Journal: Journal of Antimicrobial Chemotherapy

Article doi: dkt084

Article title: Predicting intestinal absorption of raltegravir using a population-based

ADME simulation

First Author: Darren M. Moss Corr. Author: Darren M. Moss



The corrections can be sent in any of the following ways: (i) marked on proofs electronically [we encourage you to use Adobe's editing tools (please see the next page for instructions)] or marked clearly on a copy of the proof that can be scanned and sent to <a href="mailtojac@bsac.org.uk">jac@bsac.org.uk</a>; (ii) listed clearly in an e-mail to <a href="mailtojac@bsac.org.uk">jac@bsac.org.uk</a>; or (iii) marked clearly on a copy of the proof and sent by fax to +44-(0)121-212-9822 (if sending a fax authors are advised to include a separate list of corrections as comments in the margins may be lost). If no corrections are required the JAC Editorial Office must be informed.

Changes should be kept to the minimum necessary. Changes that contradict journal style will not be made.

These proofs are for checking purposes only. They should not be considered as final publication format. The proof must not be used for any other purpose. In particular we request that you: do not post them on your personal/institutional web site, and do not print and distribute multiple copies (please use the <u>offprint order form</u>). Neither excerpts nor all of the article should be included in other publications written or edited by yourself until the final version has been published and the full citation details are available. You will be sent these when the article is published.

This is your last chance to check the article before publication. The list below aims to help you check the article systematically. You should use the accepted version of the article as a reference.

- 1. **Permissions:** Permission to reproduce any third party material in your paper should have been obtained prior to acceptance. If your paper contains figures or text that require permission to reproduce, please inform the JAC Editorial Office immediately by email.
- 2. Author names: Please check that all names have been spelled correctly and appear in the correct order. Please also check that all initials are present. Please check that the author surnames (family name) have been correctly identified by a pink background. If this is incorrect, please identify the full surname of the relevant authors. Occasionally, the distinction between surnames and forenames can be ambiguous, and this is to ensure that the authors' full surnames and forenames are tagged correctly, for accurate indexing online.
- 3. **Using the accepted version of the article you should check** that the text is complete and that all figures, tables and their legends are included.
- 4. **Figures/Tables:** Please check that they are complete, alignments are correct and that the legends/footnotes are correct. Figures in the proof are low resolution versions that will be replaced with high resolution versions when the journal is printed.
- 5. **Colour reproduction:** If your article contains Figures that we have agreed will appear in colour online but black and white in print, please check the black and white version of the Figures at the end of the article and let us know if you have any concerns.
- 6. **Special characters:** Please check that special characters, chemical structures, equations, dosages and units, if applicable, have been reproduced accurately.
- 7. **URLs:** Please check that all web addresses cited in the text, footnotes and reference list are up-to-date, and please provide a 'last accessed' date for each URL in the reference list.



# AUTHOR QUERIES - TO BE ANSWERED BY THE CORRESPONDING AUTHOR

Please respond to all queries and send any additional proof corrections. Failure to do so could result in delayed publication.

Query No.	Nature of Query	Author's Response
Q1	Would 'clinical trial data' be better than 'real-life data'?	$\bigcirc$
	Following are the source queries raised by the Journal Editor:	
SQ1	Reference 37: please provide the Abstract number.	
SQ2	Table 1 footnotes: should 'volume of distribution' be changed to 'volume of distribution at steady state'?	$\bigcirc$
SQ3	Table 4 footnotes: the information in the second sentence does not agree with the information in the table. Please state all necessary corrections.	
SQ4	Table 4 footnotes: please check the table citations very carefully. Please state all necessary corrections.	$\bigcirc$

## MAKING CORRECTIONS TO YOUR PROOF

These instructions show you how to mark changes or add notes to the document using the Adobe Acrobat Professional version 7.0 (or onwards) or Adobe Reader 8 (or onwards). To check what version you are using go to **Help** then **About**. The latest version of Adobe Reader is available for free from get.adobe.com/reader.

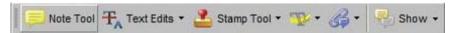
For additional help please use the **Help** function or, if you have Adobe Acrobat Professional 7.0 (or onwards), go to http://www.adobe.com/education/pdf/acrobat curriculum7/acrobat7 lesson04.pdf

### Displaying the toolbars

Adobe Reader 8: Select Tools, Comments & Markup, Show Comments and Markup Toolbar. If this option is not available, please let me know so that I can enable it for you.



Acrobat Professional 7: Select Tools, Commenting, Show Commenting Toolbar.



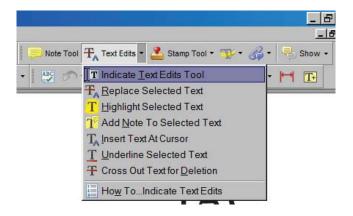
Adobe Reader 10: To edit the galley proofs, use the Comment Toolbar (Sticky Note and Highlight Text).

Ŧ



# **Using Text Edits**

This is the quickest, simplest and easiest method both to make corrections, and for your corrections to be transferred and checked.



- 1. Click Text Edits
- Select the text to be annotated or place your cursor at the insertion point.
- Click the **Text Edits** drop down arrow and select the required action.

You can also right click on selected text for a range of commenting options.

### **Pop up Notes**

With *Text Edits* and other markup, it is possible to add notes. In some cases (e.g. inserting or replacing text), a pop-up note is displayed automatically.



To **display** the pop-up note for other markup, right click on the annotation on the document and selecting **Open Pop-Up Note**.

To **move** a note, click and drag on the title area.



To **resize** of the note, click and drag on the bottom right corner.



To **close** the note, click on the cross in the top right hand corner.



To **delete** an edit, right click on it and select **Delete**. The edit and associated note will be removed.

### **SAVING COMMENTS**

In order to save your comments and notes, you need to save the file (**File, Save**) when you close the document. A full list of the comments and edits you have made can be viewed by clicking on the Comments tab in the bottom-left-hand corner of the PDF.

