<u>Abstract</u>

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- 2 Plasmodium falciparum is a protozoan parasite that causes the most severe form of human
- 3 malaria. Four other *Plasmodium* species can also infect humans *P. vivax*, *P. ovale*, *P.*
- 4 malariae and P. knowlesi but P. falciparum is the most prevalent Plasmodium species in
- 5 the African region, where 90% of all malaria occurs, and it is this species that causes the
- 6 great majority of malaria deaths. These were reported at 438,000 in 2015 (1) from an
- 7 estimated 214 million cases; importantly, however, figures for the global burden of malaria
- 8 tend to have wide margins of error due to poor and inaccurate reporting (2-4). In this
- 9 Perspective, the unique features of the *P. falciparum* parasite are highlighted and current
- issues surrounding the control and treatment of this major human pathogen are discussed.

What is special about Plasmodium falciparum?

All *Plasmodium* parasites share unique and fascinating biological features, enabling them to invade, colonise, replicate and persist in diverse host environments. They have a complex and highly-evolved lifecycle that requires both an insect vector and a vertebrate host (fig 1A), and their cell and molecular biology is highly unusual. *Plasmodium* belongs to an early-diverging lineage of eukaryotes, the Apicomplexan phylum, which evolved from a free-living algal ancestor into an obligate intracellular parasite (5): the resultant cells carry a relic plastid (6) as well as other special organelles that facilitate invasion of host cells (7) (fig 1B). Inside these cells they grow via various syncytial modes of cell division rather than conventional binary fission (8). *Plasmodium* parasites are widespread throughout the animal kingdom but tend to be highly specialised for particular hosts – avian, reptilian, mammalian, etc.: *P. falciparum* infects only humans and great apes (9, 10), and although its basic biology shares all the above characteristics, it also has additional features that can cause unique and severe pathology in humans. The case fatality rate of falciparum malaria is ~0.3-0.45%, but in a subset of severe malaria cases it can exceed 20% (11) (table 1).

Table 1: Manifestations of severe falciparum malaria

| Clinical manifestation | Frequency | | Prognostic of poor outcome? | | Linked laboratory indices |
|---|-----------|--------|-----------------------------|--------|--|
| | Children | Adults | Children | Adults | |
| Cerebral coma or impaired consciousness | +++ | ++ | +++ | +++ | Parasite sequestration in brain |
| Repeated convulsions | +++ | + | + | ++ | Parasite sequestration in brain, Hypoglycaemia |
| Prostration | +++ | +++ | + | + | |
| Respiratory distress | +++ | ++ | +++ | +++ | Metabolic acidosis/Hyperlactataemia, |
| | | | | | Severe anaemia |
| Pregnancy malaria | - | +++ | - | +++ | Parasite sequestration in placenta, Hypoglycaemia |

Table shows a non-exhaustive list of key disease features in severe falciparum malaria.

Adapted from data in references (4) and (11).

Firstly, in the human-pathogenic stage of its lifecycle, which consists of growth inside erythrocytes (fig 1A), *P. falciparum* can infect erythrocytes of all ages. This distinguishes it from the other major human malaria species, *P. vivax*, which is restricted to rare immature erythrocytes called reticulocytes. Accordingly, whereas *P. vivax* growth is limited by the scarcity of reticulocytes, *P. falciparum* can swiftly reach parasitaemias of 10-20%, and each infected cell can produce ~20-30 new parasites every 48 hours. This capacity to reach very high parasitaemias exacerbates malaria pathologies such as severe anaemia, metabolic acidosis and respiratory distress (11, 12) (table 1).

Secondly, *P. falciparum* has a family of major virulence genes called *var* genes which are unique to this parasite and its close relatives (ape malaria parasites in the '*Laverania*' subgenus (13)). These virulence factors play key roles in other lethal pathologies, such as cerebral malaria and pregnancy malaria (14) (table 1). *Var* genes encode an adhesive protein, 'PfEMP1', which is exported and expressed on the surface of infected erythrocytes, allowing the cells to be sequestered in capillaries as they mature. Thus they avoid clearance by the spleen, but sequestered cells obstruct blood flow and cause inflammatory responses that are particularly harmful in vessels of the brain or placenta (15). Most other malaria parasites do not adhere in this way and do not cause cerebral comas or pregnancy complications. Furthermore, the *var* gene family is variantly expressed, giving rise to antigenic variation in PfEMP1, and hence immune evasion (16), which is one reason why sterile immunity to falciparum malaria is rarely achieved in humans.

