Review article

Vector-focused approaches to curb malaria transmission in the Brazilian Amazon: an overview of current and future challenges and strategies

Elerson Matos Rocha¹; Ricardo de Melo Katak¹; Juan Campos de Oliveira¹; Maisa da Silva Araujo²; Bianca Cechetto Carlos³,⁴; Roberto Galizi⁵; Frederic Tripet⁵; Osvaldo Marinotti⁶*; Jayme A. Souza-Neto³,⁴*

- Programa de Pós-Graduação em Biotecnologia, Universidade Federal do Amazonas PPGBIOTEC / UFAM, Manaus, Brazil
- ² Laboratory of Medical Entomology, Oswaldo Cruz Foundation, FIOCRUZ, Porto Velho, RO, Brazil
- ³ São Paulo State University (UNESP), School of Agricultural Sciences, Department of Bioprocesses and Biotechnology, Botucatu, Brazil
- ⁴ São Paulo State University (UNESP), School of Agricultural Sciences, Central Multiuser Laboratory, Botucatu, Brazil
- 5 Centre of Applied Entomology and Parasitology, School of Life Sciences, Keele University, Staffordshire, UK
- ⁶ MTEKPrime, Aliso Viejo, CA, USA
- *Correspondence: jayme.souza-neto@unesp.br (JASN); omarinotti@gmail.com (OM)

Abstract: In Brazil, malaria transmission is mostly confined to the Amazon, where substantial progress has been achieved towards disease control in the past decade. Vector control has been historically considered a fundamental part of the main malaria control programs implemented in Brazil. However, the conventional vector-control tools have been insufficient to eliminate local vector populations due to the complexity of the Amazonian rainforest environment and ecological features of malaria vector species in the Amazon, especially *Anopheles darlingi*. Malaria elimination in Brazil and worldwide eradication will require a combination of conventional and new approaches that takes into account the regional specificities of vector populations and malaria transmission dynamics. Here we present an overview on both conventional and novel promising vector-focused tools to curb malaria transmission in the Brazilian Amazon. If well designed and employed, these new vector-based approaches may improve the implementation of malaria-control programs, particularly in remote or difficult-to-access areas and in regions where existing interventions have been unable to eliminate disease transmission. However, much effort still has to be put on research expanding the knowledge of neotropical malaria vectors to set the steppingstones for the development of such innovative tools.

Keywords: Malaria; Amazon; Brazil; *Anopheles darlingi; Plasmodium*; Control; Challenges; Strategies; Conventional; Novel; Vector; Mosquito

1. INTRODUCTION

Malaria eradication, defined as the permanent reduction to zero of the worldwide incidence of malaria infection, has been a major global and public health objective for decades. Progress toward eradication includes efforts for controlling and eventually eliminating malaria in specific geographic areas or countries. While control measures aim at reducing the number of new infections and the number of people infected in local settings, malaria elimination is accomplished when transmission ceases completely to occur locally. To achieve malaria elimination in Brazil and worldwide eradication, a combination of conventional and new approaches and tools will be necessary [1, 2].

In 2015, the World Health Organization (WHO) adopted The Global Technical Strategy for Malaria 2016–2030 providing guidance to countries in their efforts to achieve malaria elimination and setting a goal of reducing global malaria incidence and mortality rates by at least 90% by 2030 [3]. The Pan American Health Organization (PAHO) followed with the resolution CD55.R7, a Plan of action for malaria elimination in the Americas [4]. In the same year, the Brazilian National Malaria Control Program (NMCP) of the Ministry of Health launched the Plan for Elimination of the malaria-causing parasite *Plasmodium falciparum* in Brazil [5], acknowledging that *Plasmodium vivax* elimination is more challenging and may take longer, requiring specific tools and strategies for its containment, especially regarding the prevention of relapses [6].

Malaria is a vector-borne disease transmitted by anopheline mosquitoes [7]. Hence, vector control is a vital component of malaria prevention, control and elimination strategies [7, 8]. Here we review malaria control measures focused on mosquito vectors presently applied in the Brazilian Amazon and discuss their advantages and limitations. Furthermore, we discuss progresses in innovative vector control approaches aimed at curbing malaria transmission, which once fully developed may be incorporated into integrated mosquito management programs.

2. Malaria in Brazil, a brief history and current status

In Brazil, malaria affecting members of the native Tupinambá people was first reported in 1587, however, no epidemics were reported during the colonial period [9, 10]. By the end of the 19th century and beginning of the 20th century, in a changed scenario, malaria was endemic throughout the country, with approximately six million cases per year [9]. Since then, the disease was virtually eliminated in the southern areas, where nowadays only a few cases of autochthonous malaria transmission are reported annually [11, 12]. In contrast, malaria remains a major public health problem in Northern Brazil, mostly in the Amazon region where more than 99% of malaria cases currently occur [13, 14].

The first noticeable increase in the number of malaria cases in the Amazon region occurred during the Amazon Rubber Boom (1879 to 1912), when approximately half million immigrants moved to the area attracted by job opportunities in latex extraction, natural rubber industrial processing and related activities. Railroads were built to facilitate the transport and export of rubber products and improve accessibility to settled but isolated areas. One of these, the Madeira Mamoré railway, built between 1907 and 1912, was nicknamed the "Devil Railway" because thousands of workers died during its construction [9], largely due to the high number of mosquitoes spreading malaria in the settlements [15]. Just a few decades later, in the 1930s, *Anopheles arabiensis*, coming from Africa by sea, was introduced in Brazil [16, 17]. The spread of this efficient vector throughout Northeastern Brazil resulted in an epidemic with more than 150,000 cases and 14,000 deaths from malaria between 1938 and 1939. Fortunately, the invasive mosquito population was eliminated in 1940, thanks to management of breeding sites and insecticides sprayed in homes and vehicles [18, 19, 20].

Despite the elimination of *An. arabiensis* in the early 1940s, malaria continued to be transmitted in the Amazonian states by local anopheline vectors, with estimates of six to eight million people infected and 80,000 deaths annually during that decade [12]. In 1947, the Brazilian National Malaria Service implemented the use of dichlorodiphenyltrichloroethane (DDT) for vector control, and in 1950 of chloroquine for the treatment of patients infected with parasites [21, 22]. These measures resulted in significant reduction of malaria transmission, with only 36,900 cases reported in 1961 [18,

21]. The spread of malaria in the Brazilian Amazon raised again in the 60's with the building of new roads, followed in the 70's with the establishment of hydroelectric projects and in the 80's with the emergence of the gold prospecting sites [23, 24].

In the early 1970's another immigratory flow was triggered by land ownership opportunities in the Amazon region, when the National Institute for Colonization and Agrarian Reform - INCRA was donating plots nearby the new road network [25, 26]. These immigrants, unprepared and unaware, entered a region infested by mosquitoes, resulting in an epidemic with numbers that reached 300 thousand cases in a population of one million inhabitants in Rondônia state alone [18, 24, 25, 27]. In addition to the migratory wave, between 1970 and 1980, the increase in malaria cases in the Amazonian region was fueled by cuts in funding to sustain the social sectors, including vector control programs [28].

From 1990 to 2006, an average of 600,000 malaria cases were recorded annually [18, 19], however, intensification of vector control and other malaria prevention and treatment measures in the following years resulted in a steady and significant reduction in the incidence of malaria [27, 29]. Transmission intensity was maintained at 140 thousand cases yearly in 2014, 2015 and 2016. However, in 2017 and 2018, 197 and 207 thousand cases were registered respectively, a considerable growth compared to previous years [7]. The recent increase in reported malaria cases is mostly due to transmission occurring near the borders between Brazil and its neighbors French Guiana and Venezuela [30, 31, 32]. A timeline of malaria in the Brazilian Amazon is presented in figure 1.

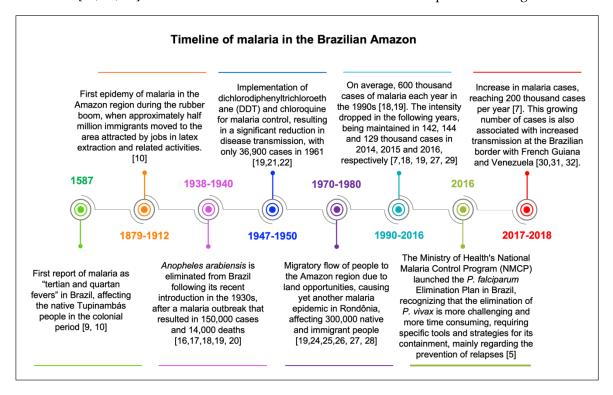


Figure 1. A brief timeline of the main events concerning malaria transmission and control in the Brazilian Amazon.

3. Malaria vectors in the Brazilian Amazon and the importance of vector population surveillance

Effective vector control in the Amazon is a complex and multifactorial task because of the sheer geographical scale of the region, its markedly heterogeneous ecology and complex human demographic aspects [33]. Urban, peri-urban and rural environments and areas of special interest (see below) present their unique challenges for controlling malaria transmission. It is clear that mosquito density and ecology as well as environmental conditions and human activity must be considered when designing vector-control measures adapted to regional specificities (Figure 2).

Zoophilic

Simbling species Breeding sites Biting activity Day/Night Crepuscular Biting behavior Frontier malaria Endophagic Exophagic Malaria vector Host preferences Vector Surveillance Anthropophilic

Challenges for malaria vector control

Figure 2. Main challenges for the development of novel vector-focused malaria control tools. Created with BioRender.com.

For instance, malaria control in areas of special interest such as native people settlements, areas of mining and national borders, present their own challenges. Malaria incidence in Brazilian native people is associated with environmental changes, their difficulty in accessing health services, and their mobility, at times resulting in migration to areas of more intense malaria transmission [34, 35]. Furthermore, culturally determined activities such as hunting, fishing, working in the fields, bathing along rivers and streams expose indigenous people to the risk of malaria infection. Gold mining plays a major role in spreading malaria in the area [36] since this activity creates puddles of water, ideal habitat for the reproduction of Anopheles mosquitoes. Transmission of malaria among miners (garimpeiros) is greatly affected by their constant mobility, and a work regime that coincides with the peak of biting activity of vector mosquitoes. Populations of border towns are generally more vulnerable, especially those living in remote areas, as recently observed along the border between French Guiana and Brazil where local inhabitants were affected by a malaria outbreak [37]. Recent events in Venezuela, causing mass migrations, has been responsible for an increase in malaria transmission across the frontier between Brazil and Venezuela [30, 31]. Each one of these scenarios demand unique malaria control programs, that include vector-control measures adapted to the regional context.

A total of fifty-four species of *Anopheles* are known to occur in Brazil, 33 of which are found in the Amazon region [38, 39]. Anopheles darlingi, An. albitarsis, An. braziliensis, An. argyritarsis, An. nunesztovari, An. oswaldoi, An. triannulatus, An. mattogrossensis, An. mediopunctatus and An. peryassui among other anopheline species are found throughout the Amazonian region [27] (Figure 3). All of those listed above are susceptible to natural Plasmodium infection, as demonstrated by using ELISA detection of the circumsporozoite protein (CSP), microscopy and/or PCR [27, 40]. Among those, the primary vector in the Amazonian Rainforest malaria transmission system is An. darlingi [27], which is abundant and highly anthropophilic [41, 42]. An. darlingi readily adapts to environmental changes caused by human activity and can easily develop in either artificial breeding sites such as fish farm tanks or in the midst of nature [43, 44].

Vector surveillance is essential to inform vector control strategies and evaluate their impact on malaria transmission. In the Amazon, vector surveillance has evidenced geographical and temporal differences in mosquito densities and species composition in malaria endemic areas [38, 45]. Factors driving this diversity are environmental [46, 47], anthropogenic [44] and biological [38, 48]. Hydrological cycles of the Amazon region include heavy rainfall between the months of November and June, resulting in the flooding of approximately 85,000 km2 of the Amazonian plain [49]. The end of each flooding cycle creates numerous large and small pools of water and slow-flowing streams suitable for mosquito breeding, as water levels slowly recede along rivers [44]. The large-scale climate interaction caused by environmental phenomena like the El Niño are associated with warmer temperatures, higher dew points, as well as reduced precipitation and river discharge in the Amazon, which may influence the dynamics of malaria transmission [50, 51].

