Article

Terminalia bentzoë, a Mascarene endemic plant, inhibits human hepatocellular carcinoma cells growth *in vitro* via G0/G1 phase cell cycle arrest

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Abstract: Tropical forests constitute prolific sanctuary of unique floral diversity and potential medicinal sources, however, many of them remain unexplored. The scarcity of rigorous scientific data on the surviving Mascarene endemic taxa renders bioprospecting of this untapped resource of utmost importance. Thus, in view of valorising the native resource, this study has as objective to investigate the bioactivities of endemic leaf extracts. Herein, seven Mascarene endemic plants leaves were extracted and evaluated for their *in vitro* antioxidant properties and antiproliferative effects on a panel of cancer cell lines using MTT and clonogenic cell survival assay. Flow cytometry and comet assay were used to investigate the cell cycle and DNA damaging effects, respectively. Bioassay guided-fractionation coupled with LC-Mass spectrometry (MS), gas chromatography-MS, and nuclear magnetic resonance spectroscopy analysis were used to identify the bioactive compounds. Among the seven plants tested, *Terminalia bentzoë* was comparatively the most potent antioxidant extract with significantly ($p < 0.05$) higher cytotoxic activities. *T. bentzoë* extract further selectively suppressed the growth of human hepatocellular carcinoma cells and significantly halted the cell cycle progression in G0/G1 phase, decreased the cells replicative potential and induced significant DNA damage. Ten phenolic compounds including punicalagin and ellagic acid were identified and likely contributed to the extract potent antioxidant and cytotoxic activities. These results established a promising basis for further in-depth investigations on the potential use of *T. bentzoë* as supportive therapy in cancer management.

Keywords: *Terminalia bentzoë*; Mascarene endemic; cytotoxicity; antioxidant; cell cycle arrest; phenolics; bioassay-guided fractionation

1. Introduction

The plant kingdom is known to be a prolific sanctuary of phytochemicals with unique therapeutic potential. At least 25 % of the 1562 clinical drugs approved by the US Food and Drugs Administration are known to have been emanated from terrestrial plants [1,2]. Moreover, an estimate of about 28187 plant taxa, globally, are documented to have medicinal values with over 3000 species reported with the ethnomedicinal application against cancer [3,4]. The continued dependence of

mankind on plants was further evidenced during the recent outbreak of the COVID-19 pandemic whereby, herbal medicines were used in an attempt to mitigate the symptoms of the novel coronavirus infection [5–7].

Madagascar and its neighboring islands in the Western Indian Ocean regions are known as biodiversity hotspots [8]. Undoubtedly, untapped endemic plant species from these niche areas broadened the structural variation of novel chemotypes [9,10]. The tropical forest of Madagascar was once acknowledged as a fertile source of economically valuable plants with pharmacologically active ingredients [11]. Indeed, the anticancer drugs vinblastine and vincristine were derived from the Madagascan endemic *Catharanthus roseus* (Apocynaceae) [12]. Certainly, the market value of vincristine alone was estimated to be 15 million USD per kilogram, in the year 2016 [13].

Phytogeography investigations revealed that the Mascarene endemic plants islands have their ancestral lineages traced back from Madagascar [14]. As such, the unique floral biodiversity of Mauritius is expected to possess similar medicinal and therapeutic prolificacy as the Madagascan rain forest. However, instead of conserving such valuable biodiversity, human activities are pushing endemic taxa towards an unprecedented extinction crisis. In less than 400 years of human settlement, Mauritius has witnessed the shrinking of its native forest to around 5 % of the original cover, leading to the permanent loss of 30 (10.9 %) of its endemic plant species and driving 81.7 % of the remnant endemic taxa on the brink of extinction [15–17]. For instance, fewer than 500 adults trees of *Terminalia bentzoë* (L.) L.f. subsp. *bentzoë* is recorded in the wild, defining the species survival as vulnerable, as per the IUCN red list criteria [18]. Nevertheless, the remnant areas of the pristine forest are still home to a plethora of endemic flora rich in high genetic diversity representing interesting sources for complementary and alternative medicine, nutraceuticals as well as pharmaceutical leads [16].

Initiation and progression of cancer are involved in oxidative stress via DNA damage and increase of DNA mutations. Conventional chemo- and radiotherapy caused cancer cell death often through generation of reactive oxygen species (ROS), but also unfortunately leading to severe side effects. It is highly desired to develop more effective therapies with less toxic effects [19]. Plant polyphenols exert anticarcinogenic activity by interfering with the different hallmarks of cancer, including sustained tumor cell proliferation, angiogenesis and apoptotic cell death through various signaling pathways [19,20]. Polyphenolics can also behave as antioxidants thereby maintaining the integrity of DNA from oxidative stress attack and prevent the initiation stage of carcinogenesis [21,22].