A third unusual feature of *P. falciparum* is its extremely biased genome, the implications of which are not yet understood. At 81% A/T, this is one of the most biased genomes ever sequenced (17). Not all human malaria parasites share this bias – the *P. vivax* genome is only ~58% A/T – and although elegant studies have recently elucidated *how* the bias is maintained at a molecular level (18), they have not established *why*. It may be that A/T-rich DNA favours permissive transcription (19) or rapid DNA replication (20, 21) – both signature features of *P. falciparum*. Certainly, this genome bias presents a severe challenge to biologists in sequencing, cloning, expressing and working with *P. falciparum* genes.

Current issues in P. falciparum biology

 Hot topics in *P. falciparum* biology range from the molecular to the epidemiological. On the molecular and cellular level, the parasite has unusual basic biology that is both academically interesting and medically important. Understanding the parasite's unique features will help scientists to focus on new targets for antimalarial drugs and vaccines. These features include the unusual genome mentioned above; the unusual cell cycles (8)(fig 1A); the biochemical specialisations for an intracellular lifecycle (22); and the invasion pathways (7),

transport pathways (23) and capacities for host-cell remodelling (24, 25) that have evolved to facilitate life inside anucleate erythrocytes. Interestingly, a recent screen for genes that are essential for growth in the rodent malaria parasite *P. berghei* revealed an unprecedentedly high proportion of indispensable genes, which was extrapolated to be similarly true for *P. falciparum* (26). This raises conceptual questions about reductive specialisation for an intracellular parasitic lifestyle and importantly it also raises the prospect of an abundance of genetically-essential targets for antimalarial drugs.

Moving to the epidemiological level, falciparum malaria has historically been a major scourge of humans throughout the tropics and subtropics, and attempts to control the disease have a long and complex history. Excitingly and controversially, the prospect of global malaria eradication has recently returned to the fore (27), after the failure of the first in 'WHO Global Malaria Eradication Programme' in the mid-20th century. The original programme was built upon considerable success in eliminating the disease from Europe and North America in the early 1900s via large-scale environmental insecticide treatment which targeted the mosquito vector, together with drug treatment of malaria cases in humans. Reasons for its ultimate failure included the development of mosquito resistance to the insecticide DDT and parasite resistance to the antimalarial chloroquine, as well as the more challenging dynamics of disease transmission in hyperendemic tropical areas. Figure 1C illustrates the subsequent rebound in the global burden of malaria. Encouragingly, however, more modern interventions have reduced this burden once again, with a drop of ~40% in malaria in Africa over the past 15 years, attributed primarily to the use of insecticide-treated bednets (28, 29).

Whether or not malaria can actually be eradicated with current tools remains a topic of debate (30, 31), and some of these tools are now increasingly threatened, as discussed below. There is a clear historical trend for disease resurgence when control measures fail or when funding to sustain them fails (not only malaria (fig 1C) but other parasitic diseases such as sleeping sickness have illustrated this (32)). Nevertheless, striking successes have already been achieved in eliminating malaria from island nations such as Sri Lanka (33), as well as non-island nations on the margins of transmission zones, such as Morocco and Kyrgyzstan. Others including China and Malaysia, benefitting from regional cross-border collaboration, are close to the elimination goal (34).

Current challenges in *P. falciparum* biology

Key challenges in the *P. falciparum* field range from basic science to real-world intervention. At the level of basic science, the unusual biology of this parasite makes it challenging to work with, although it remains one of only two human malaria parasites that can be grown in laboratory culture at all (35) (the other is the zoonotic macaque parasite *P. knowlesi* (36)).

The ability to culture *P. falciparum* in human erythrocytes makes genetic experiments feasible (37), albeit painfully inefficient when compared to model systems. Nevertheless, in the past decade great advances have been made in developing genetic tools for gene tagging, gene knockouts, knockdowns, inducible approaches and gene editing (38), and the scope for further improvement remains substantial. Collaborative efforts to sequence hundreds of *P. falciparum* strains from around the world are fast revealing the genetic diversity of the species (39), but challenges remain in efficiently adapting field strains to *in vitro* culture (40) and there are persistent concerns about the relevance of laboratory experiments conducted exclusively in strains that have been in culture for decades.