Figure Read by: S.S Copy Edited by: J.P

Journal of Antimicrobial Chemotherapy

65

70

75

80

85

90

95

100

J Antimicrob Chemother doi:10.1093/jac/dkt084

15

20

25

30

35

40

55

# Predicting intestinal absorption of raltegravir using a population-based ADME simulation

Darren M. Moss<sup>1\*</sup>, Marco Siccardi<sup>2</sup>, David J. Back<sup>2</sup> and Andrew Owen<sup>2</sup>

<sup>1</sup>Molecular and Biochemical Parasitology Group, Liverpool School of Tropical Medicine, Liverpool, UK; <sup>2</sup>Department of Molecular and Clinical Pharmacology, University of Liverpool, Liverpool, UK

\*Corresponding author. Tel: +44-(0)151-705-3350; Fax: +44-(0)151-794-5656; E-mail: darren.moss@liverpool.ac.uk

Received 29 October 2012; returned 13 December 2012; revised 24 January 2013; accepted 10 February 2013

**Objectives:** Raltegravir pharmacokinetics (PK) show high intra- and inter-patient variability and are also influenced by co-administered substances that alter the gastrointestinal tract environment, such as pH-altering or metal-containing agents. The aim of this investigation was to develop a population-based absorption, distribution, metabolism and excretion (ADME) model to investigate the effects of gastrointestinal pH and ingested magnesium on raltegravir PK.

**Methods:** In vitro data describing the disposition of raltegravir were obtained from literature sources or generated by standard methods. Raltegravir (400 mg single dose) PK were simulated in healthy volunteers (50 subjects per group, 20–50 years old, 0.5 proportion female subjects) over a 12 h period.

**Results:** Simulated raltegravir PK correlated well with data from clinical trials, with a mean deviation in  $C_{\text{max}}$ , AUC<sub>0-12</sub> and  $C_{\text{trough}}$  of <20%. Solubility of raltegravir in the gastrointestinal tract was increased at higher luminal pH. Increased intestinal pH and transit time both correlated with higher raltegravir absorption (P<0.001). Magnesium ingestion reduced raltegravir exposure in simulated subjects, with mean  $C_{\text{trough}}$  reduced by 32% (P<0.001).

**Conclusions:** The *in vitro-in vivo* extrapolation model developed in this study predicted raltegravir PK in virtual individuals with different gastrointestinal pH profiles. The main PK variables were predicted with good accuracy compared with reference data, and both luminal pH and magnesium were able to influence drug absorption. This modelling system provides a tool for investigating the absorption of other drugs, including HIV integrase inhibitors currently in development, which have also shown interactions with food and metal-containing products.

Keywords: HIV, pharmacokinetics, integrase inhibitors, permeability, IVIVE

#### Introduction

Raltegravir shows marked variability in pharmacokinetics (PK) both between patients and in the same patient over time. High variability in raltegravir  $C_{\rm trough}$ , with median coefficients of variation (CV%) of 128% and 245% for inter- and intra-patient variability, respectively, has been observed in a clinical cohort. Similar high variability has been observed when assessing raltegravir plasma AUC. High intra-individual variability in plasma raltegravir concentrations and inconsistent concentration–response relationships have complicated the use of therapeutic drug monitoring and predictive PK modelling of raltegravir.

The primary route of raltegravir elimination is glucuronidation via the liver, with around 30% being excreted in urine.<sup>3</sup> Metabolism is achieved by UDP-qlucuronosyltransferase 1A1 (UGT1A1)

and the drug is neither a substrate for nor an inhibitor of major cytochrome P450 enzymes.<sup>3,4</sup> Atazanavir inhibits UGT1A1, causing an increase in raltegravir exposure, and rifampicin mediates induction of UGT1A1, causing a decrease in raltegravir exposure.<sup>5,6</sup> In addition, raltegravir is a weak substrate for the drug transporters ABCB1, SLC22A6 and SLC15A1 and also inhibits SLC22A6 *in vitro*.<sup>7</sup> Raltegravir shows uneven distribution in tissues, with a 2.3-fold higher concentration in vaginal fluid and a 17-fold lower concentration in CSF compared with plasma.<sup>8,9</sup> These factors do not fully explain the high intrapatient variability in raltegravir PK, and recent clinical studies suggest that variation in raltegravir absorption may be more relevant.<sup>10</sup>

The PK of raltegravir are influenced by co-administered agents that raise gastrointestinal (GI) tract pH, such as omeprazole and

© The Author 2013. Published by Oxford University Press on behalf of the British Society for Antimicrobial Chemotherapy. All rights reserved. For Permissions, please e-mail: journals.permissions@oup.com

120

125

130

135

140

145

150

155

160

165

170

famotidine.  $^{11}$  The rate of disintegration of standard 400 mg raltegravir tablets increases when dissolved at higher pH.  $^{12}$  High-fat food has been shown to increase raltegravir exposure, which may be attributable to a fat-induced increase in raltegravir solubility, although in the same study low-fat food decreased raltegravir exposure by an unknown mechanism.  $^{13}$  Furthermore, antacids containing divalent metals have been shown to reduce raltegravir  $C_{\rm trough}$  in healthy subjects, which is believed to be the result of reduced raltegravir absorption caused by magnesium binding in the lumen.  $^{14}$  These factors may contribute to the high variability observed in raltegravir PK.

Two drug efficacy studies found that 18% (2 of 11) and 33% (2 of 6) of patients who failed raltegravir therapy had no evidence of HIV integrase mutations.  $^{15,16}$  These studies suggest that raltegravir therapy failure may occur independently of HIV resistance mutations. Raltegravir exposure may be an influencing factor; however, this was not investigated in the studies. Nonadherence to treatment may have also contributed to therapy failure without known resistance mutations. A statistically significant correlation between raltegravir exposure and virological response was recently demonstrated in patients taking 800 mg raltegravir once a day, where  $C_{\rm trough}$  was a predictor of raltegravir treatment failure. These data suggest that raltegravir exposure contributes to treatment failure, although high HIV RNA at baseline is the strongest correlate.

A population PK model has been published based on data from six fasted healthy volunteers given a single 400 mg dose of raltegravir, and another model has been published based on the data from 145 HIV-infected patients and 19 healthy volunteers. <sup>10,18</sup> The PK data used in these studies were highly variable and both concluded that the lack of information on factors with a potential

effect on raltegravir absorption limits the interpretation of the results. Our previous investigations set out to determine the impact of drug transporters, pH, cationic metals, raltegravir tablet breakdown rate and various other factors on raltegravir disposition *in vitro*. <sup>7,12,19</sup> By incorporating these mechanistic data into physiologically based PK (PBPK) models describing physiological parameters, predictions of potential interactions between raltegravir and co-administered agents may be made *a priori*.