The tropical island of Mauritius is known for its endemic biodiversity richness [8]. However, human activities on the island have provoked the irreversible loss of a considerable fraction of this genetic resource. For instance, in less than four centuries, Mauritius has lost 95 % of its pristine forest cover accounting for the extinction of 10.9 % of the island endemic flora [23]. The scarcity of rigorous scientific data on the surviving endemic taxa renders bioprospecting of this untapped resource of utmost importance. With this in mind, and in view of providing solid foundations to enforce stringent conservation policies, the *in vitro* antioxidant propensities of leaf extracts from seven plants endemic to Mascarene islands were investigated, in relation to their polyphenolic content. The plants under study have documented traditional uses against ailments ranging from dermatological conditions, asthma to infectious diseases (**Table 1**). The cytotoxic effect of *Terminalia bentzoë*, on a panel of cancer cell lines, and its ability to impede the cell cycle progression in hepatocellular carcinoma (HepG2) cells were determined. The bioactive constituents in *T. bentzoë* leaf extract were characterised following bioassay-guided fractionation.

2. Materials and Methods

2.1. Plant material and preparation of total extracts.

Healthy fresh leaves of seven Mascarene endemic plants were collected in Mauritius and deposited at the Mauritius herbarium, where plant species were authenticated by the botanist (**Table 1**). The leaves were air-dried followed by exhaustive maceration with aqueous methanol (80 %, v/v) and freeze-dried as described previously [24]. The assay results were expressed in terms of the lyophilised weight of extracts.

2.2. Estimation of polyphenolic contents.

The total phenolic, flavonoid, and proanthocyanidin level in the crude extracts were estimated using the Folin-Ciocalteu assay, aluminium chloride assay and HCl/Butan-1-ol assay as described [24].

2.3. In-vitro antioxidant capacities of extracts.

The antioxidant potential of the extracts was investigated according to reported methods [24,25]. Extract vehicle and gallic acid (or otherwise stated) were used as negative and positive controls respectively. The percentage activity of the extracts was calculated relative to the negative control. GraphPad Prism 6 software (GraphPad Inc., USA) was used to plot the dose-response curves and to generate the half-maximal inhibitory concentration (IC_{50}) values. All experiments were performed in triplicates in three independent assays. The results were expressed as mean ± SEM.

2.3.1. Ferric reducing antioxidant potential assay

The final reaction volume of 3.4 mL of the ferric reducing antioxidant power assay contained 100 µL of extract and 300 µL of water followed by addition of 3 mL FRAP reagent. The FRAP reagent was prepared immediately before use by mixing 100 mL of 0.25 M acetate buffer (pH 3.6), 10 mL of 20 mM ferric chloride (source of Fe^{3+}) and 10 mL of 10 mM 2,4,6-tripyridyl-s-triazine. After incubating the mixture for 4 minutes at ambient temperature, the absorbance was read at 593 nm against a blank. Results were reported as in mmol Fe^{2+}.

2.3.2. 2,2-diphenyl-1-picrylhydrazyl (DPPH) assay

The DPPH assay protocol involved mixing varying concentrations, between 0 to 25 µg/mL, of 100 µL of the methanolic extract with 200 µL of 100 µM methanolic DPPH and absorbance were read at 492 nm, 30 minutes post-incubation at ambient temperature.

2.3.3. Iron chelating assay

The reaction mixture for iron-chelating activity contained 40 µL of plant extract (concentrations between 0 to 10 mg/mL), 10 µL of $FeCl_2 \cdot 4H_2O$ (0.5 mM) and 150 µL of distilled deionized water. The mixture was incubated at ambient temperature for 5 minutes, before the addition of 10 µL of ferrozine (2.5 mM) and the absorbance was read at 562 nm.

2.3.4. Superoxide scavenging assay

The final 250 µL reaction volume for superoxide anion scavenging assay contained 25 µL of plant extract (concentrations between 0 to 300 µg/mL), 100 µL of 156 µM of nitroblue tetrazolium and 100 µL of 200 µM beta-nicotinamide adenine dinucleotide reduced disodium salt hydrate and 30 µL of phenazine methosulphate. The absorbance was read at 560 nm following 30 minutes incubation at 25 ˚C.

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Table 1. The investigated Mascarene endemic plant species.