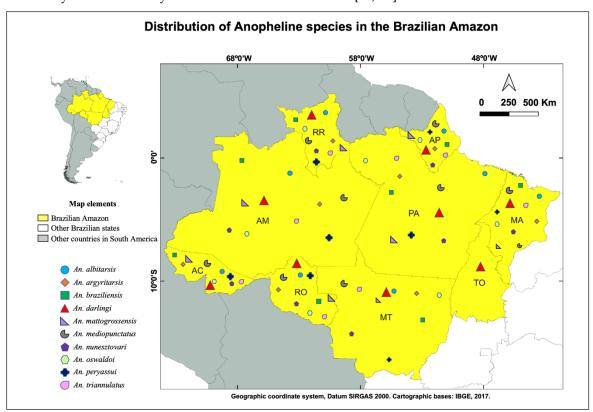


Figure 3. Distribution of the main anopheline species known to occur in the Brazilian Amazon, with emphasis in *Anopheles darlingi*, the main malaria vector in the region.

Anthropic activity in the Amazon has been associated with "Frontier malaria" a term commonly used to describe malaria transmission associated with deforestation and with the unplanned development of new agricultural settlements [52]. Newly deforested areas create multiple new *An. darlingi* breeding sites, favoring malaria transmission [53, 54]. However, anthropic activity may favor other vector species too. In Macapa, such changes reduced the suitability of breeding sites for *An. darlingi* and led to an increased density of *An. marajoara* [55].

Phenotypic plasticity, including features such as anthropophily, and endo-exophily, have been revealed by entomological surveys of neotropical malaria vectors [56, 57]. *An. darlingi* typically displays a single peak of biting activity before midnight [58, 59, 60, 61, 62, 63], however, in certain areas two peaks of biting activity are observed: one at dusk and another at dawn [64, 65, 66, 67, 68]. Furthermore, *An. darlingi* biting activity may occur in three peaks at sunset, midnight and dawn [69]. Observations at one locality showed considerable plasticity of this species' biting patterns as well, indicating that intra-population variation of biting activity can be as significant as inter-population

variation [70]. Single or multiple blood meals in each gonotrophic cycle, determined by biological or environmental factors also may influence the vectorial capacity of malaria mosquitoes [71].

The investigation of cryptic species among neotropical malaria vectors is essential for a better understanding of species distributions, behavior and population dynamics, leading to a better understanding of malaria transmission and adequate strategies for effective vector control in the Amazon. In Africa and in Asia several of the major malaria vectors belong to species complexes, including cryptic species that differ in host feeding preference, breeding sites, feeding behavior and role in malaria transmission [72, 73, 74]. These complexes may include both malaria vectors and nonvector species, which may either occur sympatrically or have distinct geographical distributions. While An. darlingi, the major malaria vector in the Brazilian Amazon is a monotypic species, other neotropical malaria vectors such as An. nuneztovari, An. albitarsis, An. triannulatus and An. oswaldoi are complexes of closely related, morphologically similar species [75]. For instance, An. konderi is often mistaken for An. oswaldoi [76], however An. konderi is present in human impacted or open areas, whereas An. oswaldoi is restricted to forested ones [77]. Marrelli et al. (1999) observed that both An. oswaldoi and An. konderi developed P. vivax oocysts in their midgut, but the complete cycle of the parasite, with sporozoites reaching the salivary glands, was only observed in An. oswaldoi, suggesting these species differ in vector competence [78]. Therefore, the refinement of taxonomic tools, including molecular taxonomic tools [79, 80] is paramount for the knowledge and understanding of the biology of these neotropical species complexes and for the investigation of malaria epidemiology. Furthermore, proper identification of species and knowledge of their ranges, often affected by changes in land use and lately in the climate, is vital for appropriate allocation of vector control resources.

4. Conventional measures for vector control and their limitations

The core and supplemental interventions for malaria vector control advocated by the World Health Organization and the Brazilian Ministry of Health are very similar as they represent a set of evidence-based guidelines (Table 1). The Brazilian Health Ministry acknowledges that not all areas should be subject to the same malaria control programs and follows a decentralized system in which each municipality adopts different control strategies. Also, it recognizes the existence of areas of special importance, particularly vulnerable and with limited access to interventions including. These include: Indigenous areas/indigenous tribes; gold mining camps; settlement areas; and frontier areas in the northern and western Brazilian Amazon rainforest.

Table 1. Current guidelines for malaria vector control in the world and in Brazil.

World Health Organization

Brazilian ministry of health

GUIDELINES FOR MALARIA VECTOR CONTROL 2019

https://apps.who.int/iris/bitstream/handle/10665/310862 /9789241550499-eng.pdf?ua=1

Malaria vector control, Policy guidance, Recommendations, accessed on May 5, 2020 https://www.who.int/malaria/policy-guidance/vector-control#tab=tab-2

Malaria vector control, Policy guidance, Operational manuals, accessed on May 5, 2020 https://www.who.int/malaria/policy-guidance/vector-control#tab=tab 3

Malaria: what it is, causes, symptoms, treatment, diagnosis and prevention, accessed on May 5, 2020 https://saude.gov.br/saude-de-a-z/malaria

Plano de Eliminação da malária no Brasil 2016 https://www.saude.gov.br/images/pdf/2017/janeiro/04/ Plano-eliminacao-malaria-pub.pdf

Guia de tratamento da malária no Brasil 2020 https://portalarquivos2.saude.gov.br/images/pdf/2020/janeiro/29/af-guia-tratamento-malaria-28jan20-isbn.pdf

Core interventions

INSECTICIDE-TREATED NETS - Pyrethroid-only Long-lasting insecticidal nets (LLINs) prequalified by WHO are recommended for deployment as a core intervention in all malaria-endemic settings. Pyrethroid-PBO nets prequalified by WHO are conditionally recommended for deployment instead of pyrethroid-only LLINs where the principal malaria vector(s) exhibit pyrethroid resistance.

Strongly recommended as an intervention of public health value, high-certainty evidence.

INSECTICIDE-TREATED NETS - Use of long-lasting impregnated mosquito nets in priority locations for each municipality and increase coverage in locations where LLIN is already used, together with monitoring of LLIN replacement plan to ensure availability.

INDOOR RESIDUAL SPRAYING (IRS) -

IRS deploying a product prequalified by WHO is recommended as a core intervention in all malaria-endemic settings. DDT has not been prequalified; it may be used for IRS if no equally effective and efficient alternative is available, and if it is used in line with the Stockholm Convention on Persistent Organic Pollutants.

Strongly recommended as an intervention of public health value, low-certainty evidence.

INDOOR RESIDUAL SPRAYING - IRS following technical recommendations of

the Brazilian Health Surveillance Secretariat (SVS), in buildings located in areas responsible for 80% of malaria transmission by

Infection, and in cycles that allow residual insecticide to be maintained throughout the year.

HOUSING AND WORKING PLACE
IMPROVEMENT - doors and windows screen
installation and maintenance.

Supplementary interventions

LARVICIDING - Regular application of biological or chemical insecticides to water bodies is recommended in areas where high coverage with a core intervention has been achieved, where aquatic habitats of the principal malaria vector(s) are few, fixed and findable, and where its application is both feasible and costeffective. Conditionally recommended as an intervention of public health value, low-certainty evidence.

LARVICIDING - Carrying out management of water collections to eliminate breeding sites of anopheles in urban locations with malaria transmission. Drainage; minor sanitation work to eliminate vector breeding sites; landfill; cleaning the margins of breeding sites; modification of water flow; control of aquatic vegetation.

Personal protection measures

TOPICAL REPELLENTS - Deployment of topical repellents is not recommended as a public health intervention; however, topical repellents may be beneficial as an intervention to provide personal protection.

Conditionally recommended against deployment as an intervention with public health value, low-certainty evidence.

TOPICAL REPELLENTS - DEET (N-N-dietilmetatoluamida)

INSECTICIDE-TREATED CLOTHING - Use of insecticide-treated clothing is not recommended as an intervention with public health value; however, insecticide-treated clothing may be beneficial as an intervention to provide personal protection in specific population groups.

Conditionally recommended against deployment as an intervention with public health value, low-certainty evidence.

CLOTHING - Clothing that protects legs (pants) and arms (long sleeve)

SPACE SPRAYING - Space spraying should not be undertaken for malaria control, and IRS or LLINs should be prioritized instead.

Conditionally recommended against deployment, very low-certainty evidence.

SPACE SPRAYING - Performing chemical spatial control, when in outbreak situations.

4.1. Long-lasting insecticidal nets (LLINs)

The use of LLINs is a highly cost-effective strategy for malaria prevention which has contributed to a significant reduction in disease morbidity and mortality worldwide. Until 2007, the WHO advocated distribution of LLINs only to pregnant women, children, and human immunodeficiency virus (HIV)-positive individuals. Since then, LLINs are recommended by WHO to all individuals at risk in endemic areas [81]. Accordingly, in Brazil, the Ministry of Health recommends the distribution and installation of LLINs, which are intended for personal overnight protection. Currently, LLINs impregnated with pyrethroid have a shelf life of 2-3 years [82]. Insecticide-treated bed-net incorporating a mixture of the pyrethroid alpha-cypermethrin and pyriproxyfen, an insect growth regulator, has been proposed as an alternative to pyrethroid only nets. Experimental evaluations showed more than 80% mortality and higher than 90% blood-feeding inhibition in the African malaria vector *An. gambiae* exposed to these LLINs. Furthermore, blood-fed female mosquitoes surviving net exposure suffered 83% reduction in oviposition and 95% reduction in offspring, indicating a potential improvement of malaria vector control when compared to standard pyrethroid-only LLINs [83].

Three hundred thousand LLINs were purchased by the Ministry of Health between 2015 and today. Despite the advantages of using LLINs subsidized by the government, data regarding the actual distribution and use of impregnated mosquito nets in this region is scarce [84, 85]. Local surveys indicate negative perceptions of LLINs, as they may cause skin irritations and allergies and are not effective in preventing malaria transmission occurring outdoors [86]. Low compliance, nets misuse, lack of LLIN replacement program, and local epidemiological factors may curtail the efficacy of impregnated bed nets for malaria control in the Amazon region.

4.2. Indoor residual spraying (IRS)

Another core measure for malaria vector control consists of spraying the walls of commercial buildings and residences with insecticides that remain on the applied surfaces [87]. Etofenprox PM 20% is the insecticide used in Brazil for residual spraying for malaria vector control. This product has a residual effect for 4 months requiring three annual applications [82]. In the Amazon, factors such as the operational cost of mobilizing teams to perform insecticide spraying, the difficulty in accessing remote areas at adequate frequency, the variability of dwellings, and variable environmental conditions, may compromise the efficacy of IRS [88]. In fact, few systematic evaluations on the impact these measures have on suppressing anopheline populations and reducing levels of malaria transmission in the Brazilian Amazon have been performed [82, 88]. While IRS is applicable and effective in urban and peri-urban environments, issues need to be addressed regarding gold miners who often live in huts without walls, continuously exposed to mosquito bites, unconventional indigenous house architecture that may not favor IRS and the lack of studies regarding the stability of insecticides applied on unconventional surfaces and under extreme humidity and temperature environmental conditions [89].

4.3. Larvae control

Supplemental interventions based on larval control are effective in reducing vector density and malaria transmission where aquatic habitats of the principal malaria vector(s) are few, fixed and findable, and where its application is both feasible and cost-effective [90, 91, 92]. Historical interventions largely based on larval control, including the previously mentioned eradication of *An. arabiensis* from Brazil [93], suggest that this approach may be an important asset in the battle for achieving malaria elimination and eradication.

Early larvicidal interventions involved environmentally damaging measures including elimination of breeding sites by filling depressions or draining swamps and application of toxic diesel or Paris green, impacting all organisms living in ponds, swamps, and other breeding sites. Nowadays, environmentally friendly alternatives are widely available. For example, biolarvicides based on the bacteria *Bacillus thuringiensis israelensis* (Bti) and/or *Lysinibacillus sphaericus*, (syn. *Bacillus sphaericus*, Bs), have been successfully applied for mosquito control in various ecological settings in sub Saharan Africa [94], Europe [95], Asia [96] and South America [97]. Aquatic insect predators and larvivorous fish have also been proposed as mosquito biocontrol agents, however, there is only limited evidence of their impact on disease transmission [98]. Density, diversity and habitat effects on the efficacy of natural mosquito larvae enemies must be considered. For instance, the presence of alternative preys, normally present in the extremely biodiverse Amazonian environment, together with the scarce selectivity of predators may limit the impact of such approaches on mosquito population [99].