In vitro-in vivo extrapolation (IVIVE) is the process of using in vitro physicochemical, permeability and metabolic parameters to predict in vivo drug characteristics using PK simulation modelling. The Simcyp population-based absorption, distribution, metabolism and excretion (ADME) simulator utilizes a 'bottom-up' approach that integrates demographic, anatomical, physiological and drug-specific factors.<sup>20</sup> These data are included in PBPK models, based on physiological compartments that mimic ADME processes in the human body. This approach can be used to investigate drug disposition in drug discovery and development, such as PK profiling, drug-drug interactions, bioavailability and drug exposure in special populations. <sup>21–23</sup> Moreover, design of a clinical trial can be optimized using IVIVE by determining how factors such as sample size, sex ratio and age may influence PK.<sup>24</sup> Oral absorption of a drug can be simulated using different approaches and taking into account the dynamic interplay of tablet disintegration, dissolution, solubility, intestinal permeation and, for some drugs, metabolism. The advanced dissolution, absorption and metabolism (ADAM) model within Simcyp represents the GI tract as compartments based upon their physiological and anatomical characteristics, and the relationship between permeability, metabolism and dissolution, amongst other factors, can be simulated.<sup>25</sup> The aim of this investigation was to develop an IVIVE

200

205

210

215

220

225

230

SQ2

**Table 1.** Input parameters detailing raltegravir properties

Input parameters	Value	Value Reference	
Physical chemistry			
mol. wt	445.16	42	
LogP <sub>O:W</sub>	0.58	19	
рКа	6.67	19	
B/P	0.6	43	
fu	0.17	43	
Absorption			
$P_{app}$ pH 7.4, $\times 10^{-6}$ cm/s	6.6	19	
tablet release rate	increased at higher pH	12	
solubility-pH profile	see Figure S1, available as	ACD/PhysChem Suite	
	Supplementary data at JAC Online	predicted	
stribution V <sub>ss</sub> L/kg	0.308	Simcyp predicted	
limination hepatic CL <sub>int</sub> , µL/min/10 <sup>6</sup> hepatocytes	12.4	18	
CL <sub>R</sub> , L/h	3.6	32	

 $LogP_{O:W}$ , log partition coefficient between octanol and water (pH 7); B/P, blood to plasma drug ratio; fu, free drug fraction in plasma;  $P_{app}$  pH 7.4, apical to basolateral apparent permeability in Caco-2 monolayer at pH 7.4;  $V_{SS}$ , volume of distribution;  $CL_{int}$ , intrinsic clearance;  $CL_R$ , renal clearance.

2 of 8

**JAC** 

Table 2. Values for GI pH, raltegravir solubility and raltegravir PK values in each population group after a single 400 mg dose of raltegravir

	Group 1 (very low pH)	Group 2 (low pH)	Group 3 (medium pH)	Group 4 (high pH)	Group 5 (very high pH)	
Stomach Mean pH	1	2.5	3	5	6	29
Raltegravir solubility, mg/mL	0.04	0.04	0.04	0.12	0.98	
Duodenum Mean pH Raltegravir solubility, mg/mL	4 0.04	4.5 0.08	5 0.12	6 0.98	6.4 2.4	30
Jejunum Mean pH Raltegravir solubility, mg/mL	4.4 0.07	5 0.12	5.5 0.33	6.8 5.8	7 8.9	
Ileum Mean pH Raltegravir solubility, mg/mL	6.4 2.4	6.8 5.8	7.4 20.3	7.6 30.0	8 37.3	30
Colon Mean pH Raltegravir solubility, mg/mL	5 0.12	5.5 0.33	6 0.98	6.2 1.6	6.5 2.9	31
C <sub>max</sub> , ng/mL (5th-95th percentile)	1491 (821–2836)	1980 (1376-3403)	2094 (1416-3657)	2541 (1676-4383)	2545 (1645-4148)	
AUC <sub>0-12</sub> , ng h/mL (5th–95th percentile)	5756 (2486-12335)	7285 (3770-14645)	8165 (4308-16231)	10137 (5321-18916)	11046 (6169-19691)	31!
T <sub>max</sub> , h (range)	3.7 (1.3-5.3)	3.3 (1.4-4.4)	3.2 (1.3-4.3)	2.3 (1.3-3.1)	2.1 (1.2 – 2.9)	
C <sub>trough</sub> , ng/mL (5th–95th percentile) fa	30.9 (8.4-60.9) 0.39	51.4 (21.6-93.6) 0.49	113.2 (58.4-210.1) 0.55	153.2 (87.3-254.7) 0.69	188.2 (95.4-312.8) 0.75	320
(5th-95th percentile)	(0.16-0.60)	(0.26-0.67)	(0.34-0.73)	(0.45-0.86)	(0.54-0.91)	

GI pH values are the mean for each group and had a CV% set at 2% when determining pH variation for individuals within a group. Raltegravir solubility values in each GI segment are the mean for each group and were determined using the ACD/PhysChem Suite (see Figure S1, available as Supplementary data at JAC Online). Groups 1-5 each contained 50 individuals and represented very low (Group 1) to very high (Group 5) GI pH. Geometric mean values for  $C_{max}$ , AUC<sub>0-12</sub>,  $C_{trough}$  and fa are given with 5th and 95th percentiles. Median  $T_{max}$  is given with range.

model for simulating raltegravir PK and to investigate the role of GI pH and ingested magnesium using the ADAM model. The model detailed in this investigation includes data from previous clinical trials and also *in vitro* data from previous studies. <sup>7,12,19</sup> This combined approach provides a system to determine which physiological and drug-based attributes are important in explaining high raltegravir PK variability.

predicted using the ACD/PhysChem Suite (Toronto, Canada) and data were added to the IVIVE model (Figure S1, available as Supplementary data at JAC Online). The volume of distribution and penetration into tissues were predicted with the full PBPK model within Simcyp using tissue:plasma partition coefficients predicted using the method of Poulin and Theil.<sup>26</sup> All other factors were obtained from published literature.