Species	Family	Vernacular names	Ethnomedicinal uses [16]	Collection site	Collection date	Mauritius herbarium accession code	% Yield
***Antirhea borbonica* J.F.Gmel**	Rubiaceae	Bois lousteau, Bois d'oiseau	Astringent, diarrhoea, dysentery, stop bleeding, promote wound repair, skin diseases, tambave, Urinary tract infections	Gaulettes Serrées	14-Oct-2014	MAU 0009462	5.53
***Dictyosperma album* (Bory) H. Wendl. & Drude ex Scheff var. *conjugatum* H. E. Moore & Guého**	Arecaceae	Palmiste blanc	Not described	Réduit, Joseph Guého Arboretum	19-Aug-2014	MAU 0016674	8.52
***Erythroxylum sideroxyloides* Lam**	Erythroxylaceae	Bois de ronde	Renal stones	Lower Gorges National Park, 'Morne Sec'	15-Oct-2014	MAU 0016542	13.81
***Ficus mauritiana* Lam**	Moraceae	Figuier du pays	Not described	Gaulettes Serrées	14-Nov-2014	MAU 0011002	3.10
***Hancea integrifolia* (Willd.) S.E.C. Sierra, Kulju & Welzen**	Euphorbiaceae	Bois pigeon	Clean the blood and improve blood circulation, tonic.	Gaulettes Serrées	14-Nov-2014	MAU 0016431	10.42
***Stillingia lineata* Muell. Arg**	Euphorbiaceae	Fangame; Bois de lait; Tanguin de pays	Eczema, skin disease	Lower Gorges National Park, 'Morne Sec'	27-Nov-2014	MAU 0016545	6.28
Terminalia bentzoë* (L.) L.f. subsp. *bentzoë	Combretaceae	Bois benjoin	Asthma, antipyretic, antimalarial, chills, dysentery, diarrhoea, depurative, emmenagogue, haemorrhages, Sexually transmissible diseases	Réduit, Joseph Guého Arboretum	7-Oct-2014	MAU 0016557	7.29

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2.3.5. Nitric oxide scavenging assay

The nitric oxide radical scavenging activity was conducted in a 96-well plate. 50 µL of aqueous extract (0 to 100 µg/mL) and 100 µL of 5 mM of sodium nitroprusside (in phosphate saline buffer, pH 7.4) was incubated at 25 ˚C for 150 minutes. After incubation, 125 µL of the reaction mixture was transferred to another 96-well plate, to which 100 µL of 0.33 % sulfanilic acid in 20 % glacial acetic acid was added. After 5 minutes, 100 µL of 0.1 % of N-1-napthyethylenediamine dihydrochloride was added and the pink coloration formed was read at 546 nm.

2.3.6. Deoxyribose degradation inhibitory assay

The deoxyribose degradation inhibitory assay protocol was optimised to a 24-well microtiter plate format. Each well contained 50 µL of aqueous extract, 50 µL of 1 mM EDTA, 100 µL of 500 µM $FeCl_3$, 50 µL of 1 mM ascorbic acid, 50 µL of 10 mM hydrogen peroxide, 100 µL of 100 mM KH_2PO_4-KOH buffer (pH 7.4) and 100 µL of 15 mM 2-deoxyribose. The reaction mixture was incubated at 37 ˚C for 90 minutes. At the end of the incubation period, 500 µL of 10 % (w/v) trichloroacetic acid followed by 500 µL of 1% (w/v) thiobarbituric acid were added to each well and the solutions were heated in a water bath at 80 ˚C for 20 minutes to develop the pink chromogen. The absorbance of the reaction mixture was read at 532 nm, both before and after incubation. Results were given in mg lyophilised extract/mL.

2.4. Human cell lines and culture conditions

Human liposarcoma cells (SW872), human lung carcinoma cells (A549), human hepatocellular carcinoma cells (HepG2), and human ovarian carcinoma cell lines OVCAR-4 and OVCAR-8 were purchased from American Type Culture Collection (USA). Human ovarian epithelial (HOE) cells immortalized using SV40 large T antigen was obtained from Applied Biological Materials Inc (Canada). All cell lines, except the ovarian cell lines, were cultured in Dulbecco's Modified Eagle's Medium. Roswell Park Memorial Institute (RPMI) 1640 medium was used in the case of ovarian cell lines. Culture medium was supplemented with 10% fetal bovine serum, 2 mM L-glutamine and 100 U/L streptomycin-penicillin. Cells were grown in a humidified atmosphere of 5 % carbon dioxide and 95 % humidity at 37 ˚C.

2.5. Cell-based assays

The viability of the investigated cells treated with test samples was evaluated using the methyl thiazolyl diphenyl-tetrazolium bromide (MTT) cell viability assay. Following overnight acclimatisation of cells in 96-well plate, cells were treated with test samples for 48 hours and assayed for different parameters. For 96-well plate, the seeding densities for cancer cell lines and HOE cells were 5×10^3 cells and 2×10^3 cells per well, respectively. All experiments were performed in triplicates (unless otherwise specified) in three independent assays. To compare the chemotherapeutic potential of the active extracts, the cytotoxicity of the extracts/purified compounds was evaluated against, etoposide, a clinically used oncologic agent.

2.5.1. MTT viability assay

The MTT assay was performed as previously described [26]. The percentage cell viability relative to DMSO control (0.025 % v/v, final concentration) was calculated and the IC_{50} value determined using GraphPad Prism 6 software (GraphPad Inc., USA).