Larvae control interventions require substantial knowledge of larval ecology due to the effects of weather and physical and biological characteristics of larval habitats on their efficacy. Furthermore, larviciding interventions are labor-intensive and to be effective must cover multiple *Anopheles* larval habitats often dispersed in vast areas to be effective [94]. Therefore, the effectiveness of larvicides in the Amazon, is limited to urban and peri urban environments, during the dry season and where the number of mosquito breeding sites is limited and easily accessible. Larviciding is not recommendable when and where breeding sites are inaccessible and countless, and in rural and frontier environments. Because of the territorial dimensions of the Amazon and the characteristics of breeding sites, in rural areas this procedure is restricted to the vicinity of inhabited settlements in ranches and farms and fishponds [100]. Recent developments using unmanned aerial vehicles coupled with high-resolution multispectral imagery to locate anopheline breeding sites could contribute to lower costs and improve coverage of larvae control programs in the Amazonian region [101].

4.4. Personal protection

Supplementary prophylactic measures against malaria include personal protection such as screens installed in windows and doors, clothing covering exposed parts of the body during biting periods, mosquito repellents, insecticides and air-conditioning [102]. Although highly recommended for travelers entering malaria endemic areas [103], most types of continuous personal protection may be neither reasonable nor affordable for local inhabitants due to economic and cultural issues. The typical climate conditions of the Amazon, with both high temperature and humidity, makes wearing long sleeves shirts and pants uncomfortable as a way of mosquito bite protection. Insecticide impregnated clothing is expensive and requires replacement or special treatment to maintain its protective function [104]. Mosquito repellents (DEET) require continuous reapplication and costs may be prohibitive. In Brazil DEET-based products (100 ml, of 6.7-7.1% DEET) costs on average 15.80 BRL equivalent to US\$3.84 [105]. Furthermore, the efficacy of commercially available repellents in protecting people from neotropical anophelines bites is still largely unknown. Local insecticide spraying is only manageable, sustainable and effective in confined environments, and its continuous application harms the environment [106]. Finally, housing improvements including netting and air conditioning are dependent on traditional architecture, windows and doors, as well as access to electricity and sufficient income that allows purchase, installation and maintenance of such home improvements.

5. Promising novel for vector control approaches

In view of the limitations of conventional techniques for vector-focused malaria control in the Amazon, novel and promising approaches to curb malaria transmission are under investigation and consideration.

5.1. Genetic control of malaria vectors

Genetic engineering of mosquitoes offers solutions and novel strategies to tackle the challenges encountered by current vector control interventions, i.e.; difficulties in deploying control measures to the affected regions, largely rural and dispersed in large areas; broad-spectrum activity of available insecticides and increasing spread of insecticide resistance. Such strategies rely on the release of modified insects carrying specific genetic traits, which act upon mating with the compatible species. This limits their impact on the ecosystem and, at the same time, facilitates the deployment of the intervention by taking advantage of the dispersal and mate-seeking behavior of the released mosquitoes.

5.1.1. Population suppression or replacement

Genetic control strategies can be aimed either to the "suppression" of the overall number of vectors or to their "replacement" with modified insects that are incapable or refractory to the transmission of the pathogen. Suppression strategies usually exploit the engineering of genetic traits that interfere with the reproductive capacity of insects and/or their fitness. Conversely, genetic modification for population replacement involves the introduction and expression of exogenous antiparasitic genes or the editing of the genetic components involved in vector-pathogen interactions to block or reduce the parasite development within the vector. Replacement modifications can be also intended to hinder the host-seeking behavior of insects thereby reducing their vector competence [107, 108, 109].

5.1.2. Self-limiting or sustaining strategies

The genetic control traits carried by the modified insect can be engineered to achieve different levels of persistence in the population after being released; this may vary from one, up to a virtually unlimited number of generations. Classic sterile insect technique (SIT) rely on repeated inundative releases of radio or chemical-sterilized males which can suppress the targeted populations by exploiting the single-mating capacity of female mosquitoes [110]. However, poor survival and mating competitiveness of the sterile males released are detrimental for the efficacy of these strategies [111].

The availability of genomic sequences and tools for the genetic modification of the mosquito genome allowed the engineering of alternative approaches based on the release of genetically modified male insects carrying a dominant lethal gene (RIDL), with the added benefit of a reduced impact on the fitness of released males compared to the classic methods [112]. With both approaches (SIT and RIDL), the sterile or dominant lethal traits carried by released males are not transmitted to the following generations minimizing long-term impacts and simplifying the risk assessment process leading to field applications. Both these technologies have been successfully applied for suppression of agricultural pests and vectors [112, 113, 114, 115] including the New World screwworm fly *Cochliomyia hominivorax* (cause of myiasis) in several American and African countries [116, 117], the malaria vector *Anopheles albimanus* in El Salvador [118], the dengue virus-transmitting *Aedes aegypti* in Brazil [119] and tsetse flies (*Glossina* spp.) carriers of the African trypanosomiasis (sleeping sickness) in the Zanzibar Island Unguja [120] among other examples. However, self-limiting methods, such as SIT and RIDL, require repeated mass releases of the modified insects, challenging their use for the treatment of large or remote geographic regions and/or non-isolated vector populations.

Beside the dominant sterility phenotype associated with these specific methods, genetic modifications are at best neutral or, in most of the cases, conferring a reduction of fitness to the carriers, resulting in a gradual removal of these traits from the population after release [121].

Approaches to overcome these limitations were theorized in the first half of the 20th century when both threshold-dependent and self-sustaining strategies were initially proposed [110, 122]. Threshold-dependent strategies, such as genetic underdominance, involve the introduction of genetic elements able to invade a population only if seeded above a certain frequency, which depend upon the fitness of released insects relative to wildtype [123]. On the other hand, self-spreading technologies such as gene drives (GD) offer the advantage of reducing the size of releases necessary to either suppress the targeted population or replace it with insects unable to transmit the parasite [121, 124, 125].

GD elements can be engineered to bias their own transmission by hijacking the mendelian partition of genetic material during germline formation of the vectors hosting such modification in their genome. For example, site-specific endonucleases, such as the increasingly popular CRISPR-Cas system, can be inserted into specific genomic sequences to disrupt the function of haplosufficient genes with role on female development [126, 127] or fertility [128]. The same endonuclease, active during the diploid stages of germline formation, is programmed to cut the target site on the homologous chromosome (not containing the CRISPR drive element). The double-strand DNA break stimulates the homology directed repair (HDR) machinery of the germ cell to repair the broken chromosome by using its homologous twin, carrying the GD, as a genetic template. As a result, the GD element is copied ("homed") to the homologous chromosome and transmitted to the entire progeny, instead of only half, thereby increasing in frequency over generations.

The short life cycle of mosquitoes allows a rapid increment of individuals heterozygous for the GD element (and the associated genetic disruption) in the population, even if released at low frequencies, progressively reducing the number of wildtypes. Mating between GD heterozygous mosquitoes generates individuals without functional copies of the targeted haplosufficient gene manifesting the disruptive phenotype; e.g. female sterile [128] or intersex XX individuals [126], causing suppression of the population. The same CRISPR-based gene drive element was also linked to a second nuclease targeting the X chromosome during male meiosis [129] to bias transmission of sperm in favor of those carrying the Y. As a result, super-mendelian transmission of the GD is accompanied by male biased progenies, thus presenting the advantage of reducing the fraction of biting females whilst suppressing the population [127]. Similarly, CRISPR-based GD constructs homing in neutral genetic loci were also engineered to spread anti-parasite molecules through caged mosquito populations [130, 131].

5.1.3. Challenges, alternatives and transfer to neotropical species (e.g. An. darlingi)

Besides the technical challenges, such as the selection of genetic resistance to the driving component or to the anti-malarial molecule, consistent research efforts have been focused over the last few years towards the development of new methods to limit or mitigate the spread of gene drive elements. The flexibility and modularity of CRISPR endonucleases prompted a variety of genetic control flavors with reduced penetrance as well as the development of novel countermeasures to gene drive spread [132, 133, 134, 135, 136]. The rapid progress in the laboratory and the potentials offered by these technologies are progressively shifting challenges towards the assessment of risk and ecological impact, regulation and acceptance prior field applications [137].

The flexibility offered by the molecular components used for genetic control offer the opportunity to transfer, with relative ease, these technologies to other species, such as *An. darlingi*, albeit the following resources being available: a laboratory-adapted inbred colony for genetic manipulation and testing in the laboratory; annotated genome and, favorable but not essential, a transcriptome for the selection of candidate genes and regulatory sequences for the expression of molecular effectors in the mosquitoes. Ad hoc transcriptomes may be unnecessary in the case orthologous genes may be retrievable from sibling species [138, 139, 140, 141, 142]. Successes with *An. darlingi* colonization [143, 144, 145] and genome sequence and annotation [146] and similar advances with other neotropical anophelines [147, 148, 149, 150] offer optimism that these technologies will soon be transferred to neotropical Amazonian malaria vectors.

5.2. Microbial-based approaches to control malaria transmission and malaria vector populations

5.2.1. Entomopathogenic organisms

Mosquitocidal microorganisms, including viruses, fungi and bacteria have been investigated as potential ecologically friendly alternatives to chemical insecticides [151, 152, 153, 154]. *Bacillus thuringiensis* var. *israelensis* (*Bti*) and *Lysinibacillus sphaericus* or *Bacillus sphaericus* (Bs) selectively kill mosquito larvae and have been used for decades with high efficacy and safety records [155, 156]. However, conventional Bti and Bs have low residual activity requiring repeated applications and increasing the cost of interventions [157, 158]. Long lasting microbial larvicide formulations with sustained release of Bti and Bs active ingredients for up to six months are currently commercially available [91, 159, 160, 161]. These new long-lasting larvicidal formulations associated with the use of drones to identify and map mosquito breeding sites, associated with aerial application of granular or aqueous Bti formulations may assist reducing the complex operational challenges that affect mosquito control in the Amazon environments [101, 162]. Besides Bti and Bs, microorganisms such as the bacteria *Chromobacterium* sp. Panama [163, 164] and the fungi *Beauveria bassiana* [165] and *Metarhizium anisopliae* [166], among others, have mosquitocidal activities. The present challenge is to convert these promising observations into products that are ready to be incorporated in mosquito control interventions.

Genetic engineering methods have been proposed to increase the pathogenicity, improve longer-term efficacy, and prevent or delay insect resistance to entomopathogenic microorganisms [166, 167]. The addition of *Bti* genes into *Bs* genomes to increase infectivity to mosquitoes [168, 169] and the expression of *Bt* and non-*Bt* derived mosquito toxins in readily transformable microorganisms such as *Chlorella desiccate*, *Pichia pastoris* and *Saccharomyces cerevisiae* have been investigated as alternatives to develop new microbial products for mosquito control [170, 171, 172].

5.2.2. Naturally occurring symbiotic microorganisms with anti-pathogen activity

Mosquito microbiomes modulate insect immunity and some naturally occurring symbiotic microorganisms are capable of hampering or blocking malaria parasites development within their vectors [173, 174, 175]. These symbionts have been proposed as agents to render mosquito populations refractory to *Plasmodium* [176]. For example, the *Serratia marcescens* strain Y1 promotes the activation of the insect immune system resulting in a reduction of the number of developing oocysts after mosquitoes are challenged with an infective blood meal [177]. Similarly, a *S. marcescens* strain isolated from *An. albimanus* impairs *P. vivax* infection in that vector [178]. Enterobacter species isolated from wild mosquito populations in Zambia also show anti-*Plasmodium* activity, likely through the production of reactive oxygen species (ROS) [173, 178, 179]. Bacteria of the genus *Asaia* induce a basal immunity in *Anopheles* mosquitoes leading to a decrease in the development of malaria parasites within their vectors [180].

The symbiotic yeast *Wickerhamomyces anomalus* strain (WaF17.12), isolated from the malaria vector mosquito *Anopheles stephensi*, has shown strong in-vitro anti-plasmodial activity. Mosquitoes colonized with WaF17.12 developed 65.2% fewer parasites than the control group [181]. More recently, a symbiont microsporidian (*Microsporidia MB*) that colonizes *An. arabiensis* from Kenya, were shown capable of blocking *P. falciparum* development and transmission, providing a new prospect for malaria control [182]. Host-baited traps, odour-baited traps, resting traps and sugarbaited traps have been proposed as possible ways of delivering these agents to mosquitoes [183].