#### Methods

#### In vitro-in vivo extrapolation: drug parameters

Raltegravir PK properties were simulated using the full PBPK model implemented in the Simcyp population-based simulator (Version 11.0, Simcyp Ltd, UK). Data describing raltegravir physiochemical and pharmacological properties were added to the model and are summarized in Table 1.

Raltegravir logP, pKa and apparent permeability ( $P_{\rm app}$ ) have previously been determined in-house. <sup>19</sup> The dissolution rate of standard 400 mg film-coated raltegravir tablets was previously determined in a physiological range of pH buffers, and a higher dissolution rate was observed with increased pH. <sup>12</sup> The intrinsic solubility of raltegravir at pH 1–9 was

#### In vitro-in vivo extrapolation: system parameters

All system parameters were taken from the North European Caucasian population library within Simcyp, with the exception of patient GI pH values, the ranges of which were obtained from the literature. <sup>27–30</sup> All simulations were performed in five separate population groups, named Group 1, 2, 3, 4 and 5. These groups contained individuals with the same physiological attributes as each other (allowing for variation around the mean) except for mean GI pH values, which increased from Group 1 to Group 5 (Table 2). All simulated groups consisted of 50% female subjects. The mean transit times of drug in the different sections of the GI tract were 0.4, 3.3 and 12 h in the stomach, intestine and colon, respectively. Each transit time is the mean for a population representative

325

330

335

345

290

270

275

280

350

355

360

370

375

380

385

395

400

405

and each segment had a CV% of 30% when determining GI transit time for individuals in a group.

#### In vitro-in vivo extrapolation: simulation design

Raltegravir (400 mg single dose) PK were simulated in healthy volunteers (20–50 years old, 0.5 proportion female subjects) over a 12 h period. Fifty subjects per population group were chosen as the number to use in all subsequent simulations. Simulated PK results were compared with actual PK results in published data and were used to calculate simulated geometric mean values for  $C_{\rm max}$  (ng/mL), AUC<sub>0-12</sub> (ng h/mL),  $C_{\rm trough}$  (ng/mL) and fa (fraction of dose absorbed), which were given with 5th and 95th percentiles. Median  $T_{\rm max}$  (h) was calculated and given with range. The transit time of raltegravir through the small intestine was collected from all simulated subjects and any correlation between transit time and fraction of raltegravir absorbed was determined.

A parallel trial was performed as described above to simulate the impact of elevated GI magnesium concentrations on raltegravir PK values. Drug parameters were altered so that the Caco-2 monolayer  $P_{\rm app}$  of raltegravir was reduced from 6.6 to 3.4 cm/s, which was taken from previously published data, simulating the impact of 25 mM magnesium salt on raltegravir  $P_{\rm app}.^{19}$ 

#### Statistical analyses

Data were analysed using SPSS 19.0 for Windows. All data were tested for normality using the Shapiro–Wilk test. An independent t-test was used to determine the significance of normally distributed data and the Mann–Whitney U-test was used for non-normal data. A two-tailed P value of <0.05 was accepted as being statistically significant. The correlation between simulated raltegravir intestinal transit time (h) and fraction of drug absorbed was assessed using linear regression analysis. The regression coefficient ( $\beta$ ) was calculated from the linear regression plot and given with 95% CI.

#### **Results**

#### Impact of GI pH on raltegravir solubility

The impact of pH on raltegravir solubility in the GI tract is shown in Table 2. A general trend towards increased raltegravir solubility in all GI segments can be observed by progressing from Group 1 to 5. Between groups the subjects differ only in mean GI pH values (subjects in Group 1 have very low GI pH, progressing to Group 5 subjects with very high GI pH); therefore the increase in raltegravir solubility is likely to be a direct effect of increased GI pH.

# Impact of GI pH and drug intestinal transit time on raltegravir PK parameters

The impact of GI pH on raltegravir PK parameters in subject Groups 1 to 5 is also shown in Table 2. Using linear regression analysis, a general trend towards increased raltegravir exposure can be observed by progressing from Group 1 (very low GI pH) to Group 5 (very high GI pH) ( $\beta=0.088$  for each group progression (95% CI=0.077-0.099,  $R^2=0.496$ , P<0.001). Median  $T_{\rm max}$  decreased as GI pH increased, which may be influenced by decreased tablet disintegration time and higher drug solubility during the earlier stages of the GI tract. Raltegravir intestinal transit time data were collated from subjects in all groups tested (50 subjects from each group, 250 subjects in total) and

ranged from 68 to 350 min. Linear regression analysis using intestinal transit time (independent variable) and fraction of drug absorbed (dependent variable) gave a  $\beta$ -value of 0.099 (95% CI=0.085-0.113,  $R^2$ =0.427), which equates to a 9.9% increase in fraction of drug absorbed for every 1 h increase in intestinal transit time (P<0.001).

The fraction of raltegravir absorbed from each GI segment is shown for subjects in Groups 1, 2, 3, 4 and 5 (Figure 1). Raltegravir absorption via the jejunum was low in subjects from Groups 1, 2 and 3 but became a more substantial route of absorption in subjects from Groups 4 and 5. Similarly, raltegravir absorption via the colon was greater in subjects from Groups 4 and 5. Group 5 subjects showed a mean drug fraction absorbed via the colon of 20% compared with small intestine. A fraction absorbed of 28% was observed for the hydrophilic drug cefradine in a separate study using rats.<sup>31</sup> As subjects differ only in their mean GI pH values between groups, the differences in absorption profiles between groups are likely to be pH related.