Meanwhile, major challenges persist for *P. falciparum* control in the real world. Drug resistant parasites (as well as insecticide-resistant mosquito vectors) are a recurring problem, as are sufficiently accurate and sensitive diagnostics, while the gold-standard disease-prevention tool of a vaccine against *P. falciparum* remains elusive despite decades of scientific effort.

P. falciparum parasites have historically developed resistance to every antimalarial deployed (fig 1C). Current first-line treatments are all based on artemisinin derivatives, which are highly effective but very short-lived in the bloodstream. Therefore they are always supplied with a second longer-lasting antimalarial as a combination therapy or 'ACT'. Resistance to ACTs is now found in much of the greater Mekong region (41, 42). As yet, there is no strong evidence that resistance has spread from Asia to Africa, where it would be particularly devastating, but this has previously happened with antimalarials such as chloroquine and antifolates, and the ever-increasing global movement of people makes the transport of resistant parasites very likely. The current picture is complicated by the unusual nature of artemisinin resistance: a phenotype of 'delayed parasite clearance' in which parasites are cleared only slowly from the blood, and may go 'dormant' to survive the brief period of drug exposure before recrudescing (41, 43). This phenotype is difficult to measure in vitro, its genetic basis is only partially understood (44, 45), and it may be dependent on the genetic background of the parasite, perhaps explaining why it has developed in Asian but not yet in African strains (46, 47). As highlighted below, it will be imperative to preserve the effectiveness of the ACT antimalarials for as long as possible.

Developing an effective vaccine remains a huge challenge owing to the parasite's antigenic complexity, antigenic redundancy and capacity for antigenic variation (48). In 2018, the first ever vaccine for falciparum malaria, Mosquirix™, will begin to be supplied in three African countries, Ghana, Kenya and Malawi, supported by the WHO 'Malaria Vaccine Implementation Programme'. However, this programme remains exploratory and the vaccine is unlikely to be a game-changer in global malaria control because it does not offer

sterile or long-term protection. Mosquirix™ features an epitope from the invading 'sporozoite' stage of the parasite (fig 1A), and thus targets the pre-erythrocytic parasite stages in order to stimulate an immune response pre-empting symptomatic blood-stage malaria. Unfortunately, Phase-3 clinical trials revealed that the vaccine offered only ~30% protection, waning rapidly over a four-year period (49). Deployment in young children across Africa might still prevent millions of severe malaria episodes and deaths, but this must be weighed against concerns about cost, uptake, impact on other malaria control interventions, and the potential risk of shifting severe disease to older age groups (50).

Finally, in order to prevent malaria transmission it is vital to detect and then treat *P. falciparum* infections accurately, even when asymptomatic, because they may nevertheless still be transmitted by mosquitoes (51). People who have been repeatedly exposed tend to develop functional – albeit non-sterile – immunity to the parasite, suppressing infections to a level that causes few symptoms. These can be difficult to detect (because asymptomatic people do not seek treatment) and if the parasitaemia is very low they can also be difficult to diagnose without sensitive PCR-based tests or expert microscopy. Field diagnosis is often limited to antibody-based 'rapid diagnostic tests' (52), which frequently have lower sensitivity.

Future perspectives on P. falciparum

Arguably the most urgent current issue in the *P. falciparum* field is the threat now posed by artemisinin-resistant parasites. The community must work to understand the underlying biology of resistance, develop and deploy the right assays for it in the field – genetic, phenotypic or a combination of both – and thus track its spread across the malaria-endemic world. Retrospective studies have traced the emergence and spread of chloroquine resistance in the mid-1900s (53), but for the first time we now have the capacity to do this in real time, putting in place proactive interventions. Indeed, the genetic basis of artemisinin resistance was at least partially elucidated almost as it emerged, via a massive multicentre effort to sequence parasites and perform genome-wide association studies (44). It may be possible to prevent, or at least impede, the spread or *de novo* emergence of artemisinin-resistant parasites in Africa via in-depth surveillance, preventing the use of artemisinin monotherapy and using the right partner drugs in ACTs (since partner drugs are also at risk of resistance, invalidating the ACT approach (54)).