5.2.3. Paratransgenesis

Paratransgenesis consists in genetically transformed mosquito symbionts such as fungi, viruses or bacteria to disrupt the transmission of vector-borne pathogens [184]. The perspective of applying paratransgenesis for malaria control has driven exciting research with promising results. Recombinant densovirus such as AgDNV, isolated from *An. gambiae*, can be used to infect mosquitoes and drive expression of anti-*Plasmodium* peptides to block parasite transmission or insect-specific toxins to reduce mosquito population density or mosquito lifespan [185]. Fungi carrying effector

genes that hinders *Plasmodium* development have been created [186]. Among the symbiotic bacteria found in malaria vectors, *Asaia*, *Pantoea*, *Serratia*, *Pseudomonas* and *Thorsellia* have been evaluated as candidates for paratransgenesis [187, 188, 189].

Proof-of-principle experiments conducted with *An. gambiae* and *An. stephensis* [190, 191, 192] suggest paratransgenesis can be developed in an actual tool for malaria control. Engineered *Pantoea agglomerans* expressing anti-plasmodial genes inhibit the development of malaria parasites by up to 98% and reduce the proportion of infected mosquitoes by 84% in lab settings [191]. Similar results were obtained with *Serratia*, strain AS1 [190]. With this in mind, the microbial flora associated with *An. darlingi* and other neotropical malaria vectors have been investigated [178, 193, 194, 195, 196, 197, 198, 199, 200, 201, 202]. Expanding the knowledge of culturable bacteria associated with this malaria vector and identifying symbiotic bacterial strains that are amenable to genetic manipulation, colonize *An. darlingi* efficiently and are transferred from adult females to their progeny, is essential for moving forward and testing the viability of paratransgenesis for malaria control in the Amazon. Recent detection of *Asaia sp* in *An. darlingi* further supports the prospect of using these bacteria as tools for malaria control through paratransgenesis in the Amazon [193, 199] although several challenges remain to be addressed for field applications (i.e. effectiveness, safety and release methods).

5.2.4. Wolbachia

Wolbachia are common intracellular endosymbiont bacteria present in up to 60% of insect species, including mosquitoes [203, 204, 205]. They are maternally inherited and can cause three kinds of reproductive alterations in their hosts: cytoplasmic incompatibility (CI), parthenogenesis and feminization [204, 206, 207, 208]. Furthermore, Wolbachia can inhibit the replication of pathogens in its arthropod hosts making these organisms a promising tool to combat mosquito-transmitted diseases [209, 210]. The successes of Wolbachia-based biocontrol of dengue and other arboviruses [211] suggest the possibility of similar Wolbachia-based strategies for malaria control.

Evidences of natural Wolbachia infections in malaria vectors [212, 213, 214] triggered investigations on the possible use of Wolbachia-Anopheles associations to limit malaria transmission. These efforts generated remarkable results showing reduced egg laying (population reduction) and a significantly reduced *Plasmodium* prevalence in mosquitoes carrying native *Wolbachia* infection (population replacement) [213, 215]. However, challenges remain for naturally occurring Wolbachia to be applicable as tools for malaria control. These strategies must rely on CI for Wolbachia to spread in natural populations and at present, it is not clear whether native Wolbachia can induce CI in anophelines. Induction of CI was not observed in caged experiments using wAnga-BF-infected An. coluzzii [215]. Nonetheless, Wolbachia-based malaria control strategies, such as population suppression or blocking of parasite development, are not only reliant on Wolbachia symbionts naturally associated with a given mosquito species. Successful dengue control was achieved with Aedes aegypti mosquitoes artificially infected with Wolbachia from a different insect species. Hence, the Wolbachia-based vector population suppression and disease transmission blocking can work in species not commonly infected with Wolbachia in the wild [216]. So far, the only Anopheles species amenable to Wolbachia transinfection in the laboratory is An. stephensi [217]. The wAlbB strain was used to stably infect An. stephensi, inducing complete CI, and conferring resistance to malaria parasites [218]. Recent studies suggest paratransgenesis could be exploited to circumvent difficulties in infecting malaria vectors with living Wolbachia strains. Wolbachia-derived molecules able to stimulate the immune system and modulate the mosquito vector competence, can be expressed in symbiotic bacteria affecting parasite development [219].

Wolbachia-based approaches for malaria control in the Amazon have not been investigated to date. Successful laboratory colonization of *An. darlingi* and other local malaria vectors [143, 144, 145], will allow attempts to transinfect these mosquitoes with *Wolbachia*. However, *Wolbachia*-based approaches for malaria control in the Amazon will depend on vertical transmission of *Wolbachia* to offspring. Additionally, research is needed to investigate if CI, parthenogenesis or feminization could be induced by *Wolbachia* infected and to identify *Wolbachia* strains which affect malaria parasites development in these mosquitoes.

6. Final remarks and perspectives

Despite the WHO's global efforts to control and eliminate malaria, the present malaria situation is still alarming, with an estimated 228 million of yearly cases of malaria occurring worldwide, causing more than 400 thousand deaths and predominantly affecting the poor and underprivileged (7WHO, 2019). While most malaria cases and related deaths occur in the World Health Organization (WHO) African Region (213 million or 93%), in 2018 the Americas reported more than 750 thousand confirmed malaria cases, with 130 million people living in areas at risk of malaria transmission. Approximately 200 thousand malaria cases were registered in the Brazilian Amazon in 2018. With the goal of providing the deserved health care to the thousands of people living in the Amazon, and in accordance with United Nations Sustainable Development Goal (SDG) 3, "ensure healthy lives and promote well-being for all at all ages" [220], increased investment and adequate planning will be necessary to eliminate malaria transmission in the area. Investments in malaria elimination and eradication are worthwhile, resulting in millions of lives saved, stimulating the economy and fostering prosperity, ensuring return on investment of billions of dollars [221, 222, 223, 224, 225, 226].

While malaria elimination in Brazil in the near future remains unlikely [6], researchers are exploring and developing novel and promising vector-based approaches to curb malaria transmission. Along with improvements of vaccines, drugs, diagnostic tools and insecticide-treated nets, these new vector-based approaches may prove crucial for the implementation of malaria-control programs, especially in regions where existing interventions have been unable to eliminate disease transmission. The efficacy and biosafety concerning these new technologies will need to be addressed via a stepwise regulatory framework before they can be incorporated into malaria control programs. Meanwhile, research expanding the knowledge of neotropical malaria vectors biology, ecology, behavior, physiology, genetics, biochemistry and insecticide resistance, primarily *An. darlingi*, is warranted as the basis on which vector-based malaria control in the Amazon may be founded.

Author contributions: All authors have read and agreed to the published version of the manuscript

Funding: JASN (FAPESP Young Investigator Award grant # 2013/11343-6).

Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

References

- 1. WHO WHO. From malaria control to malaria elimination: a manual for elimination scenario planning. 2014.
- 2. Antiporta DA, Rosas-Aguirre A, Chang J, Llanos-Cuentas A, Lescano AG. Malaria eradication. The Lancet. 2020;395 10233:e67.
- 3. WHO WHO. Global technical strategy for malaria 2016-2030. World Health Organization; 2015.
- 4. PAHO PAHO. Plan of action for malaria elimination 2016-2020. 55th Directing Council of PAHO, 68th Session of the Regional Committee of WHO for the Americas; Washington, DC, USA, Sep 26-30 2016 2016. https://iris.paho.org/bitstream/handle/10665.2/31440/CD55-13-e.pdf?sequence=1&isAllowed.
- 5. MS MdSdB. Plano de eliminação de malária no Brasil. 2016; Objetivos de Desenvolvimento Sustentável: ONU https://www.saude.gov.br/images/pdf/2017/janeiro/04/Plano-eliminacao-malaria-pub.pdf
- 6. Melo J, Padilha M, Barbosa R, Alonso W, Vittor A, Laporta G. Evaluation of the malaria elimination policy in Brazil: a systematic review and epidemiological analysis study. Tropical Biomedicine. 2020;37 2:513-35.
- 7. WHO WHO. World malaria report. 2019. https://www.who.int/publications/i/item/9789241565721.

- 8. Wilson AL, Courtenay O, Kelly-Hope LA, Scott TW, Takken W, Torr SJ, et al. The importance of vector control for the control and elimination of vector-borne diseases. PLoS neglected tropical diseases. 2020;14 1:e0007831.
- 9. Coura JR, Suárez-Mutis M, Ladeia-Andrade S. A new challenge for malaria control in Brazil: asymptomatic Plasmodium infection-a review. Memórias do Instituto Oswaldo Cruz. 2006;101 3:229-37.
- 10. Deane LM. Malaria vectors in Brazil. Mem Inst Oswaldo Cruz. 1986;81 Suppl II:5-14.
- 11. Multini LC, Marrelli MT, Beier JC, Wilke AB. Increasing Complexity Threatens the Elimination of Extra-Amazonian Malaria in Brazil. Trends in parasitology. 2019;35 6:383-7.
- 12. Pina-Costa Ad, Brasil P, Santi SMD, Araujo MPd, Suárez-Mutis MC, Oliveira-Ferreira J, et al. Malaria in Brazil: what happens outside the Amazonian endemic region. Memórias do Instituto Oswaldo Cruz. 2014;109 5:618-33.
- 13. MS MdSdB. Malária: o que é, causas, sintomas, tratamento, diagnóstico e prevenção Situação epidemiológica da malária. 2020. http://saude.gov.br/saude-de-a-z/malaria.
- 14. Sampaio VS, Siqueira AM, Alecrim MdGC, Mourão MPG, Marchesini PB, Albuquerque BC, et al. Malaria in the State of Amazonas: a typical Brazilian tropical disease influenced by waves of economic development. Revista da Sociedade Brasileira de Medicina Tropical. 2015;48:4-11.
- 15. Cruz OG. Madeira-Mamoré Railway Company: Considerações gerais sobre as condições sanitárias do rio Madeira. 1910. Cruz, Oswaldo Opera ominia Rio de Janeiro: Instituto Oswaldo Cruz. 1972:564-624.
- 16. Lopes G. Anopheles gambiae in Brazil: the background to a" silent spread," 1930-1932. Historia, ciencias, saude--Manguinhos. 2019;26 3:823-39.
- 17. Parmakelis A, Russello MA, Caccone A, Marcondes CB, Costa J, Forattini OP, et al. Historical analysis of a near disaster: Anopheles gambiae in Brazil. The American journal of tropical medicine and hygiene. 2008;78 1:176-8.
- 18. Ferreira MU, Castro MC. Challenges for malaria elimination in Brazil. Malaria journal. 2016;15 1:1-18.
- 19. Oliveira-Ferreira J, Lacerda MV, Brasil P, Ladislau JL, Tauil PL, Daniel-Ribeiro CT. Malaria in Brazil: an overview. Malaria journal. 2010;9 1:115.
- 20. Killeen GF. Following in Soper's footsteps: northeast Brazil 63 years after eradication of Anopheles gambiae. The Lancet Infectious diseases. 2003;3 10:663-6; doi: 10.1016/s1473-3099(03)00776-x.
- 21. Silva Rd, Paiva CHA. The Juscelino Kubitschek government and the Brazilian Malaria Control and Eradication Working Group: collaboration and conflicts in Brazilian and international health agenda, 1958-1961. História, Ciências, Saúde-Manguinhos. 2015;22 1:95-114.
- 22. da Silva R, Hochman G. Um método chamado Pinotti: sal medicamentoso, malária e saúde internacional (1952-1960). História, Ciências, Saúde-Manguinhos. 2011;18 2:519-43.
- 23. Marques AC. Migrations and the dissemination of malaria in Brazil. Mem Inst Oswaldo Cruz. 1986;81 Suppl II:17-30.
- 24. Marques AC. Human migration and the spread of malaria in Brazil. Parasitology today. 1987;3 6:166-70.
- 25. Camargo EP. Malária, maleita, paludismo. Ciência e cultura. 2003;55 1:26-9.
- 26. Fearnside PM. Deforestation in Brazilian Amazonia: history, rates, and consequences. Conservation biology. 2005;19 3:680-8.
- 27. Carlos BC, Rona LD, Christophides GK, Souza-Neto JA. A comprehensive analysis of malaria transmission in Brazil. Pathogens and Global Health. 2019;113 1:1-13.
- 28. Eichemberg Silva LC. O que mostram os Indicadores sobre a pobreza na década perdida. Rio de Janeiro: IPEA (Texto para discussão, 274). 1992.
- 29. Conn JE, Grillet ME, Correa M, Sallum MAM. Malaria transmission in South America—present status and prospects for elimination. Towards malaria elimination—a leap forward London: InTech. 2018:281-313.