425

450

# Comparison between reference and simulated raltegravir PK values

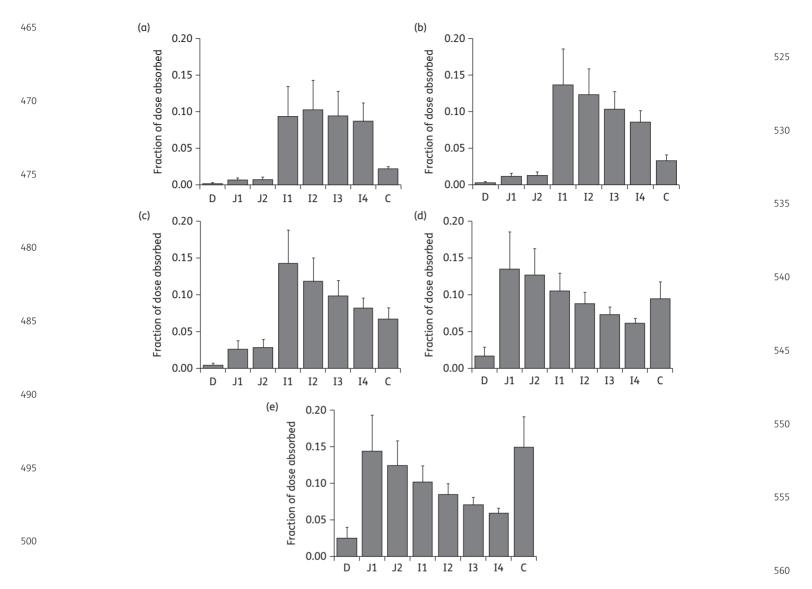
Reference PK values (mean  $\pm$  SD) for  $C_{\text{max}}$  (2519  $\pm$  1930 ng/mL),  $AUC_{0-12}$  (7076 ± 4071 ng h/mL) and  $C_{trough}$  (71 ± 50 ng/mL) in Table 3 are the mean of geometric mean values for fasted, healthy volunteers in selected previous studies. 3,11,13,14,18,32 All reference values displayed high inter-study variability, as shown by the large standard deviations. Simulated PK values obtained from subjects in Groups 1 to 5 fell within the range obtained for reference PK values. Group 2 displayed the best match with the reference value for  $AUC_{0-12}$  (+3% deviation) and  $C_{trough}$ (-28% deviation), although the reference  $C_{\text{max}}$  value was most closely predicted in Group 4 (+1% deviation). Groups 2 and 4 represent populations with 'low' and 'high' GI pH values, respectively (Table 2). When simulated data from all groups were combined, reference data matched well with simulated geometric mean  $C_{max}$  (2091+801 ng/mL; -17% deviation), AUC<sub>0-12</sub>  $(8255 \pm 4214 \text{ ng h/mL}; 17\% \text{ deviation})$  and  $C_{trough}$   $(77 \pm 79 \text{ ng/s})$ mL; 8% deviation).

#### Impact of magnesium on raltegravir exposure

The impact of magnesium binding on raltegravir PK values in subject Groups 1 to 5 is shown in Table 4. Geometric mean ratios (GMRs) were generated that were the ratios of raltegravir PK values in the presence of elevated GI magnesium concentrations and PK values under the standard conditions given in Table 2. Simulated  $C_{\rm max}$ , AUC and  $C_{\rm trough}$  were all significantly (P<0.008) reduced by the presence of magnesium in the GI tract. Raltegravir  $T_{\rm max}$  was not significantly altered by magnesium in Groups 1 (P=0.49), 2 (P=0.29) or 3 (P=0.34) but was significantly increased in Groups 4 (2.6 h versus 2.3 h, P=0.003) and 5 (2.5 h versus 2.1 h, P=0.001). The increased  $T_{\rm max}$  in Groups 4 and 5 are likely a result of an inhibited rate of raltegravir absorption, caused by reduced tablet breakdown and lower drug solubility. Data from all groups were combined and are shown in Figure 2.

4 of 8

**JAC** 



**Figure 1.** Mean simulated fraction of raltegravir absorbed (drug absorbed in segment/drug administered) ± SD in each GI segment from 50 individuals from Group 1 (a; very low pH), Group 2 (b; low pH), Group 3 (c; medium pH), Group 4 (d; high pH) and Group 5 (e; very high pH). Table 2 gives details of GI pH values for each group. D, duodenum; J1, jejunum section 1; J2, jejunum section 2; I1, ileum section 1; I2, ileum section 2; I3, ileum section 2; I3, ileum section 2; I4,

Table 3. Comparison of reference raltegravir PK values obtained from the literature with simulated PK values

	C <sub>max</sub> (ng/mL)	Difference from reference (%)	AUC <sub>0-12</sub> (ng h/mL)	Difference from reference (%)	C <sub>trough</sub> (ng/mL)	Difference from reference (%)
Reference value	2519±1930	_	7076±4071	_	71 ± 50	_
Group 1	$1491 \pm 582$	-41%	$5756 \pm 3092$	-19%	$31 \pm 27$	-56%
Group 2	$1980 \pm 623$	-21%	$7285 \pm 3399$	3%	$51 \pm 31$	-28%
Group 3	$2094 \pm 665$	-17%	$8165 \pm 3618$	15%	$113 \pm 48$	59%
Group 4	$2541 \pm 808$	1%	$10137 \pm 4300$	43%	$153 \pm 60$	115%
Group 5	$2545 \pm 792$	1%	$11046 \pm 4335$	56%	$188 \pm 78$	164%
Combined data	2091 + 801	-17%	8255 + 4214	17%	77 + 79	8%

Reference PK values for  $C_{\text{max}}$ ,  $\text{AUC}_{0-12}$  and  $C_{\text{trough}}$  are the mean  $\pm$  SD values of geometric mean values for fasted, healthy volunteers in selected previous studies.  $^{3,11,13,14,18,32}$  Each simulated population group contained 50 individuals and geometric mean  $\pm$  SD values are presented for  $C_{\text{max}}$ ,  $\text{AUC}_{0-12}$  and  $C_{\text{trough}}$ . The percentage difference of each simulated result to the reference value is given.

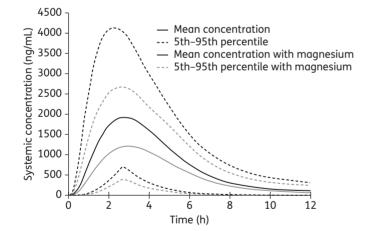
ileum section 4; C, colon.