In parallel with this effort, since artemisinin will inevitably be lost sooner or later as an effective first-line drug, it is crucial to develop new drugs with different modes of action, and to improve their transit through the drug development pipeline (55). Product Development Partnerships such as the Medicines for Malaria Venture (MMV) are key players here.

Finally, returning to the eradication agenda, there are strong advocates for an audacious plan which was backed by the WHO in 2015 to halt the spread of artemisinin resistance by entirely eliminating *P. falciparum* from the Mekong region, where resistant parasites currently reside (56). This would probably require the unprecedented use of mass drug administration in complete populations – a logistical and ethical challenge – but the concept merits serious consideration if the WHO target of reducing malaria cases and deaths by 90% by 2030 is to be met. If successful, it could set a template for other regional elimination programmes.

P. falciparum is a fascinating and sophisticated parasite that has co-evolved with humans for thousands of years, shaping human genetics (57) and remaining a major public health problem to this day. There has never been a better moment for a concerted effort at the elimination, and eventually the global eradication, of this parasite.

Summary points

- *Plasmodium falciparum* is responsible for most of the global burden of death from malaria approximately half a million per annum.
- The *P. falciparum* parasite is an early-diverging eukaryote with many unusual and interesting biological features.
- Studying this parasite in the laboratory is challenging but great advances have been made in recent decades.
- Control of falciparum malaria has improved greatly in the past 15 years but is threatened by the repeated emergence of drug resistant parasites.

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203 **References**

- 1. WHO. World Malaria Report 2016. WHO website. 2016.
- 205 2. Snow RW. Sixty years trying to define the malaria burden in Africa: have we made
- 206 any progress? BMC Med. 2014;12:227.
- 207 3. Snow RW, Guerra CA, Noor AM, Myint HY, Hay SI. The global distribution of clinical
- episodes of Plasmodium falciparum malaria. Nature. 2005;434(7030):214-7.
- 209 4. WHO. Severe malaria. Trop Med Int Health. 2014;19 Suppl 1:7-131.
- 5. Janouskovec J, Horak A, Obornik M, Lukes J, Keeling PJ. A common red algal origin
- of the apicomplexan, dinoflagellate, and heterokont plastids. Proceedings of the National
- Academy of Sciences of the United States of America. 2010;107(24):10949-54.
- 213 6. van Dooren GG, Striepen B. The algal past and parasite present of the apicoplast.
- 214 Annu Rev Microbiol. 2013;67:271-89.
- 215 7. Cowman AF, Berry D, Baum J. The cellular and molecular basis for malaria parasite
- invasion of the human red blood cell. The Journal of cell biology. 2012;198(6):961-71.
- 217 8. Striepen B, Jordan CN, Reiff S, van Dooren GG. Building the perfect parasite: cell
- division in apicomplexa. PLoS pathogens. 2007;3(6):e78.
- 219 9. Liu W, Li Y, Learn GH, Rudicell RS, Robertson JD, Keele BF, et al. Origin of the
- 220 human malaria parasite Plasmodium falciparum in gorillas. Nature. 2010;467(7314):420-5.
- 10. Duval L, Fourment M, Nerrienet E, Rousset D, Sadeuh SA, Goodman SM, et al.
- 222 African apes as reservoirs of Plasmodium falciparum and the origin and diversification of the
- Laverania subgenus. Proceedings of the National Academy of Sciences of the United States
- 224 of America. 2010;107(23):10561-6.
- 11. Marsh K, Forster D, Waruiru C, Mwangi I, Winstanley M, Marsh V, et al. Indicators of
- life-threatening malaria in African children. N Engl J Med. 1995;332(21):1399-404.
- 12. Taylor WRJ, Hanson J, Turner GDH, White NJ, Dondorp AM. Respiratory
- 228 manifestations of malaria. Chest. 2012;142(2):492-505.
- 229 13. Otto TD, Rayner JC, Bohme U, Pain A, Spottiswoode N, Sanders M, et al. Genome
- 230 sequencing of chimpanzee malaria parasites reveals possible pathways of adaptation to
- 231 human hosts. Nat Commun. 2014;5:4754.
- 232 14. Smith JD, Rowe JA, Higgins MK, Lavstsen T. Malaria's deadly grip: cytoadhesion of
- Plasmodium falciparum-infected erythrocytes. Cellular microbiology. 2013;15(12):1976-83.
- 15. Milner DA, Jr., Whitten RO, Kamiza S, Carr R, Liomba G, Dzamalala C, et al. The
- 235 systemic pathology of cerebral malaria in African children. Front Cell Infect Microbiol.
- 236 2014;4:104.
- 16. Scherf A, Hernandez-Rivas R, Buffet P, Bottius E, Benatar C, Pouvelle B, et al.
- 238 Antigenic variation in malaria: in situ switching, relaxed and mutually exclusive transcription