- 30. Arruda-Barbosa Ld, Sales AFG, Souza ILLd. Reflexes of Venezuelan immigration on health care at the largest hospital in Roraima, Brazil: qualitative analysis. Saúde e Sociedade. 2020;29:e190730.
- 31. Grillet ME, Moreno JE, Hernandez JV, Vincenti-Gonzalez MF, Noya O, Tami A, et al. Malaria in Southern Venezuela: The Hottest Hotspot in Latin America. bioRxiv. 2020.
- 32. Mosnier E, Roux E, Cropet C, Lazrek Y, Moriceau O, Gaillet M, et al. Prevalence of Plasmodium spp. in the Amazonian Border Context (French Guiana–Brazil): Associated Factors and Spatial Distribution. The American Journal of Tropical Medicine and Hygiene. 2020;102 1:130-41.
- 33. Tauil PL. Perspectives of vector borne diseases control in Brazil. Revista da Sociedade Brasileira de Medicina Tropical. 2006;39 3.
- 34. Mendes AM, Lima MdS, Maciel AGP, Menezes RAdO, Eugênio NCC. Malaria among indigenous peoples on the Brazil-French Guiana border, 2007-2016: a descriptive study. Epidemiologia e Serviços de Saúde. 2020;29:e2019056.
- 35. Robortella DR, Calvet AA, Amaral LC, Fantin RF, Guimarães LFF, França Dias MH, et al. Prospective assessment of malaria infection in a semi-isolated Amazonian indigenous Yanomami community: Transmission heterogeneity and predominance of submicroscopic infection. PloS one. 2020;15 3:e0230643.
- 36. Douine M, Lambert Y, Musset L, Hiwat H, Blume LR, Marchesini P, et al. Malaria in Gold Miners in the Guianas and the Amazon: Current Knowledge and Challenges. Current Tropical Medicine Reports. 2020:1-11.
- 37. Mosnier E, Dusfour I, Lacour G, Saldanha R, Guidez A, Gomes MS, et al. Resurgence risk for malaria, and the characterization of a recent outbreak in an Amazonian border area between French Guiana and Brazil. BMC infectious diseases. 2020;20:1-14.
- 38. Consoli RA, Oliveira RLd. Principais mosquitos de importância sanitária no Brasil. Editora Fiocruz; 1994.
- Tadei WP, Dutary Thatcher B. Malaria vectors in the Brazilian Amazon: Anopheles of the subgenus Nyssorhynchus. Revista do Instituto de Medicina Tropical de São Paulo. 2000;42 2:87-94.
- 40. Marrelli MT, Sallum MAM, Marinotti O. The second internal transcribed spacer of nuclear ribosomal DNA as a tool for Latin American anopheline taxonomy: a critical review. Memórias do Instituto Oswaldo Cruz. 2006;101 8:817-32.
- 41. Galardo AKR, Arruda M, COUTO AADA, Wirtz R, Lounibos LP, Zimmerman RH. Malaria vector incrimination in three rural riverine villages in the Brazilian Amazon. The American journal of tropical medicine and hygiene. 2007;76 3:461-9.
- 42. Klein TA, Lima J. Seasonal distribution and biting patterns of Anopheles mosquitoes in Costa Marques, Rondonia, Brazil. Journal of the American Mosquito Control Association. 1990;6 4:700-7.
- 43. Arcos AN, da Silva Ferreira FA, da Cunha HB, Tadei WP. Characterization of artificial larval habitats of Anopheles darlingi (Diptera: Culicidae) in the Brazilian Central Amazon. Revista Brasileira de Entomologia. 2018;62 4:267-74.
- 44. Tadei WP, Rodrigues IB, Rafael MS, Sampaio RTdM, Mesquita H, Pinheiro VCS, et al. Adaptative processes, control measures, genetic background, and resilience of malaria vectors and environmental changes in the Amazon region. Hydrobiologia. 2017;789 1:179-96.
- 45. Lourenço-de-Oliveira R, Luz SL. Simian malaria at two sites in the Brazilian Amazon-II: Vertical distribution and frequency of anopheline species inside and outside the forest. Memórias do Instituto Oswaldo Cruz. 1996;91 6:687-94.
- 46. Forattini OP. Culicidologia médica: identificação, biologia e epidemiologia: v. 2. Culicidologia médica: identificação, biologia e epidemiologia: v 2; 2002. p. 860-.
- 47. Tadei WP, Santos JMMd, Rodrigues IB, Rafael MS. Malária e Dengue na Amazônia: vetores e estratégias de controle. Pesquisa e Ações em Saúde nos Institutos de Pesquisa do Ministério da Ciência e Tecnologia, pgs 113-125. 2010.

- 48. Rafael MS, Rohde C, Bridi LC, Tadei WP. Salivary polytene chromosome map of Anopheles darlingi, the main vector of neotropical malaria. The American journal of tropical medicine and hygiene. 2010;83 2:241-9.
- 49. Wolfarth-Couto B, Filizola N, Durieux L. Seasonal pattern of malaria cases and the relationship with hydrologic variability in the Amazonas State, Brazil. Revista Brasileira de Epidemiologia. 2020;23:e200018.
- 50. Olson SH, Gangnon R, Elguero E, Durieux L, Guégan J-F, Foley JA, et al. Links between climate, malaria, and wetlands in the Amazon Basin. Emerging infectious diseases. 2009;15 4:659.
- 51. Wolfarth BR, Filizola N, Tadei WP, Durieux L. Epidemiological analysis of malaria and its relationships with hydrological variables in four municipalities of the State of Amazonas, Brazil. Hydrological sciences journal. 2013;58 7:1495-504.
- 52. Souza PF, Xavier DR, Suarez Mutis MC, da Mota JC, Peiter PC, de Matos VP, et al. Spatial spread of malaria and economic frontier expansion in the Brazilian Amazon. Plos one. 2019;14 6:e0217615.
- 53. Chaves LSM, Fry J, Malik A, Geschke A, Sallum MAM, Lenzen M. Global consumption and international trade in deforestation-associated commodities could influence malaria risk. Nat Commun. 2020;11 1:1258; doi: 10.1038/s41467-020-14954-1.
- 54. MacDonald AJ, Mordecai EA. Amazon deforestation drives malaria transmission, and malaria burden reduces forest clearing. Proceedings of the National Academy of Sciences of the United States of America. 2019;116 44:22212-8; doi: 10.1073/pnas.1905315116.
- 55. Conn JE, Wilkerson RC, Segura MNO, de Souza RT, Schlichting CD, Wirtz RA, et al. Emergence of a new Neotropical malaria vector facilitated by human migration and changes in land use. The American journal of tropical medicine and hygiene. 2002;66 1:18-22.
- 56. Charlwood J. Biological variation in Anopheles darlingi Root. Memórias do Instituto Oswaldo Cruz. 1996;91 4:391-8.
- 57. Montoya-Lerma J, Solarte YA, Giraldo-Calderón GI, Quiñones ML, Ruiz-López F, Wilkerson RC, et al. Malaria vector species in Colombia: a review. Memórias do Instituto Oswaldo Cruz. 2011;106:223-38.
- 58. Elliott R. The influence of vector behavior on malaria transmission. The American Journal of Tropical Medicine and Hygiene. 1972;21 5_Suppl:755-63.
- 59. Hudson J. Anopheles darlingi Root (Diptera: Culicidae) in the Suriname rain forest. Bull Entomol Res. 1984;74 01:129-42.
- 60. Magris M, Rubio-Palis Y, Menares C, Villegas L. Vector bionomics and malaria transmission in the Upper Orinoco River, Southern Venezuela. Memórias do Instituto Oswaldo Cruz. 2007;102 3:303-12.
- 61. Quiñones M, Suarez M. Indoor resting heights of some anophelines in Colombia. Journal of the American Mosquito Control Association. 1990;6 4:602-4.
- 62. Rozendaal J. Biting and resting behavior of Anopheles darlingi in the Suriname rainforest. J Am Mosq Control Assoc. 1989;5 3:351-8.
- 63. Vittor AY, Gilman RH, Tielsch J, Glass G, Shields T, Lozano WS, et al. The effect of deforestation on the human-biting rate of Anopheles darlingi, the primary vector of falciparum malaria in the Peruvian Amazon. The American journal of tropical medicine and hygiene. 2006;74 1:3-11.
- 64. Charlwood J, Wilkes T. Studies on the age-composition of samples of Anopheles darlingi Root (Diptera: Culicidae) in Brazil. Bulletin of Entomological Research. 1979;69 2:337-42.
- 65. Forattini OP. Comportamento exófilo de Anopheles darlingi Root, em região meridional do Brasil. Revista de Saúde Pública. 1987;21:291-304.
- 66. Lourenço-de-Oliveira R, Guimarães AEdG, Arlé M, Silva TFd, Castro MG, Motta MA, et al. Anopheline species, some of their habits and relation to malaria in endemic areas of Rondonia State, Amazon region of Brazil. Memórias do Instituto Oswaldo Cruz. 1989;84 4:501-14.
- 67. Roberts D, Alecrim W, Tavares A, Radke M. The house-frequenting, host-seeking and resting behavior of Anopheles darlingi in southeastern Amazonas, Brazil. Journal of the American Mosquito Control Association. 1987;3 3:433.

- 68. Tadei WP, Thatcher BD, Santos J, Scarpassa VM, Rodrigues IB, Rafael MS. Ecologic observations on anopheline vectors of malaria in the Brazilian Amazon. The American journal of tropical medicine and hygiene. 1998;59 2:325-35.
- 69. Freitas-Sibajev M, Conn J, Mitchell S, Cockburn A, Seawright J, Momen H. Mitochondrial DNA and morphological analyses of Anopheles darlingi populations from Brazil (Diptera: Culicidae). Mosq Syst. 1995;27:78-99.
- 70. Voorham J. Intra-population plasticity of Anopheles darlingi's (Diptera, Culicidae) biting activity patterns in the state of Amapá, Brazil. Revista de Saúde Pública. 2002;36:75-80.
- 71. de Oliveira CD, Tadei WP, Abdalla FC, Paolucci Pimenta PF, Marinotti O. Multiple blood meals in Anopheles darlingi (Diptera: Culicidae). Journal of Vector Ecology. 2012;37 2:351-8.
- 72. Barrón MG, Paupy C, Rahola N, Akone-Ella O, Ngangue MF, Wilson-Bahun TA, et al. A new species in the major malaria vector complex sheds light on reticulated species evolution. Scientific Reports. 2019;9 1:14753; doi: 10.1038/s41598-019-49065-5. https://doi.org/10.1038/s41598-019-49065-5.
- 73. Sinka ME, Bangs MJ, Manguin S, Chareonviriyaphap T, Patil AP, Temperley WH, et al. The dominant Anopheles vectors of human malaria in the Asia-Pacific region: occurrence data, distribution maps and bionomic précis. Parasit Vectors. 2011;4 1:89; doi: 10.1186/1756-3305-4-89. https://doi.org/10.1186/1756-3305-4-89.
- 74. Tripathy A, Samanta L, Das S, Parida SK, Marai N, Hazra RK, et al. Distribution of sibling species of Anopheles culicifacies s.l. and Anopheles fluviatilis s.l. and their vectorial capacity in eight different malaria endemic districts of Orissa, India. Memórias do Instituto Oswaldo Cruz. 2010;105:981-7. http://www.scielo.br/scielo.php?script=sci_arttext&pid=S0074-02762010000800006&nrm=iso.
- 75. Rosa-Freitas MG, Lourenço-de-Oliveira R, Carvalho-Pinto CJd, Flores-Mendoza C, Silva-do-Nascimento TF. Anopheline Species Complexes in Brazil. Current Knowledge of Those Related to Malaria Transmission. Memórias do Instituto Oswaldo Cruz. 1998;93:651-5. http://www.scielo.br/scielo.php?script=sci arttext&pid=S0074-02761998000500016&nrm=iso.
- 76. Saraiva JF, Souto RNP, Scarpassa VM. Molecular taxonomy and evolutionary relationships in the Oswaldoi-Konderi complex (Anophelinae: Anopheles: Nyssorhynchus) from the Brazilian Amazon region. PLoS One. 2018;13 3:e0193591; doi: 10.1371/journal.pone.0193591.
- 77. Klein TA, Lima JB, Tada MS. Comparative susceptibility of anopheline mosquitoes to Plasmodium falciparum in Rondonia, Brazil. Am J Trop Med Hyg. 1991;44 6:598-603; doi: 10.4269/ajtmh.1991.44.598.
- 78. Marrelli MT, Honório NA, Flores-Mendoza C, Lourenco-de-Oliveira R, Marinotti O, Kloetzel JK. Comparative susceptibility of two members of the Anopheles oswaldoi complex, An. oswaldoi and An. konderi, to infection by Plasmodium vivax. Transactions of The Royal Society of Tropical Medicine and Hygiene. 1999;93 4:381-4; doi: 10.1016/s0035-9203(99)90123-2. https://doi.org/10.1016/S0035-9203(99)90123-2.
- 79. Bourke BP, Conn JE, de Oliveira TMP, Chaves LSM, Bergo ES, Laporta GZ, et al. Exploring malaria vector diversity on the Amazon Frontier. Malaria journal. 2018;17 1:342; doi: 10.1186/s12936-018-2483-2.
- 80. Swain SN, Makunin A, Dora AS, Barik TK. SNP barcoding based on decision tree algorithm: A new tool for identification of mosquito species with special reference to Anopheles. Acta Trop. 2019;199:105152; doi: 10.1016/j.actatropica.2019.105152.
- 81. WHO WHO. Global Malaria Programme: Insecticide-Treated Mosquito Nets: A WHO Position Statement. World Health Organization. 2007.
- 82. Baia-da-Silva DC, Brito-Sousa JD, Rodovalho SR, Peterka C, Moresco G, Lapouble OMM, et al. Current vector control challenges in the fight against malaria in Brazil. Revista da Sociedade Brasileira de Medicina Tropical. 2019;52.
- 83. Ngufor C, Agbevo A, Fagbohoun J, Fongnikin A, Rowland M. Efficacy of Royal Guard, a new alpha-cypermethrin and pyriproxyfen treated mosquito net, against pyrethroid-resistant malaria vectors. Scientific reports. 2020;10 1:1-15.