2.5.2. Clonogenic cell survival assay

The effect of *T. bentzoë* leaf extract on the cell reproductive death was assessed by clonogenic cell survival assay according to reported methods, with slight modifications [27,28]. HepG2 cells were seeded in a 6-well plate (500/well) and allowed to attach overnight. Following 48 hours treatment period with extract/ control, the media was replaced with fresh complete culture media and cells were grown under standard recommended culture conditions for an additional 7 days to allow large

colonies formation. Colonies were then fixed with 4 % paraformaldehyde for 30 minutes and stained with 0.5 % (w/v) crystal violet. The individual wells were imaged using a digital camera and the colonies counted using ImageJ software (the US, National Institute of Health). The cytotoxic effect was expressed as the percentage of surviving colonies relative to untreated control.

2.5.3. Single cell gel electrophoresis

Comet assay was carried out according to the method described [29,30] with minor modifications. Briefly, 30 µL of pre-treated cells were mixed with 70 µL of 1 % (w/v) low melting agarose (LMA) and 40 µL of the cell-LMA mixture was placed on frosted microscope slides pre-coated with 1.5 % normal melting agarose. A coverslip was placed on top of the cell-LMA mix and allowed to solidify at 4°C for 1 hour in dark. Following solidification, the coverslip was gently slid off and the slides were immersed in pre-chilled lysis buffer (2.5 M NaCl, 0.1 M EDTA, 10 mM Tris base, 1% v/v Triton X-100 (added 30 minutes before use), pH 10, 4 °C) for 1 hour in dark. Following this period, the excess lysis solution was drained and the slides were submerged in electrophoresis buffer (0.2 M NaOH, 1 mM EDTA, pH 13, 4 °C) for 30 minutes in dark, to allow the DNA to unwind.

Electrophoresis was conducted for 30 minutes at 30 volts and 350 mA. The gels were neutralized by immersing in pre-chilled neutralisation buffer (0.4 Tris-HCL, pH 7.5, 4 °C) for 10 minutes in dark. The slides were washed in distilled water, fixed with 4 % formalin solution for 20 minutes and allowed to air-dry overnight. The slides were stained with Hoechst 33342 (1 µg/mL), air-dried in dark and visualised at 200 X magnification in DAPI, using EVOS fluorescence microscope (Life Technologies). Damaged DNA was measured for 100 randomly selected cells (for each independent experiment) using the Comet Assay IV 4.3.1 (Perceptive instrument, UK).

2.5.4. Flow cytometric analysis

Apoptosis/necrosis analysis was performed on HepG2 cells after 48 hours treatment by flow cytometry (Beckman Coulter's CytoFLEX and Cytexpert software) using Annexin V-FITC and Propidium iodide (PI) double staining as described in a previous study [31]. Cell cycle analysis was performed using propidium iodide for DNA staining as described [32]. Percentage of cells in different phases (G_O/G_1, S and G_2M phases) were quantified from propidium iodide fluorescence intensity-area (PI-A) histograms corresponding to the DNA content of HepG2 cells.

2.5.5. MTT-guided fractionation and identification of bioactive molecules

The total extract of *T. bentzoë* was solubilised in distilled water and sequentially partitioned with ethyl acetate, followed by n-butanol. Each fraction was dried and their cytotoxicity evaluated against SW872, A549, HepG2 using the MTT cell viability assay. The butanol fraction being selective towards HepG2 cells was subjected to Sephadex LH-20 column chromatography (30 cm X 2.1 cm internal diameter) and eluted with water, water: methanol (3:1 v/v, 1:1 v/v and 1:3 v/v respectively), methanol and acetone. The flow rate was maintained at 1.5 mL/min. Guided by the cytotoxicity against HepG2 cells and purity profile, potent sub-fraction was further fractionated using semi-preparative HPLC column. The crude extract and thereof derived purified fractions were analysed by extensive spectroscopic methods including GC-MS, LC-MS, HPLC, ^{1}H-NMR and ^{13}C-NMR (Supporting Information). For analytical HPLC, the concentration of standards in the crude extract was determined from the linear regression of the analytical standards curve namely $y = 12.317\ x$, $R^2 = 0.9996$: gallic acid and $y = 16.066\ x$, $R^2 = 0.9992$: methyl gallate.

2.6. Statistical analysis

Statistical analyses were performed using GraphPad Prism 6 software (GraphPad Inc., San Diego, California). The mean values among extracts were compared using One-Way ANOVA. Student *t*-test and/or Tukey's multiple comparisons as Post Hoc test was used to determine significances in mean phytochemicals, antioxidants and cytotoxic activities among different species. All charts were generated using GraphPad Prism 6 software (GraphPad Inc., San Diego, California).

3. Results

3.1. Estimation of polyphenols level in the investigated leaf extracts.

The phenolic content varied significantly among the seven leaf extracts under study ($p < 0.05$) with amounts ranging between 70.2 ± 4.72 mg and 385 ± 24.1 mg gallic acid equivalent/g. Total flavonoid levels ranged between 2.43 ± 0.06 mg and 12.9 ± 0.45 mg quercetin equivalent/g. Based on the spectrophotometric assay results, the estimated level of phenolics and flavonoids were significantly highest ($p < 0.05$) in *T. bentzoë* leaf extract as compared to the other investigated leaf extracts (**Table 2**). While the proanthocyanidin content prevailed in *E. sideroxyloides* leaf, both *S. lineata* leaf and *T. bentzoë* leaf had a remarkably negligible amount of proanthocyanidin detected by the butanol/HCl assay.