505

Table 4. Raltegravir PK values in the presence of elevated magnesium GI concentrations

	Group 1 (very low pH)	Group 2 (low pH)	Group 3 (medium pH)	Group 4 (high pH)	Group 5 (very high pH)	
C <sub>max</sub> , ng/mL	842	1215	1301	1604	1616	
5th-95th percentile	447-1611	831-2114	858-2331	1014-2842	1028-2672	
GMR	0.56	0.61	0.62	0.63	0.63	64
P value	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	
AUC <sub>0-12</sub> , ng h/mL	3435.7	4635	5208	6712	7359	
<del>Range</del>	1394-7542	2215-9767	2623-10698	3306-13023	3769-13570	
GMR	0.60	0.64	0.64	0.61	0.67	
P value	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	650
T <sub>max</sub> , h	3.8	3.5	3.3	2.6	2.5	
Range	1.4-5.7	1.4-4.7	1.4-4.5	1.3-3.5	1.3-3.4	
Median ratio	1.03	1.06	1.03	1.13	1.19	
P value	0.49	0.29	0.34	0.003	0.001	655
C <sub>trough</sub> , ng/mL	21.9	33.3	66.2	90.3	152.1	
5th-95th percentile	4.6-58.2	11.8-63.2	32.5-119.8	48.4-153.5	88.9-253.6	
GMR	0.71	0.65	0.58	0.59	0.81	
P value	0.008	< 0.001	< 0.001	< 0.001	0.005	
fa	0.23	0.31	0.35	0.45	0.50	660
5th-95th percentile	0.09-0.38	0.15-0.46	0.20-0.50	0.27-0.64	0.31-0.67	
GMR	0.59	0.63	0.64	0.65	0.67	
P value	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	

Each simulated population group contained 50 individuals. Geometric mean values for  $C_{\text{max}}$ ,  $AUC_{0-12}$ ,  $C_{\text{trough}}$  and fa are given with 5th and 95th percentiles. Median  $T_{\text{max}}$  (h) is given with range. Values in Table  $\frac{6}{4}$  have been divided by corresponding values in Table  $\frac{4}{4}$  to calculate the GMRs ( $C_{\text{max}}$ ,  $AUC_{0-12}$ ,  $C_{\text{trough}}$  and fa) or median ratios ( $T_{\text{max}}$ ). P-values were calculated by comparing values between Table 4 and Table 2.



**Figure 2.** Mean simulated raltegravir plasma concentration—time profile (black continuous line) with 5th and 95th percentiles (black broken lines) using combined data from Groups 1–5 (50 subjects from each group, 250 subjects in total). Also shown is the mean simulated raltegravir plasma concentration—time profile (grey continuous line) with 5th and 95th percentiles (grey broken lines) using combined data from Groups 1–5 following co-ingestion of magnesium (50 subjects from each group, 250 subjects in total).

#### **Discussion**

The IVIVE model developed in this study predicted the PK of raltegravir in individuals with different GI pH profiles. The main PK

variables were predicted with good accuracy when compared with reference data. In simulated subjects, increased GI pH and small intestinal transit time were both associated with improved raltegravir exposure. Importantly, both factors are capable of causing intra-patient PK variability *in vivo* under certain conditions.<sup>33,34</sup>

SQ3

SQ4

675

HIV-infected patients, particularly those with advanced disease progression, have both higher gastric pH and higher prevalence of gastric microbe colonization than in uninfected individuals.<sup>35,36</sup> The higher gastric pH in HIV-infected patients may explain why co-administration of omeprazole with raltegravir shows less impact on raltegravir PK in HIV-infected patients (40% increase in AUC) compared with healthy volunteers (210% increase in AUC). 11,37 However, this hypothesis needs empirical confirmation as GI pH was not directly measured in either study and there are no published data showing the effect of pH on raltegravir solubility in vivo. A recent study found a statistically significant 65% increase (P=0.01) in raltegravir bioavailability in females compared with males. 10 Females have higher gastric pH and longer GI transit time than males and the results from our simulations suggest both of these factors could potentially increase raltegravir exposure.<sup>20</sup> However, conflicting reports suggest no increased raltegravir exposure in females and factors other than GI pH, such as lower hepatic blood flow and reduced intestinal P-glycoprotein expression in females, may better explain any difference in bioavailability. 16,32

The simulations showed evidence of decreased raltegravir exposure when individuals were exposed to magnesium in the GI

6 of 8

610

615

620

630

Effect of pH and metals on simulated raltegravir PK

JAC

tract. Previously determined permeability values were used in the simulations, where 25 mM magnesium chloride significantly reduced raltegravir  $P_{app}$  through Caco-2 monolayers. <sup>19</sup> It must be acknowledged that the concentration of GI magnesium in real subjects would fluctuate depending on compartment volume and magnesium absorption, therefore the intestinal permeation of raltegravir would also fluctuate. However, it can be seen from the model that the rate of raltegravir absorption is a limiting factor in ultimate exposure. The data from these simulations support the hypothesis that the combined effect of increased solubility and reduced intestinal permeability could explain the interaction observed *in vivo*, where raltegravir  $T_{max}$  and  $C_{trough}$  were lower in subjects co-administered raltegravir and a magnesium-containing antacid. <sup>14</sup>

700

705

710

715

735

740

750

The model created in this study has limitations and the inclusion of further in vitro data could potentially improve the model's predictive ability. Elimination of raltegravir from virtual subjects was determined by whole organ clearance obtained from published literature: information about the role of individual metabolic enzymes was not included. 18,32 The metabolism of raltegravir involves enzymes UGT1A1, UGT1A3 and UGT1A9.4 A model that included the clearance of raltegravir by these individual enzymes, combined with known phenotypic variations and expression levels in the virtual population, could provide a more accurate model for determining raltegravir elimination. Similarly, the impact of drug transporters on raltegravir exposure in vivo is not fully understood and was not included in the final model. The simulated population in our model used physiological parameters based on data from healthy adults, and the model at present does not contain a population representing HIV-infected individuals. Consequently, we could not investigate the influence of the disease on raltegravir exposure in our model. Most raltegravir PK data predicted by our model were within the range of those found in clinical data; however, the volume of raltegravir distribution is substantially lower than in previously published models. 10,18 This discrepancy may be due to the differences in modelling strategy, where our model utilized in vitro data in PBPK simulations (bottom-up) and the previously published models used patient PK data (top-down).