- 84. de Oliveira Sousa J, de Albuquerque BC, Coura JR, Suárez-Mutis MC. Use and retention of long-lasting insecticidal nets (LLINs) in a malaria risk area in the Brazilian Amazon: a 5-year follow-up intervention. Malaria journal. 2019;18 1:100.
- 85. Vieira GdD, Basano SdA, Katsuragawa TH, Camargo LMA. Insecticide-treated bed nets in Rondônia, Brazil: evaluation of their impact on malaria control. Revista do Instituto de Medicina Tropical de São Paulo. 2014;56 6:493-7.
- 86. Murta FLG, Mendes MO, Sampaio VS, Junior ASB, Díaz-Bermúdez XP, Monteiro WM, et al. Misperceptions of patients and health workers regarding malaria elimination in the Brazilian Amazon: a qualitative study. Malaria journal. 2019;18 1:223.
- 87. WHO WHO. Indoor residual spraying: an operational manual for indoor residual spraying (IRS) for malaria transmission control and elimination. World Health Organization; 2015.
- 88. Corrêa APS, Galardo AK, Lima LA, Câmara DC, Müller JN, Barroso JFS, et al. Efficacy of insecticides used in indoor residual spraying for malaria control: an experimental trial on various surfaces in a "test house". Malaria journal. 2019;18 1:345.
- 89. Dengela D, Seyoum A, Lucas B, Johns B, George K, Belemvire A, et al. Multi-country assessment of residual bio-efficacy of insecticides used for indoor residual spraying in malaria control on different surface types: results from program monitoring in 17 PMI/USAID-supported IRS countries. Parasit Vectors. 2018;11 1:1-14.
- 90. Antonio-Nkondjio C, Sandjo NN, Awono-Ambene P, Wondji CS. Implementing a larviciding efficacy or effectiveness control intervention against malaria vectors: key parameters for success. Parasit Vectors. 2018;11 1:57.
- 91. Fontoura PS, da Costa AS, Ribeiro FS, Ferreira MS, Castro MC, Ferreira MU. Field Efficacy of VectoMax FG and VectoLex CG Biological Larvicides for Malaria Vector Control in Northwestern Brazil. Journal of medical entomology. 2020;57 3:942-6.
- 92. WHO WHO. Larval source management: a supplementary malaria vector control measure: an operational manual. 2013. https://apps.who.int/iris/bitstream/handle/10665/85379/9789241505604_eng.pdf.
- 93. Soper FL, Wilson DB. Anopheles gambiae in Brazil 1930 to 1940. Anopheles gambiae in Brazil 1930 to 1940. 1943.
- 94. Derua YA, Kweka EJ, Kisinza WN, Githeko AK, Mosha FW. Bacterial larvicides used for malaria vector control in sub-Saharan Africa: review of their effectiveness and operational feasibility. Parasit Vectors. 2019;12 1:426.
- 95. Becker N. Microbial control of mosquitoes: management of the Upper Rhine mosquito population as a model programme. Parasitology Today. 1997;13 12:485-7.
- 96. Kumar A, Sharma V, Sumodan P, Thavaselvam D. Field trials of biolarvicide Bacillus thuringiensis var. israelensis strain 164 and the larvivorous fish Aplocheilus blocki against Anopheles stephensi for malaria control in Goa, India. Journal of the American Mosquito Control Association. 1998;14 4:457-62.
- 97. Kroeger A, Horstick O, Riedl C, Kaiser A, Becker N. The potential for malaria control with the biological larvicide Bacillus thuringiensis israelensis (Bti) in Peru and Ecuador. Acta Tropica. 1995;60 1:47-57.
- 98. Dambach P. The use of Aquatic Predators for Larval Control of Mosquito Disease Vectors: Opportunities and Limitations. Biological Control. 2020:104357.
- 99. Cuthbert RN, Dalu T, Wasserman RJ, Weyl OL, Froneman PW, Callaghan A, et al. Alternative prey impedes the efficacy of a natural enemy of mosquitoes. Biological Control. 2020;141:104146.
- 100. Cavados CFG, Tadei WP, Roque RA, Regis LN, de Oliveira CMF, Gil HB. Bacillus Entomopathogenic Based Biopesticides in Vector Control Programs in Brazil. Bacillus thuringiensis and Lysinibacillus sphaericus: Springer; 2017. p. 223-37.
- 101. Carrasco-Escobar G, Manrique E, Ruiz-Cabrejos J, Saavedra M, Alava F, Bickersmith S, et al. High-accuracy detection of malaria vector larval habitats using drone-based multispectral imagery. PLoS neglected tropical diseases. 2019;13 1:e0007105.
- 102. Schoepke A, Steffen R, Gratz N. Effectiveness of personal protection measures against mosquito bites for malaria prophylaxis in travelers. Journal of travel medicine. 1998;5 4:188-92.

- 103. Loutan L. Malaria: still a threat to travellers. International journal of antimicrobial agents. 2003;21 2:158-63.
- 104. Richards SL, Agada N, Balanay JAG, White AV. Permethrin treated clothing to protect outdoor workers: evaluation of different methods for mosquito exposure against populations with differing resistance status. Pathogens and global health. 2018;112 1:13-21.
- 105. Tan K, Faierstein GB, Xu P, Barbosa RM, Buss GK, Leal WS. A popular Indian clove-based mosquito repellent is less effective against Culex quinquefasciatus and Aedes aegypti than DEET. PloS one. 2019;14 11:e0224810.
- 106. Roberts DR, Tren R. International advocacy against DDT and other public health insecticides for malaria control. Research and Reports in Tropical Medicine. 2011;2:23.
- 107. Hoermann A, Tapanelli S, Capriotti P, Masters EK, Habtewold T, Christophides GK, et al. Converting endogenous genes of the malaria mosquito into simple non-autonomous gene drives for population replacement. bioRxiv. 2020.
- 108. Raban RR, Marshall JM, Akbari OS. Progress towards engineering gene drives for population control. Journal of Experimental Biology. 2020;223 Suppl 1.
- 109. Terenius O, Marinotti O, Sieglaff D, James AA. Molecular genetic manipulation of vector mosquitoes. Cell Host & Microbe. 2008;4 5:417-23.
- 110. Serebrovsky A. On the possibility of a new method for the control of insect pests. Sterile-male technique for eradication or control of harmful insects Proceedings of a panel on application of the sterile-male technique for the eradication or control of harmful species of insects, organised by the Joint FAO/IAEA Division of Atomic Energy in Food and Agriculture and held in Vienna, 27-31 May 1968. 1969:123-237.
- 111. Benedict MQ, Robinson AS. The first releases of transgenic mosquitoes: an argument for the sterile insect technique. Trends in parasitology. 2003;19 8:349-55.
- 112. Thomas DD, Donnelly CA, Wood RJ, Alphey LS. Insect population control using a dominant, repressible, lethal genetic system. Science. 2000;287 5462:2474-6.
- 113. Knipling EF. Sterile-Male Method of Population Control: Successful with some insects, the method may also be effective when applied to other noxious animals. Science. 1959;130 3380:902-4.
- 114. Mastrangelo T, Welch JB. An overview of the components of AW-IPM campaigns against the New World screwworm. Insects. 2012;3 4:930-55.
- 115. Orozco-Dávila D, Quintero L, Hernández E, Solís E, Artiaga T, Hernández R, et al. Mass rearing and sterile insect releases for the control of A nastrepha spp. pests in Mexico–a review. Entomologia Experimentalis et Applicata. 2017;164 3:176-87.
- 116. Lindquist D, Abusowa M, Hall M. The New World screwworm fly in Libya: a review of its introduction and eradication. Medical and Veterinary Entomology. 1992;6 1:2-8.
- 117. Vargas-Terán M, Hursey B, Cunningham E. Eradication of the screwworm from Libya using the sterile insect technique. Parasitology today. 1994;10 3:119-22.
- 118. Lowe RE, Bailey DL, Dame DA, Savage KE, Kaiser PE. Efficiency of techniques for the mass release of sterile male Anopheles albimanus Wiedemann in El Salvador. The American journal of tropical medicine and hygiene. 1980;29 4:695-703.
- 119. Carvalho DO, McKemey AR, Garziera L, Lacroix R, Donnelly CA, Alphey L, et al. Suppression of a field population of Aedes aegypti in Brazil by sustained release of transgenic male mosquitoes. PLoS neglected tropical diseases. 2015;9 7:e0003864.
- 120. Vreysen MJ, Saleh K, Mramba F, Parker A, Feldmann U, Dyck VA, et al. Sterile insects to enhance agricultural development: the case of sustainable tsetse eradication on Unguja Island, Zanzibar, using an area-wide integrated pest management approach. PLoS Negl Trop Dis. 2014;8 5:e2857.
- 121. Hammond AM, Galizi R. Gene drives to fight malaria: current state and future directions. Pathogens and global health. 2017;111 8:412-23.
- 122. Curtis C. Possible use of translocations to fix desirable genes in insect pest populations. Nature. 1968;218 5139:368-9.