3.2. In-vitro antioxidant activities of the investigated leaf extracts.

An array of five analytical models was used to benchmark the antioxidant potential of the investigated leaf extracts. The Mascarene endemic plant leaf extracts exhibited a varying degree of activities in the different antioxidant assays. All extracts showed a dose-dependent metal chelating and free radical scavenging activity (**Table 2)**. In terms of iron chelation, all the extracts were weak chelator compared to EDTA with an IC_{50} value of 0.01 ± 0.00 mg/mL (23.6 ± 0.22 μM) ($p \leq 0.05$). As depicted in **Table 2**, among the seven accessions of plants, *T. bentzoë* showed the most effective antioxidant potential in all the five antioxidant assays. Thus, the free radical quenching activity of *T. bentzoë* was further evaluated in hydroxyl radical scavenging assay. *T. bentzoë* (IC_{50} = 0.25 ± 0.03 mg/mL) exhibited a significantly ($p < 0.0001$) greater degree of protection against Fenton-mediated oxidative damage to 2-deoxyribose sugar moiety as compared to gallic acid (IC_{50} = 1.65 ± 0.09 mg/mL).

3.3. Effect of T. bentzoë leaf extract on cell survival

To assess the antiproliferative properties of *T. bentzoë* leaf extract on cancer cells, first, the influence of the extracts on the cell viability of five cancer cell lines notably, SW872, A549, HepG2, Ovcar-4 and Ovcar-8 cells, were investigated using the MTT assay. *T. bentzoë* suppressed the growth of all cancer cell lines in a dose-dependent manner. However, the dose of extract required to reach the half-maximal inhibitory concentration differed considerably among the various cancer cell types. The cytotoxicity of *T. bentzoë* was also evaluated against the non-malignant human ovarian epithelial (HOE) cells and the IC_{50} value obtained (**Table 3**). HepG2 cells were more sensitive to *T. bentzoë* treatment (selective index value 2.4 compared to HOE cells) (Table 3). At 48 hours of exposure, the highest concentration of *T. bentzoë* extract (100 μg/mL) reduced HepG2 cell viability to 20 % as compared to untreated control cells (**Supplementary figure S1 A**). Subsequently, the effect of *T. bentzoë* extract on the replicative ability of HepG2 cells was evaluated using the clonogenic cell survival assay. *T. bentzoë* treatment significantly ($\rho < 0.0001$) reduced the number of surviving HepG2 colonies as compared to untreated control cells (**Figure 1**).

Table 2. Phenolic content and antioxidant potential of investigated leaf extracts.

Extract	Total phenolics[1]	Total flavonoids[2]	Total proanthocyanidins[3]	FRAP[4]	Iron chelating activity[5]	DPPH Scavenging activity[6]	Superoxide scavenging activity[6]	Nitric oxide scavenging activity[6]
A. borbonica	70.2 ± 4.72^{e}	3.15 ± 0.07^{d,e}	5.71 ± 0.09^{e}	3.32 ± 0.16^{d,e,****}	4.05 ± 0.26^{b,c,****}	11.2 ± 1.63^{d,****}	19.0 ± 2.46^{d,****}	80.3 ± 29.0^{b,**}
D. album	75.7 ± 5.22^{d}	2.43 ± 0.06^{f}	30.9 ± 0.58^{c}	2.21 ± 0.05^{e,****}	4.83 ± 1.49^{c,d,****}	7.89 ± 0.13^{c,****}	32.7 ± 1.16^{f,****}	68.0 ± 10.0^{a,*}
E. sideroxyloides	182 ± 10.5^{b}	3.62 ± 0.13^{d}	121 ± 3.25^{a}	9.14 ± 0.85^{b,****}	1.46 ± 0.02^{a,****}	4.44 ± 0.26^{b,****}	12.7 ± 0.65^{c,****}	24.3 ± 1.39^{a}
F. mauritiana	133 ± 2.36^{c}	10.4 ± 0.21^{b}	85.7 ± 2.38^{b}	5.38 ± 0.01^{c,****}	0.43 ± 0.01^{a,****}	5.35 ± 0.23^{b,****}	24.2 ± 0.44^{e,****}	87.08 ± 28.9^{b,***}
H. integrifolia	142 ± 4.91^{c}	2.75 ± 0.06^{e,f}	18.4 ± 0.78^{d}	9.37 ± 0.29^{b,****}	0.78 ± 0.01^{a,****}	4.16 ± 0.16^{b,****}	9.55 ± 0.78^{b,***}	68.49 ± 37.5^{a,b,**}
S. lineata	97.7 ± 3.36^{d}	6.61 ± 0.19^{c}	ND	4.53 ± 0.02^{c,d,****}	6.45 ± 0.02^{d,*}	4.04 ± 0.17^{a,b,****}	3.81 ± 0.48^{a}	68.5 ± 42.9^{b,**}
T. bentzoë	385 ± 24.1^{a}	12.9 ± 0.45^{a}	ND	18.2 ± 0.01^{a,****}	0.10 ± 0.00^{a,****}	2.65 ± 0.14^{a,***}	5.20 ± 0.53^{a}	9.74 ± 3.13^{a}
Gallic acid	-	-	-	24.8 ± 0.22	8.00 ± 0.04 (47.0 ± 0.23 mM)	0.62 ± 0.05 (4.18 ± 0.32 μM)	5.52 ± 0.11 (31.4 ± 0.84 μM)	9.61 ± 1.75 (68.0 ± 13.9 μM)