- of var genes during intra-erythrocytic development in Plasmodium falciparum. The EMBO
- 240 journal. 1998;17(18):5418-26.
- 17. Gardner MJ, Hall N, Fung E, White O, Berriman M, Hyman RW, et al. Genome
- sequence of the human malaria parasite Plasmodium falciparum. Nature.
- 243 2002;419(6906):498-511.
- 18. Hamilton WL, Claessens A, Otto TD, Kekre M, Fairhurst RM, Rayner JC, et al.
- 245 Extreme mutation bias and high AT content in Plasmodium falciparum. Nucleic acids
- 246 research. 2016.
- 19. Salcedo-Amaya AM, van Driel MA, Alako BT, Trelle MB, van den Elzen AM, Cohen
- AM, et al. Dynamic histone H3 epigenome marking during the intraerythrocytic cycle of
- 249 Plasmodium falciparum. Proceedings of the National Academy of Sciences of the United
- 250 States of America. 2009;106(24):9655-60.
- 251 20. Stanojcic S, Kuk N, Ullah I, Sterkers Y, Merrick CJ. Single-molecule analysis reveals
- 252 that DNA replication dynamics vary across the course of schizogony in the malaria parasite
- 253 Plasmodium falciparum. Sci Rep. 2017;7(1):4003.
- 254 21. Kawamoto F, Alejo-Blanco R, Fleck SL, Sinden RE. Plasmodium berghei: ionic
- regulation and the induction of gametogenesis. Experimental parasitology. 1991;72(1):33-42.
- 256 22. Malaria Parasite Metabolic Pathways [Available from: http://mpmp.huji.ac.il/.
- 257 23. Spillman NJ, Beck JR, Goldberg DE. Protein export into malaria parasite-infected
- erythrocytes: mechanisms and functional consequences. Annu Rev Biochem. 2015;84:813-
- 259 41.
- 260 24. Sherling ES, van Ooij C. Host cell remodeling by pathogens: the exomembrane
- system in Plasmodium-infected erythrocytes. FEMS Microbiol Rev. 2016;40(5):701-21.
- 262 25. Moxon CA, Grau GE, Craig AG. Malaria: modification of the red blood cell and
- consequences in the human host. Br J Haematol. 2011;154(6):670-9.
- 26. Bushell E, Gomes AR, Sanderson T, Anar B, Girling G, Herd C, et al. Functional
- 265 Profiling of a Plasmodium Genome Reveals an Abundance of Essential Genes. Cell.
- 266 2017;170(2):260-72 e8.
- 267 27. WHO. Global technical strategy for Malaria 2016–2030. . 2016.
- 268 28. Bhatt S, Weiss DJ, Cameron E, Bisanzio D, Mappin B, Dalrymple U, et al. The effect
- of malaria control on Plasmodium falciparum in Africa between 2000 and 2015. Nature.
- 270 2015;526(7572):207-11.
- 271 29. Gething PW, Casey DC, Weiss DJ, Bisanzio D, Bhatt S, Cameron E, et al. Mapping
- 272 Plasmodium falciparum Mortality in Africa between 1990 and 2015. N Engl J Med.
- 273 2016;375(25):2435-45.
- 274 30. Liu J, Modrek S, Gosling RD, Feachem RG. Malaria eradication: is it possible? Is it
- worth it? Should we do it? Lancet Glob Health. 2013;1(1):e2-3.