A relationship between raltegravir exposure and treatment outcome has yet to be conclusively established in the clinic. There is evidence from studies that raltegravir exposure, when combined with virological parameters, may play a role in treatment success and drug resistance development. 15,17,38 Raltearavir PK profiles occasionally contain multiple peaks or delayed (>8 h after dosing) peaks and, in an attempt to explain these peaks, it has been suggested that raltegravir is able to undergo enterohepatic recirculation via conversion of the raltegravir glucuronide metabolite back to the parent form in the intestine, leading to subsequent reabsorption. 10 However, a study in healthy subjects using radiolabelled raltegravir did not support the theory, as there was no evidence of drug reabsorption. 15 Enterohepatic recirculation of raltegravir was therefore not included in the final model. Tablet disintegration rate, which has been shown to increase in higher pH solutions in vitro, may help explain unusual raltegravir PK profiles. 12,39 Also, preferential absorption of raltegravir from particular sections of the GI tract depending on GI pH, as demonstrated in our model (Figure 1), may further explain delayed/multiple drug concentration peaks. These data remain theoretical as no study has investigated raltegravir absorption via different sections of the gut *in vivo*. Further investigation is needed to determine the extent of raltegravir enterohepatic recirculation and these data would have the potential to further improve our model.

In conclusion, the Simcyp population-based ADME simulator has been used to create a model simulating raltegravir PK parameters with acceptable accuracy to real-life data. The most useful feature of this model is to investigate 'what-if' scenarios by directly altering subject physiological parameters such as GI pH and drug absorption rate and observing the effect on raltegravir PK. This also provides a tool for investigating the absorption of other drugs, such as the second-generation HIV integrase inhibitors elvitegravir and dolutegravir, which have also shown interactions with food and metal-containing products. <sup>40,41</sup>

#### Funding

This study was supported by internal funding.

#### **Transparency declarations**

D. J. B. has received consultancy, honoraria and grant income from Merck. All other authors: none to declare.

#### Supplementary data

Figure S1 is available as Supplementary data at *JAC* Online (http://jac.oxfordjournals.org/).

#### References

- **1** Siccardi M, D'Avolio A, Rodriguez-Novoa S *et al.* Intrapatient and interpatient pharmacokinetic variability of raltegravir in the clinical setting. *Ther Drug Monit* 2012; **34**: 232–5.
- **2** Cattaneo D, Gervasoni C, Meraviglia P *et al.* Inter- and intra-patient variability of raltegravir pharmacokinetics in HIV-1-infected subjects. *J Antimicrob Chemother* 2012; **67**: 460–4.
- **3** Kassahun K, McIntosh I, Cui D *et al.* Metabolism and disposition in humans of raltegravir (MK-0518), an anti-AIDS drug targeting the human immunodeficiency virus 1 integrase enzyme. *Drug Metab Dispos* 2007; **35**: 1657–63.
- **4** Iwamoto M, Kassahun K, Troyer MD *et al.* Lack of a pharmacokinetic effect of raltegravir on midazolam: in vitro/in vivo correlation. *J Clin Pharmacol* 2008; **48**: 209–14.
- **5** Cattaneo D, Ripamonti D, Baldelli S *et al.* Exposure-related effects of atazanavir on the pharmacokinetics of raltegravir in HIV-1-infected patients. *Ther Drug Monit* 2010; **32**: 782–6.
- **6** Wenning LA, Hanley WD, Brainard DM *et al.* Effect of rifampin, a potent inducer of drug-metabolizing enzymes, on the pharmacokinetics of raltegravir. *Antimicrob Agents Chemother* 2009; **53**: 2852–6.
- **7** Moss DM, Kwan WS, Liptrott NJ *et al.* Raltegravir is a substrate for SLC22A6: a putative mechanism for the interaction between raltegravir and tenofovir. *Antimicrob Agents Chemother* 2011; **55**: 879–87.
- **8** Croteau D, Letendre S, Best BM *et al*. Total raltegravir concentrations in CSF exceed the 50-percent inhibitory concentration for wild-type HIV-1. *Antimicrob Agents Chemother* 2010; **54**: 5156–60.
- ${f 9}$  Clavel C, Peytavin G, Tubiana R et al. Raltegravir concentrations in the genital tract of HIV-1-infected women treated with a

780

770

785

795

- raltegravir-containing regimen (DIVA 01 study). *Antimicrob Agents Chemother* 2011; **55**: 3018–21.
- **10** Arab-Alameddine M, Fayet-Mello A, Lubomirov R et al. Population pharmacokinetic analysis and pharmacogenetics of raltegravir in HIV-positive and healthy individuals. *Antimicrob Agents Chemother* 2012: **56**: 2959–66.
  - Iwamoto M, Wenning LA, Nguyen BY *et al.* Effects of omeprazole on plasma levels of raltegravir. *Clin Infect Dis* 2009; **48**: 489–92.
  - Moss DM, Siccardi M, Khoo SH *et al.* The interplay between raltegravir solubility, tablet dissolution, metal binding, charge state and cell permeability. In: *Abstracts of the Thirteenth International Workshop on Clinical Pharmacology of HIV Therapy, Barcelona, Spain, 2012.* Abstract P-27. Virology Education, Utrecht, Netherlands.
  - Brainard DM, Friedman EJ, Jin B et al. Effect of low-, moderate-, and high-fat meals on raltegravir pharmacokinetics. *J Clin Pharmacol* 2011; **51**: 422–7.
- 14 Kiser JJ, Bumpass JB, Meditz AL *et al.* Effect of antacids on the pharmacokinetics of raltegravir in human immunodeficiency virus-seronegative volunteers. *Antimicrob Agents Chemother* 2010; **54**: 4999–5003.
  - da Silva D, Van Wesenbeeck L, Breilh D *et al.* HIV-1 resistance patterns to integrase inhibitors in antiretroviral-experienced patients with virological failure on raltegravir-containing regimens. *J Antimicrob Chemother* 2010; **65**: 1262–9.
  - Kozal MJ, Lupo S, DeJesus E *et al.* A nucleoside- and ritonavir-sparing regimen containing atazanavir plus raltegravir in antiretroviral treatment-naive HIV-infected patients: SPARTAN study results. *HIV Clin Trials* 2012; **13**: 119–30.
  - **17** Rizk ML, Hang Y, Luo WL *et al.* Pharmacokinetics and pharmacodynamics of once-daily versus twice-daily raltegravir in treatment-naive HIV-infected patients. *Antimicrob Agents Chemother* 2012; **56**: 3101–6.
  - Wang L, Soon GH, Seng KY *et al*. Pharmacokinetic modeling of plasma and intracellular concentrations of raltegravir in healthy volunteers. *Antimicrob Agents Chemother* 2011; **55**: 4090–5.
  - Moss DM, Siccardi M, Murphy M *et al.* Divalent metals and pH alter raltegravir disposition in vitro. *Antimicrob Agents Chemother* 2012; **56**: 3020–6.
  - Soldin OP, Chung SH, Mattison DR. Sex differences in drug disposition. *J Biomed Biotechnol* 2011; **2011**: 187103.
  - Perloff MD, von Moltke LL, Greenblatt DJ. Fexofenadine transport in Caco-2 cells: inhibition with verapamil and ritonavir. *J Clin Pharmacol* 2002; **42**: 1269–74.
  - Storch CH, Theile D, Lindenmaier H et al. Comparison of the inhibitory activity of anti-HIV drugs on P-glycoprotein. *Biochem Pharmacol* 2007; **73**: 1573–81.
  - Sharma M, Saravolatz LD. Rilpivirine: a new non-nucleoside reverse transcriptase inhibitor. *J Antimicrob Chemother* 2013; **68**: 250–6.
  - Azijn H, Tirry I, Vingerhoets J *et al.* TMC278, a next-generation nonnucleoside reverse transcriptase inhibitor (NNRTI), active against wild-type and NNRTI-resistant HIV-1. *Antimicrob Agents Chemother* 2010; **54**: 718–27.
- Darwich AS, Neuhoff S, Jamei M et al. Interplay of metabolism and transport in determining oral drug absorption and gut wall metabolism: a simulation assessment using the 'Advanced Dissolution, Absorption, Metabolism (ADAM)' model. *Curr Drug Metab* 2010; **11**: 716–29.