[1]Values are expressed as mg of gallic acid equivalent (GAE)/g, [2]values are expressed as mg of quercetin equivalent (QE)/g; [3]values are expressed as mg of cyanidin chloride equivalent (CCE)/g; [4]Values are expressed in mmol Fe^{2+}; [5]IC_{50} values are expressed in mg/ml; [6]IC_{50} values are expressed in μg/ml; Data represent mean ± standard error of mean (n=3). ND = Not detected. Different letters between rows in each column represent significant differences between extracts ($p < 0.05$). Asterisks represent significant differences between extracts and gallic acid (positive control), * $p \leq 0.05$, ** $p \leq 0.01$, *** $p \leq 0.001$, **** $p \leq 0.0001$.

Table 3. Cytotoxicity (IC_{50} μg/ml) of *T. bentzoë* against human cancer cell lines. .

Extracts	SW872	A549	HepG2	Ovcar-4	Ovcar-8	HOE
T. bentzoë	45.4 ± 1.8****	96.8 ± 4.9****	22.8 ± 1.3****	30.1 ± 2.3	38.5 ± 4.2	55.5 ± 9.1
Etoposide	2.5 ± 0.2	6.8 ± 0.7	1.7 ± 0.2	NA	NA	1.5 ± 0.1

Data represent mean calculated IC_{50} values with a standard error of the mean (n=3). NA= Etoposide at 1 μg/ml. Inhibited above 80 % cancer cell growth, indicating a much lower concentration is required for determination of IC_{50} value. Asterisks represent significant differences between *T. bentzoë* and etoposide (positive control), **** $p \leq 0.0001$.