- 276 31. Hemingway J, Shretta R, Wells TN, Bell D, Djimde AA, Achee N, et al. Tools and
- 277 Strategies for Malaria Control and Elimination: What Do We Need to Achieve a Grand
- 278 Convergence in Malaria? PLoS biology. 2016;14(3):e1002380.
- 279 32. Van Nieuwenhove S, Betu-Ku-Mesu VK, Diabakana PM, Declercq J, Bilenge CM.
- 280 Sleeping sickness resurgence in the DRC: the past decade. Trop Med Int Health.
- 281 2001;6(5):335-41.
- 282 33. Senaratne R, Singh PK. Against the odds, Sri Lanka eliminates malaria. Lancet.
- 283 2016;388(10049):1038-9.
- Newby G, Bennett A, Larson E, Cotter C, Shretta R, Phillips AA, et al. The path to
- eradication: a progress report on the malaria-eliminating countries. Lancet.
- 286 2016;387(10029):1775-84.
- 287 35. Trager W, Jensen JB. Human malaria parasites in continuous culture. Science (New
- 288 York, NY. 1976;193(4254):673-5.
- 289 36. Moon RW, Hall J, Rangkuti F, Ho YS, Almond N, Mitchell GH, et al. Adaptation of the
- 290 genetically tractable malaria pathogen Plasmodium knowlesi to continuous culture in human
- 291 erythrocytes. Proceedings of the National Academy of Sciences of the United States of
- 292 America. 2013;110(2):531-6.
- 293 37. Fidock DA, Wellems TE. Transformation with human dihydrofolate reductase renders
- 294 malaria parasites insensitive to WR99210 but does not affect the intrinsic activity of
- 295 proguanil. Proceedings of the National Academy of Sciences of the United States of
- 296 America. 1997;94(20):10931-6.
- 297 38. Shaw PJ, Aroonsri A. Tools for attenuation of gene expression in malaria parasites.
- 298 International journal for parasitology. 2017;47(7):385-98.
- 299 39. Malaria Genomic Epidemiology N. A global network for investigating the genomic
- 300 epidemiology of malaria. Nature. 2008;456(7223):732-7.
- 301 40. White J, 3rd, Mascarenhas A, Pereira L, Dash R, Walke JT, Gawas P, et al. In vitro
- adaptation of Plasmodium falciparum reveal variations in cultivability. Malaria journal.
- 303 2016;15:33.
- 304 41. Dondorp AM, Nosten F, Yi P, Das D, Phyo AP, Tarning J, et al. Artemisinin
- resistance in Plasmodium falciparum malaria. N Engl J Med. 2009;361(5):455-67.
- 306 42. Ashley EA, Dhorda M, Fairhurst RM, Amaratunga C, Lim P, Suon S, et al. Spread of
- artemisinin resistance in Plasmodium falciparum malaria. N Engl J Med. 2014;371(5):411-
- 308 23.
- 309 43. Witkowski B, Lelievre J, Barragan MJ, Laurent V, Su XZ, Berry A, et al. Increased
- tolerance to artemisinin in Plasmodium falciparum is mediated by a quiescence mechanism.
- Antimicrobial agents and chemotherapy. 2010;54(5):1872-7.