- 123. Leftwich PT, Edgington MP, Harvey-Samuel T, Carabajal Paladino LZ, Norman VC, Alphey L. Recent advances in threshold-dependent gene drives for mosquitoes. Biochemical Society Transactions. 2018;46 5:1203-12.
- 124. Flores HA, O'Neill SL. Controlling vector-borne diseases by releasing modified mosquitoes. Nature reviews Microbiology. 2018;16 8:508-18; doi: 10.1038/s41579-018-0025-0.
- 125. Quinn CM, Nolan T. Nuclease-based gene drives, an innovative tool for insect vector control: advantages and challenges of the technology. Curr Opin Insect Sci. 2020;39:77-83; doi: 10.1016/j.cois.2020.03.007.
- 126. Kyrou K, Hammond AM, Galizi R, Kranjc N, Burt A, Beaghton AK, et al. A CRISPR–Cas9 gene drive targeting doublesex causes complete population suppression in caged Anopheles gambiae mosquitoes. Nature biotechnology. 2018;36 11:1062-6.
- 127. Simoni A, Hammond AM, Beaghton AK, Galizi R, Taxiarchi C, Kyrou K, et al. A male-biased sex-distorter gene drive for the human malaria vector Anopheles gambiae. Nature Biotechnology. 2020:1-7.
- 128. Hammond A, Galizi R, Kyrou K, Simoni A, Siniscalchi C, Katsanos D, et al. A CRISPR-Cas9 gene drive system targeting female reproduction in the malaria mosquito vector Anopheles gambiae. Nature biotechnology. 2016;34 1:78-83.
- 129. Galizi R, Doyle LA, Menichelli M, Bernardini F, Deredec A, Burt A, et al. A synthetic sex ratio distortion system for the control of the human malaria mosquito. Nature communications. 2014;5:3977.
- 130. Gantz VM, Jasinskiene N, Tatarenkova O, Fazekas A, Macias VM, Bier E, et al. Highly efficient Cas9-mediated gene drive for population modification of the malaria vector mosquito Anopheles stephensi. Proceedings of the National Academy of Sciences. 2015;112 49:E6736-E43.
- 131. Pham TB, Phong CH, Bennett JB, Hwang K, Jasinskiene N, Parker K, et al. Experimental population modification of the malaria vector mosquito, Anopheles stephensi. PLoS genetics. 2019;15 12:e1008440.
- 132. Akbari OS, Matzen KD, Marshall JM, Huang H, Ward CM, Hay BA. A synthetic gene drive system for local, reversible modification and suppression of insect populations. Current biology. 2013;23 8:671-7.
- 133. Buchman A, Marshall JM, Ostrovski D, Yang T, Akbari OS. Synthetically engineered Medea gene drive system in the worldwide crop pest Drosophila suzukii. Proceedings of the National Academy of Sciences. 2018;115 18:4725-30.
- 134. Kandul NP, Liu J, Buchman A, Gantz VM, Bier E, Akbari OS. Assessment of a split homing based gene drive for efficient knockout of multiple genes. G3: Genes, Genomes, Genetics. 2020;10 2:827-37.
- 135. Nash A, Urdaneta GM, Beaghton AK, Hoermann A, Papathanos PA, Christophides GK, et al. Integral gene drives for population replacement. Biology open. 2019;8 1.
- 136. Oberhofer G, Ivy T, Hay BA. Cleave and Rescue, a novel selfish genetic element and general strategy for gene drive. Proceedings of the National Academy of Sciences. 2019;116 13:6250-9.
- 137. Collins C, Bonds J, Quinlan M, Mumford J. Effects of the removal or reduction in density of the malaria mosquito, Anopheles gambiae sl, on interacting predators and competitors in local ecosystems. Medical and veterinary entomology. 2019;33 1:1-15.
- 138. Baker DA, Nolan T, Fischer B, Pinder A, Crisanti A, Russell S. A comprehensive gene expression atlas of sex-and tissue-specificity in the malaria vector, Anopheles gambiae. BMC genomics. 2011;12 1:1-12.
- 139. Cassone BJ, Kay RG, Daugherty MP, White BJ. Comparative transcriptomics of malaria mosquito testes: function, evolution, and linkage. G3: Genes, Genomes, Genetics. 2017;7 4:1127-36.
- 140. Papa F, Windbichler N, Waterhouse RM, Cagnetti A, D'Amato R, Persampieri T, et al. Rapid evolution of female-biased genes among four species of Anopheles malaria mosquitoes. Genome research. 2017;27 9:1536-48.
- 141. Rose G, Krzywinska E, Kim J, Revuelta L, Ferretti L, Krzywinski J. Dosage compensation in the African malaria mosquito Anopheles gambiae. Genome biology and evolution. 2016;8 2:411-25.

- 142. Taxiarchi C, Kranjc N, Kriezis A, Kyrou K, Bernardini F, Russell S, et al. High-resolution transcriptional profiling of Anopheles gambiae spermatogenesis reveals mechanisms of sex chromosome regulation. Scientific reports. 2019;9 1:1-12.
- 143. Araujo MdS, Andrade AO, Santos NACd, Pereira DB, Costa GdS, Paulo PFMd, et al. Brazil's first free-mating laboratory colony of Nyssorhynchus darlingi. Revista da Sociedade Brasileira de Medicina Tropical. 2019;52.
- 144. Moreno M, Tong C, Guzmán M, Chuquiyauri R, Llanos-Cuentas A, Rodriguez H, et al. Infection of laboratory-colonized Anopheles darlingi mosquitoes by Plasmodium vivax. The American journal of tropical medicine and hygiene. 2014;90 4:612-6.
- 145. Villarreal-Treviño C, Vásquez GM, López-Sifuentes VM, Escobedo-Vargas K, Huayanay-Repetto A, Linton Y-M, et al. Establishment of a free-mating, long-standing and highly productive laboratory colony of Anopheles darlingi from the Peruvian Amazon. Malaria journal. 2015;14 1:227.
- 146. Marinotti O, Cerqueira GC, De Almeida LGP, Ferro MIT, Loreto ELdS, Zaha A, et al. The genome of Anopheles darlingi, the main neotropical malaria vector. Nucleic acids research. 2013;41 15:7387-400.
- 147. Compton A, Sharakhov IV, Tu Z. Recent Advances and Future Perspectives in Vector-omics. Current Opinion in Insect Science. 2020.
- 148. Da Silva AN, Dos Santos CC, Lacerda RN, Santa Rosa EP, De Souza RT, Galiza D, et al. Laboratory colonization of Anopheles aquasalis (Diptera: Culicidae) in Belém, Pará, Brazil. Journal of medical entomology. 2006;43 1:107-9.
- 149. Giglio N, Sousa-Lima A, Gallardo A, Lima J. Laboratory Colonization of Anopheles (Nyssorhynchus) marajoara (Diptera: Culicidae) by Induced Copulation. Journal of medical entomology. 2015;52 1:3-8.
- 150. Horosko S, Lima JB, Brandolini M. Establishment of a free-mating colony of Anopheles albitarsis from Brazil. FIRST THINGS. 1997:95-6.
- 151. Dahmana H, Mediannikov O. Mosquito-Borne Diseases Emergence/Resurgence and How to Effectively Control It Biologically. Pathogens. 2020;9 4:310.
- 152. Poopathi S, Mani C, Thirugnanasambantham K, Praba VL, Ahangar NA, Balagangadharan K. Identification and characterization of a novel marine Bacillus cereus for mosquito control. Parasitology research. 2014;113 1:323-32.
- 153. Sharma L, Bohra N, Singh RK, Marques G. Potential of Entomopathogenic Bacteria and Fungi. Microbes for Sustainable Insect Pest Management: Springer; 2019. p. 115-49.
- 154. Soares-da-Silva J, Queirós SG, de Aguiar JS, Viana JL, dos RAV Neta M, da Silva MC, et al. Molecular characterization of the gene profile of Bacillus thuringiensis Berliner isolated from Brazilian ecosystems and showing pathogenic activity against mosquito larvae of medical importance. Acta tropica. 2017;176:197-205.
- 155. Arantes O, Vilas-Bôas L, Vilas-Bôas G. Bacillus thuringiensis: estratégias no controle biológico. Biotecnologia: avanços na agricultura e na agroindústria Caxias do Sul: Agropecuária. 2002:269-93.
- 156. Galardo AKR, Zimmerman R, Galardo CD. Larval control of Anopheles (Nyssorhinchus) darlingiusing granular formulation of Bacillus sphaericus in abandoned gold-miners excavation pools in the Brazilian Amazon Rainforest. Revista da Sociedade Brasileira de Medicina Tropical. 2013;46 2:172-7.
- 157. Fillinger U, Knols BG, Becker N. Efficacy and efficiency of new Bacillus thuringiensis var. israelensis and Bacillus sphaericus formulations against Afrotropical anophelines in Western Kenya. Tropical Medicine & International Health. 2003;8 1:37-47.
- 158. Majambere S, Pinder M, Fillinger U, Ameh D, Conway DJ, Green C, et al. Is mosquito larval source management appropriate for reducing malaria in areas of extensive flooding in The Gambia? A cross-over intervention trial. The American journal of tropical medicine and hygiene. 2010;82 2:176-84.

- 159. Afrane YA, Mweresa NG, Wanjala CL, Gilbreath III TM, Zhou G, Lee M-C, et al. Evaluation of long-lasting microbial larvicide for malaria vector control in Kenya. Malaria journal. 2016;15 1:1-9.
- 160. Derua YA, Kahindi SC, Mosha FW, Kweka EJ, Atieli HE, Wang X, et al. Microbial larvicides for mosquito control: Impact of long lasting formulations of Bacillus thuringiensis var. israelensis and Bacillus sphaericus on non-target organisms in western Kenya highlands. Ecology and evolution. 2018;8 15:7563-73.
- 161. Zhou G, Wiseman V, Atieli HE, Lee M-C, Githeko AK, Yan G. The impact of long-lasting microbial larvicides in reducing malaria transmission and clinical malaria incidence: study protocol for a cluster randomized controlled trial. Trials. 2016;17 1:423.
- 162. Johnson BJ, Manby R, Devine GJ. Performance of an aerially applied liquid Bacillus thuringiensis var. israelensis formulation (strain AM65-52) against mosquitoes in mixed saltmarsh-mangrove systems and fine-scale mapping of mangrove canopy cover using affordable drone-based imagery. Pest Management Science. 2020.
- 163. Ramirez JL, Short SM, Bahia AC, Saraiva RG, Dong Y, Kang S, et al. Chromobacterium Csp_P reduces malaria and dengue infection in vector mosquitoes and has entomopathogenic and in vitro anti-pathogen activities. PLoS pathogens. 2014;10 10:e1004398.
- 164. Short SM, Van Tol S, MacLeod HJ, Dimopoulos G. Hydrogen cyanide produced by the soil bacterium Chromobacterium sp. Panama contributes to mortality in Anopheles gambiae mosquito larvae. Scientific reports. 2018;8 1:1-13.
- 165. Valero-Jiménez CA, van Kan JA, Koenraadt CJ, Zwaan BJ, Schoustra SE. Experimental evolution to increase the efficacy of the entomopathogenic fungus Beauveria bassiana against malaria mosquitoes: Effects on mycelial growth and virulence. Evolutionary Applications. 2017;10 5:433-43.
- 166. Lovett B, Bilgo E, Diabate A, St. Leger R. A review of progress toward field application of transgenic mosquitocidal entomopathogenic fungi. Pest management science. 2019;75 9:2316-24.
- 167. Azizoglu U, Jouzani GS, Yilmaz N, Baz E, Ozkok D. Genetically modified entomopathogenic bacteria, recent developments, benefits and impacts: A review. Science of The Total Environment. 2020:139169.
- 168. Federici BA. Recombinant bacterial larvicides for control of important mosquito vectors of disease. Vector Biology, Ecology and Control: Springer; 2010. p. 163-76.
- 169. Federici BA, Park H-W, Bideshi DK, Wirth MC, Johnson JJ, Sakano Y, et al. Developing recombinant bacteria for control of mosquito larvae. Journal of the American Mosquito Control Association. 2007;23 sp2:164-75.
- 170. Borovsky D, Nauwelaers S, Shatters Jr R. Biochemical and Molecular Characterization of Pichia pastoris Cells Expressing Multiple TMOF Genes (tmfA) for Mosquito Larval Control. Frontiers in Physiology. 2020;11:527.
- 171. Deng S-Q, Zou W-H, Li D-L, Chen J-T, Huang Q, Zhou L-J, et al. Expression of Bacillus thuringiensis toxin Cyt2Ba in the entomopathogenic fungus Beauveria bassiana increases its virulence towards Aedes mosquitoes. PLoS neglected tropical diseases. 2019;13 7:e0007590.
- 172. Karabörklü S, Azizoglu U, Azizoglu ZB. Recombinant entomopathogenic agents: a review of biotechnological approaches to pest insect control. World Journal of Microbiology and Biotechnology. 2018;34 1:14.
- 173. Cirimotich CM, Dong Y, Clayton AM, Sandiford SL, Souza-Neto JA, Mulenga M, et al. Natural microbe-mediated refractoriness to Plasmodium infection in Anopheles gambiae. Science. 2011;332 6031:855-8.
- 174. Dong Y, Manfredini F, Dimopoulos G. Implication of the mosquito midgut microbiota in the defense against malaria parasites. PLoS pathogens. 2009;5 5:e1000423.
- 175. Ramirez JL, Souza-Neto J, Cosme RT, Rovira J, Ortiz A, Pascale JM, et al. Reciprocal tripartite interactions between the Aedes aegypti midgut microbiota, innate immune system and dengue virus influences vector competence. PLoS Negl Trop Dis. 2012;6 3:e1561.
- 176. Van Tol S, Dimopoulos G. Influences of the mosquito microbiota on vector competence. Advances in insect physiology. vol. 51: Elsevier; 2016. p. 243-91.