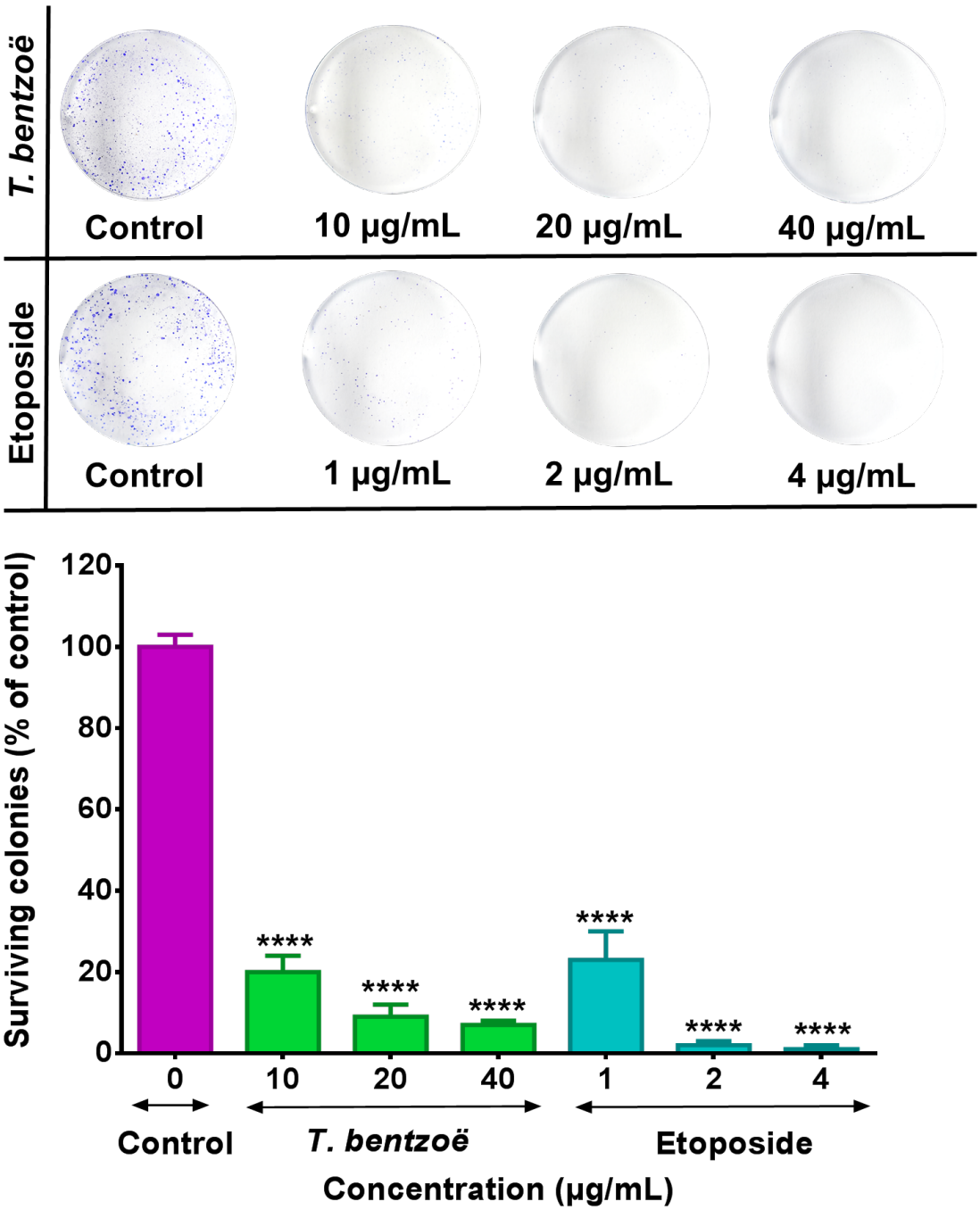


Figure 1. HepG2 cells were treated with indicated concentrations (µg/mL) of test extracts for 48 hours and subsequently allowed to grow into colonies for 14 days. After 14 days, the colonies were stained with 0.1 % crystal violet and the images of the wells were captured using a digital camera. The colonies were counted using Image J software. Each experiment was performed three times. Asterisks represent significant differences between extracts treatments and untreated control, **** $p \leq 0.0001$.

3.3. Genotoxic effect of T. bentzoë extract in HepG2 cells.

DNA damage induced by *T. bentzoë* leaf extract was assessed in HepG2 cells by the alkaline comet assay. Treatment with 10 µg/mL *T. bentzoë* leaf extract for 24 hours resulted in the induction of significant ($p \leq 0.0001$) DNA damage in HepG2 cells as compared to untreated control cells. The occurrence of DNA damaged was scored in terms of tail length, tail intensity and olive tail moment of cell cultures by comet assay software (**Table 4)**. The increased olive tail moment in response to *T.*

bentzoë treatment was 4-fold higher than that of untreated control cells. Cells treated with 200 µM H_2O_2 for 30 minutes, was used as a positive control.

Table 4. Tail length, tail intensity and olive tail moment of H_2O_2 and *T. bentzoë* treated HepG2 cells.

Extracts	Tail length (µm)	Tail intensity	Olive tail moment
Negative control (Culture medium)	32.8 ± 0.3	0.8 ± 0.1	0.2 ± 0.0
Terminalia bentzoë	38.5 ± 0.4****	3.7 ± 0.2****	0.8 ± 0.0****
Positive control (200 µM H_2O_2)	90.8 ± 7.2****	46.8 ± 2.6****	14.1 ± 0.6****

Asterisks represent significant differences between extracts and untreated control, **** $p \leq 0.0001$.

3.4. T. bentzoë induced cell death.

The proportion and distribution of HepG2 cells stained by annexin V-FITC and PI after 48 hours of exposure to *T. bentzoë* extract are illustrated in **figure 2A** and **supplementary figure S2**. The results indicated that, at a concentration equivalent to 10 µg/mL and 20 µg/mL, the distribution pattern of the cell populations in the different quadrants (**supplementary figure S2)** is similar to that of DMSO control. However, exposure of HepG2 cells for 48 hours at 40 µg/mL, stimulated a significant increase in annexin V (1.41 fold, $p < 0.01$) and propidium iodide (1.52 fold, $p < 0.001$) fluorescence compared to the control. This increase is notably reflected in a higher percentage of necrotic cells necrotic cells (11.7%, $p < 0.01$) compared to the DMSO control (4.33%), indicating apoptotic/ necrotic cell death at this test concentration. Etoposide significantly ($p < 0.001$) increased the proportion of cells undergoing apoptosis (between 10 and 12 %)/ necrosis (between 16 and 21 %) at all three test doses, relative to DMSO control.

3.5. Cell cycle progression and T. bentzoë.

Flow cytometric analysis of DNA content, of HepG2 cells treated with test extracts for 48 hours, allowed the determination of the percentage of the cell population in each phase of the cell cycle. As emphasised in **figure 2B** and **supplementary figure S3**, exposure of HepG2 cells to 40 µg/mL of *T. bentzoë* induced G0/G1 cell cycle arrest by increasing the cell population in G0/G1 (until 73.1 ± 1.76 %, $p < 0.01$ *vs.* ctrl) to the detriment of the G2/M phase (until 17.92 ± 1.76 %, $p < 0.001$ *vs.* ctrl). In contrast, treatment with 4 µg/mL etoposide led to the accumulation of G2/M cell fraction (until 53.6 ± 5.55 %) to the detriment of G0/G1 cells fraction (42.2 ± 6.25 %) suggesting G2/M arrest of HepG2 cell progression.