- 312 44. Ariey F, Witkowski B, Amaratunga C, Beghain J, Langlois AC, Khim N, et al. A
- 313 molecular marker of artemisinin-resistant Plasmodium falciparum malaria. Nature.
- 314 2014;505(7481):50-5.
- 315 45. Mok S, Ashley EA, Ferreira PE, Zhu L, Lin Z, Yeo T, et al. Drug resistance.
- Population transcriptomics of human malaria parasites reveals the mechanism of artemisinin
- 317 resistance. Science (New York, NY. 2015;347(6220):431-5.
- 318 46. Miotto O, Amato R, Ashley EA, MacInnis B, Almagro-Garcia J, Amaratunga C, et al.
- 319 Genetic architecture of artemisinin-resistant Plasmodium falciparum. Nature genetics.
- 320 2015;47(3):226-34.
- 321 47. Malaria GENPfCP. Genomic epidemiology of artemisinin resistant malaria. Elife.
- 322 2016;5.
- 323 48. Matuschewski K. Vaccines against malaria-still a long way to go. The FEBS journal.
- 324 2017.
- 325 49. Rts SCTP. Efficacy and safety of RTS,S/AS01 malaria vaccine with or without a
- booster dose in infants and children in Africa: final results of a phase 3, individually
- 327 randomised, controlled trial. Lancet. 2015;386(9988):31-45.
- 328 50. Penny MA, Verity R, Bever CA, Sauboin C, Galactionova K, Flasche S, et al. Public
- health impact and cost-effectiveness of the RTS,S/AS01 malaria vaccine: a systematic
- comparison of predictions from four mathematical models. Lancet. 2016;387(10016):367-75.
- 331 51. mal ERACGoD, Diagnostics. A research agenda for malaria eradication: diagnoses
- and diagnostics. PLoS Med. 2011;8(1):e1000396.
- 52. Mouatcho JC, Goldring JP. Malaria rapid diagnostic tests: challenges and prospects.
- 334 J Med Microbiol. 2013;62(Pt 10):1491-505.
- 53. Sa JM, Twu O, Hayton K, Reyes S, Fay MP, Ringwald P, et al. Geographic patterns
- of Plasmodium falciparum drug resistance distinguished by differential responses to
- amodiaguine and chloroguine. Proceedings of the National Academy of Sciences of the
- 338 United States of America. 2009;106(45):18883-9.
- 339 54. Amaratunga C, Lim P, Suon S, Sreng S, Mao S, Sopha C, et al. Dihydroartemisinin-
- 340 piperaquine resistance in Plasmodium falciparum malaria in Cambodia: a multisite
- prospective cohort study. Lancet Infect Dis. 2016;16(3):357-65.
- 342 55. Wells TN, Hooft van Huijsduijnen R, Van Voorhis WC. Malaria medicines: a glass
- 343 half full? Nat Rev Drug Discov. 2015;14(6):424-42.
- 344 56. Dondorp AM, Smithuis FM, Woodrow C, Seidlein LV. How to Contain Artemisinin-
- and Multidrug-Resistant Falciparum Malaria. Trends Parasitol. 2017;33(5):353-63.
- 346 57. Kwiatkowski DP. How malaria has affected the human genome and what human
- genetics can teach us about malaria. Am J Hum Genet. 2005;77(2):171-92.

- 348 58. Murray CJ, Ortblad KF, Guinovart C, Lim SS, Wolock TM, Roberts DA, et al. Global,
- regional, and national incidence and mortality for HIV, tuberculosis, and malaria during 1990-
- 2013: a systematic analysis for the Global Burden of Disease Study 2013. Lancet.
- 351 2014;384(9947):1005-70.

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Figure legend

Fig. 1: P. falciparum lifecycle, parasite structure and disease-control timeline

- A) Schematic showing the lifecycle of *P. falciparum*. Approximate parasite numbers shown at each stage highlight the severe bottlenecks and massive expansions at various stages. In the mosquito vector the sexual cycle occurs: pre-gametes called gametocytes are taken up in a blood meal from an infected human; these mature into gametes, mate and form a motile zygote called an ookinete which crosses the gut wall and encysts to form an oocyst. In the oocyst, asexual replication occurs and sporozoites are released to migrate to the salivary glands, whence they are injected into another human host during a mosquito bite. Sporozoites migrate from the bite site to the liver, where they multiply asexually inside hepatocytes over a period of ~7 days and then release merozoites which infect erythrocytes. In erythrocytes, 48-hour cycles of asexual replication, cell lysis and reinvasion occur, causing all the symptoms of malaria. A small subset of these parasites differentiates into gametocytes ready for mosquito transmission.
- B) Structure and organelles of *P. falciparum*. The apical complex that facilitates host cell invasion includes rhoptries, micronemes and dense granules, all containing proteins that are released during host cell invasion. The merozoite surface is densely coated with proteins that aid host cell attachment and are cleaved and shed during invasion. The two endosymbiont-derived organelles, mitochondrion and plastid, are also shown.
- C) Timeline showing malaria control interventions and the global burden of malaria deaths from 1990 to 2015 (+/- 95% confidence intervals). Data are from the Global Burden of Disease study (58), which records data from 1990 onwards; comparable global data prior to 1990 are lacking.