- 177. Bai L, Wang L, Vega-Rodríguez J, Wang G, Wang S. A Gut Symbiotic Bacterium Serratia marcescens Renders Mosquito Resistance to Plasmodium Infection Through Activation of Mosquito Immune Responses. Frontiers in Microbiology. 2019;10 1580; doi: 10.3389/fmicb.2019.01580. https://www.frontiersin.org/article/10.3389/fmicb.2019.01580.
- 178. Gonzalez-Ceron L, Santillan F, Rodriguez MH, Mendez D, Hernandez-Avila JE. Bacteria in midguts of field-collected Anopheles albimanus block Plasmodium vivax sporogonic development. Journal of medical entomology. 2003;40 3:371-4.
- 179. Dennison NJ, Saraiva RG, Cirimotich CM, Mlambo G, Mongodin EF, Dimopoulos G. Functional genomic analyses of Enterobacter, Anopheles and Plasmodium reciprocal interactions that impact vector competence. Malaria journal. 2016;15 1:1-15.
- 180. Cappelli A, Damiani C, Mancini MV, Valzano M, Rossi P, Serrao A, et al. Asaia activates immune genes in mosquito eliciting an anti-Plasmodium response: implications in Malaria control. Front Genet. 2019;10:836.
- 181. Cappelli A, Valzano M, Cecarini V, Bozic J, Rossi P, Mensah P, et al. Killer yeasts exert antiplasmodial activities against the malaria parasite Plasmodium berghei in the vector mosquito Anopheles stephensi and in mice. Parasit Vectors. 2019;12 1:329.
- 182. Herren JK, Mbaisi L, Mararo E, Makhulu EE, Mobegi VA, Butungi H, et al. A microsporidian impairs Plasmodium falciparum transmission in Anopheles arabiensis mosquitoes. Nature communications. 2020;11 1:1-10.
- 183. Sougoufara S, Ottih EC, Tripet F. The need for new vector control approaches targeting outdoor biting Anopheline malaria vector communities. Parasit Vectors. 2020;13 1:1-15.
- 184. Huang W, Wang S, Jacobs-Lorena M. Use of Microbiota to Fight Mosquito-Borne Disease. Front Genet. 2020;11.
- 185. Ren X, Hoiczyk E, Rasgon JL. Viral paratransgenesis in the malaria vector Anopheles gambiae. PLoS pathogens. 2008;4 8:e1000135.
- 186. Fang W, Vega-Rodríguez J, Ghosh AK, Jacobs-Lorena M, Kang A, Leger RJS. Development of transgenic fungi that kill human malaria parasites in mosquitoes. Science. 2011;331 6020:1074-7.
- 187. Mancini MV, Spaccapelo R, Damiani C, Accoti A, Tallarita M, Petraglia E, et al. Paratransgenesis to control malaria vectors: a semi-field pilot study. Parasit Vectors. 2016;9 1:1-9.
- 188. Raharimalala FN, Boukraa S, Bawin T, Boyer S, Francis F. Molecular detection of six (endo-) symbiotic bacteria in Belgian mosquitoes: first step towards the selection of appropriate paratransgenesis candidates. Parasitology research. 2016;115 4:1391-9.
- 189. Villegas LM, Pimenta PFP. Metagenomics, paratransgenesis and the Anopheles microbiome: a portrait of the geographical distribution of the anopheline microbiota based on a meta-analysis of reported taxa. Memórias do Instituto Oswaldo Cruz. 2014;109 5:672-84.
- 190. Wang S, Dos-Santos AL, Huang W, Liu KC, Oshaghi MA, Wei G, et al. Driving mosquito refractoriness to Plasmodium falciparum with engineered symbiotic bacteria. Science. 2017;357 6358:1399-402.
- 191. Wang S, Ghosh AK, Bongio N, Stebbings KA, Lampe DJ, Jacobs-Lorena M. Fighting malaria with engineered symbiotic bacteria from vector mosquitoes. Proceedings of the National Academy of Sciences. 2012;109 31:12734-9.
- 192. Yoshida S, Ioka D, Matsuoka H, Endo H, Ishii A. Bacteria expressing single-chain immunotoxin inhibit malaria parasite development in mosquitoes. Molecular and biochemical parasitology. 2001;113 1:89-96.
- 193. Arruda A, Ferreira GS, da Silva Lima NC, dos Santos Júnior A, Custódio MGF, Benevides-Matos N, et al. A simple methodology to collect culturable bacteria from feces of Anopheles darlingi (Diptera: Culicidae). Journal of microbiological methods. 2017;141:115-7.
- 194. Bascuñán P, Niño-Garcia JP, Galeano-Castañeda Y, Serre D, Correa MM. Factors shaping the gut bacterial community assembly in two main Colombian malaria vectors. Microbiome. 2018;6 1:1-12.
- 195. Galeano-Castañeda Y, Bascuñán P, Serre D, Correa MM. Trans-stadial fate of the gut bacterial microbiota in Anopheles albimanus. Acta Tropica. 2020;201:105204.

- 196. Galeano-Castañeda Y, Urrea-Aguirre P, Piedrahita S, Bascuñán P, Correa MM. Composition and structure of the culturable gut bacterial communities in Anopheles albimanus from Colombia. Plos one. 2019;14 12:e0225833.
- 197. Kämpfer P, Glaeser SP, Marinotti O, Guy L, Håkansson S, Tadei WP, et al. Coetzeea brasiliensis gen. nov., sp. nov. isolated from larvae of Anopheles darlingi. International journal of systematic and evolutionary microbiology. 2016;66 12:5211-7.
- 198. Nilsson LK, de Oliveira MR, Marinotti O, Rocha EM, Håkansson S, Tadei WP, et al. Characterization of Bacterial Communities in Breeding Waters of Anopheles darlingi in Manaus in the Amazon Basin Malaria-Endemic Area. Microbial ecology. 2019;78 4:781-91.
- 199. Oliveira TM, Sanabani SS, Sallum MAM. Asaia (Rhodospirillales: Acetobacteraceae) and Serratia (Enterobacterales: Yersiniaceae) associated with Nyssorhynchus braziliensis and Nyssorhynchus darlingi (Diptera: Culicidae). Revista Brasileira de Entomologia. 2020;64 2.
- 200. Oliveira TMP, Sanabani SS, Sallum MAM. Bacterial diversity associated with the abdomens of naturally Plasmodium-infected and non-infected Nyssorhynchus darlingi. BMC microbiology. 2020;20 1:1-8.
- 201. Prussing C, Saavedra MP, Bickersmith SA, Alava F, Guzmán M, Manrique E, et al. Malaria vector species in Amazonian Peru co-occur in larval habitats but have distinct larval microbial communities. PLoS neglected tropical diseases. 2019;13 5:e0007412.
- 202. Terenius O, De Oliveira CD, Pinheiro WD, Tadei WP, James AA, Marinotti O. 16S rRNA gene sequences from bacteria associated with adult Anopheles darlingi (Diptera: Culicidae) mosquitoes. Journal of medical entomology. 2008;45 1:172-5.
- 203. Hilgenboecker K, Hammerstein P, Schlattmann P, Telschow A, Werren JH. How many species are infected with Wolbachia?—a statistical analysis of current data. FEMS microbiology letters. 2008;281 2:215-20.
- 204. Sinkins SP. Wolbachia and cytoplasmic incompatibility in mosquitoes. Insect biochemistry and molecular biology. 2004;34 7:723-9.
- 205. Werren JH, Baldo L, Clark ME. Wolbachia: master manipulators of invertebrate biology. Nature Reviews Microbiology. 2008;6 10:741-51.
- 206. Rousset F, Bouchon D, Pintureau B, Juchault P, Solignac M. Wolbachia endosymbionts responsible for various alterations of sexuality in arthropods. Proceedings of the Royal Society of London Series B: Biological Sciences. 1992;250 1328:91-8.
- 207. Stouthamer R, Breeuwer JA, Hurst GD. Wolbachia pipientis: microbial manipulator of arthropod reproduction. Annual Reviews in Microbiology. 1999;53 1:71-102.
- 208. Yen JH, Barr AR. New hypothesis of the cause of cytoplasmic incompatibility in Culex pipiens L. Nature. 1971;232 5313:657-8.
- 209. Caragata EP, Dutra HL, Moreira LA. Exploiting intimate relationships: controlling mosquito-transmitted disease with Wolbachia. Trends in parasitology. 2016;32 3:207-18.
- 210. Hughes GL, Koga R, Xue P, Fukatsu T, Rasgon JL. Wolbachia infections are virulent and inhibit the human malaria parasite Plasmodium falciparum in Anopheles gambiae. PLoS pathogens. 2011;7 5:e1002043; doi: 10.1371/journal.ppat.1002043.
- 211. Ferreira AG, Fairlie S, Moreira LA. Insect vectors endosymbionts as solutions against diseases. Curr Opin Insect Sci. 2020;40:56-61; doi: 10.1016/j.cois.2020.05.014.
- 212. Baldini F, Segata N, Pompon J, Marcenac P, Shaw WR, Dabiré RK, et al. Evidence of natural Wolbachia infections in field populations of Anopheles gambiae. Nature communications. 2014;5:3985.
- 213. Gomes FM, Hixson BL, Tyner MD, Ramirez JL, Canepa GE, e Silva TLA, et al. Effect of naturally occurring Wolbachia in Anopheles gambiae sl mosquitoes from Mali on Plasmodium falciparum malaria transmission. Proceedings of the National Academy of Sciences. 2017;114 47:12566-71.
- 214. Jeffries CL, Lawrence GG, Golovko G, Kristan M, Orsborne J, Spence K, et al. Novel Wolbachia strains in Anopheles malaria vectors from sub-Saharan Africa. Wellcome open research. 2018;3.
- 215. Shaw WR, Marcenac P, Childs LM, Buckee CO, Baldini F, Sawadogo SP, et al. Wolbachia infections in natural Anopheles populations affect egg laying and negatively correlate with Plasmodium development. Nature communications. 2016;7 1:1-7.

- 216. Chrostek E, Gerth M. Is Anopheles gambiae a natural host of Wolbachia? MBio. 2019;10 3.
- 217. Joshi D, Pan X, McFadden MJ, Bevins D, Liang X, Lu P, et al. The maternally inheritable Wolbachia wAlbB induces refractoriness to Plasmodium berghei in Anopheles stephensi. Frontiers in microbiology. 2017;8:366.
- 218. Bian G, Joshi D, Dong Y, Lu P, Zhou G, Pan X, et al. Wolbachia invades Anopheles stephensi populations and induces refractoriness to Plasmodium infection. Science. 2013;340 6133:748-51.
- 219. Epis S, Varotto-Boccazzi I, Crotti E, Damiani C, Giovati L, Mandrioli M, et al. Chimeric symbionts expressing a Wolbachia protein stimulate mosquito immunity and inhibit filarial parasite development. Communications biology. 2020;3 1:1-10.
- 220. GA U. Transforming our world: the 2030 Agenda for Sustainable Development. . Division for Sustainable Development Goals: New York, NY, USA 2015.
- 221. Akhavan D, Musgrove P, Abrantes A, Gusmão RdA. Cost-effective malaria control in Brazil: cost-effectiveness of a malaria control program in the Amazon Basin of Brazil, 1988–1996. Social science & medicine. 1999;49 10:1385-99.
- 222. Bôtto-Menezes C, Bardají A, dos Santos Campos G, Fernandes S, Hanson K, Martínez-Espinosa FE, et al. Costs associated with malaria in pregnancy in the Brazilian Amazon, a low endemic area where Plasmodium vivax predominates. PLoS neglected tropical diseases. 2016;10 3:e0004494.
- 223. Dev V, Manguin S. Fast forward: from malaria elimination to malaria eradication. 2020. https://blogs.biomedcentral.com/bugbitten/2020/06/16/fast-forward-from-malaria-elimination-to-malaria-eradication/.
- 224. Pang LW, Piovesan-Alves F. Economic advantage of a community-based malaria management program in the Brazilian Amazon. The American journal of tropical medicine and hygiene. 2001;65 6:883-6.
- 225. Shretta R, Avanceña AL, Hatefi A. The economics of malaria control and elimination: a systematic review. Malaria journal. 2016;15 1:593.
- 226. Huang C-y, Zhang S-y, Chen Z-y, Xie H-g, Yang RO. Cost-benefit analysis of malaria elimination phase surveillance measures in Fujian Province. 2020.