- Poulin P, Theil FP. Prediction of pharmacokinetics prior to in vivo studies. 1. Mechanism-based prediction of volume of distribution. *J Pharm Sci* 2002; **91**: 129–56.
- Jamei M, Marciniak S, Feng K *et al.* The Simcyp population-based ADME simulator. *Expert Opin Drug Metab Toxicol* 2009; **5**: 211–23.
- Charman WN, Porter CJ, Mithani S *et al.* Physiochemical and physiological mechanisms for the effects of food on drug absorption: the role of lipids and pH. *J Pharm Sci* 1997; **86**: 269–82.
- McCloy RF, Greenberg GR, Baron JH. Duodenal pH in health and duodenal ulcer disease: effect of a meal, Coca-Cola, smoking, and cimetidine. *Gut* 1984; **25**: 386–92.

- Rune SJ, Viskum K. Duodenal pH values in normal controls and in patients with duodenal ulcer. *Gut* 1969; **10**: 569–71.
- Yuasa H, Soga N, Kimura Y *et al.* Effect of aging on the intestinal transport of hydrophilic drugs in the rat small intestine. *Biol Pharm Bull* 1997; **20**: 1188–92.
- Iwamoto M, Wenning LA, Petry AS *et al.* Safety, tolerability, and pharmacokinetics of raltegravir after single and multiple doses in healthy subjects. *Clin Pharmacol Ther* 2008; **83**: 293–9.
- Ibekwe VC, Fadda HM, McConnell EL *et al.* Interplay between intestinal pH, transit time and feed status on the in vivo performance of pH responsive ileo-colonic release systems. *Pharm Res* 2008; **25**: 1828–35.
- McConnell EL, Fadda HM, Basit AW. Gut instincts: explorations in intestinal physiology and drug delivery. *Int J Pharm* 2008; **364**: 213–26.
- **35** Shelton MJ, Akbari B, Hewitt RG et al. Eradication of Helicobacter pylori is associated with increased exposure to delavirdine in hypochlorhydric HIV-positive patients. J Acquir Immune Defic Syndr 2000; **24**: 79–82.
- **36** Chave JP, Thorens J, Frohlich F *et al.* Gastric and duodenal bacterial colonization in HIV-infected patients without gastrointestinal symptoms. *Am J Gastroenterol* 1994; **89**: 2168–71.
- Rhame F, Matson M, Wood D *et al.* Effects of famotidine and omeprazole on raltegravir pharmacokinetics in HIV-infected individuals. In: *Abstracts of the Twelfth European AIDS Conference, Cologne, Germany, 2009.*
- Cohen CJ, Andrade-Villanueva J, Clotet B *et al.* Rilpivirine versus efavirenz with two background nucleoside or nucleotide reverse transcriptase inhibitors in treatment-naive adults infected with HIV-1 (THRIVE): a phase 3, randomised, non-inferiority trial. *Lancet* 2011; **378**: 229–37
- Molina JM, Cahn P, Grinsztejn B *et al.* Rilpivirine versus efavirenz with tenofovir and emtricitabine in treatment-naive adults infected with HIV-1 (ECHO): a phase 3 randomised double-blind active-controlled trial. *Lancet* 2011; **378**: 238–46.
- van Lunzen J, Maggiolo F, Arribas JR *et al.* Once daily dolutegravir (S/GSK1349572) in combination therapy in antiretroviral-naive adults with HIV: planned interim 48 week results from SPRING-1, a doseranging, randomised, phase 2b trial. *Lancet Infect Dis* 2012; **12**: 111–8.
- Shimura K, Kodama EN. Elvitegravir: a new HIV integrase inhibitor. Antivir Chem Chemother 2009; **20**: 79–85.
- Summa V, Petrocchi A, Bonelli F *et al.* Discovery of raltegravir, a potent, selective orally bioavailable HIV-integrase inhibitor for the treatment of HIV-AIDS infection. *J Med Chem* 2008; **51**: 5843–55.
- **43** Laufer R, Paz OG, Di Marco A *et al.* Quantitative prediction of human clearance guiding the development of raltegravir (MK-0518, Isentress) and related HIV integrase inhibitors. *Drug Metab Dispos* 2009; **37**: 873–83.

8 of 8