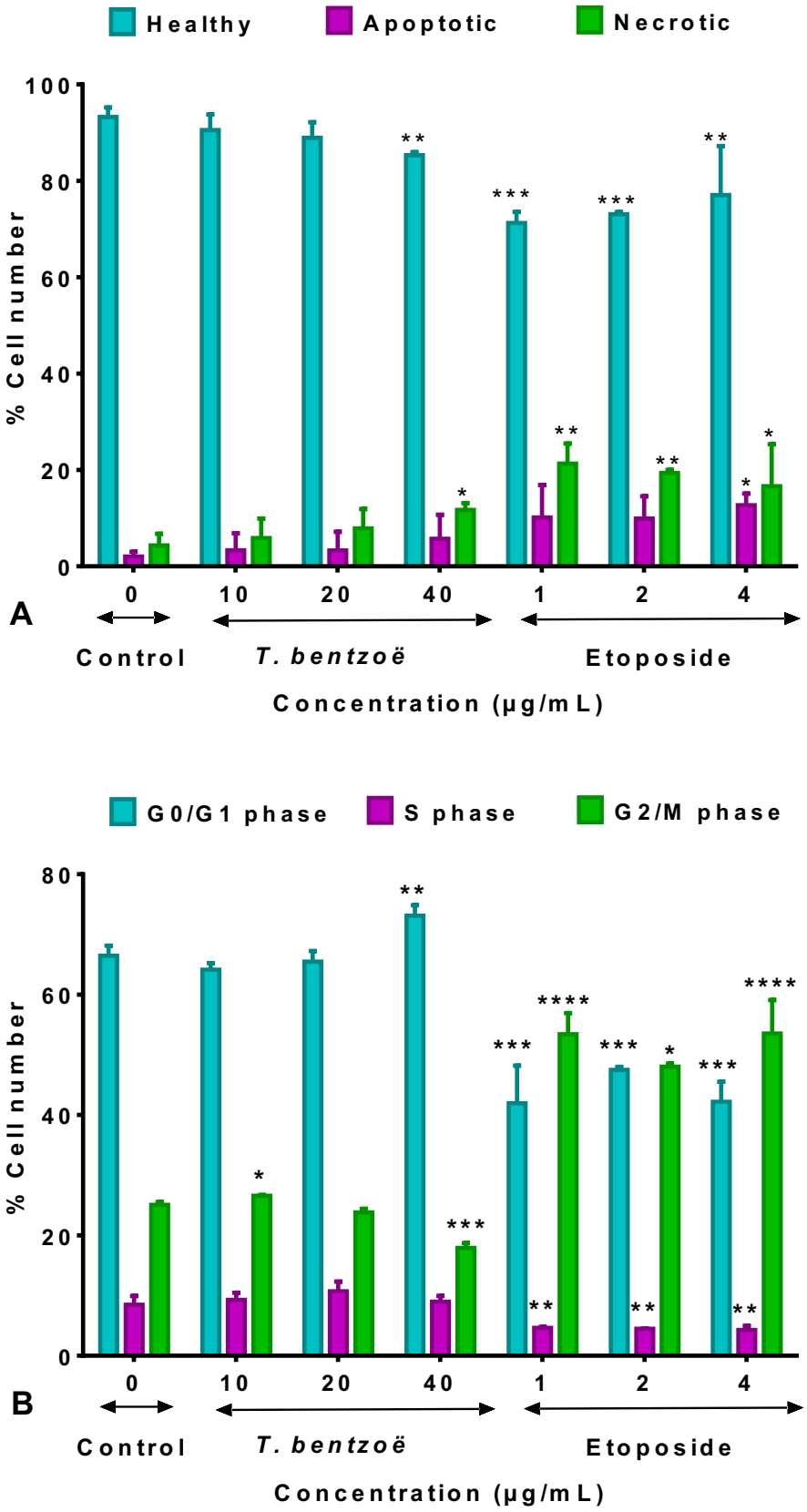


Figure 2. Effect of *Terminalia bentzoë* and etoposide on HepG2 cells as analysed by flow cytometry. (A) Annexin V-FITC/PI staining of HepG2 cells after 48 hours treatment and (B) cell cycle progression. Apoptosis and necrosis levels are expressed as percentage of the total cell population and given as mean ± SD (n=5). Percentage of cells in different phases (G0/G1, S and G2/M phases) are expressed as mean ± SD (n=3). Asterisks represent significant differences between test concentrations and DMSO control. *$p \leq 0.05$, **$p \leq 0.01$,*** $p \leq 0.001$, **** $p \leq 0.0001$.

3.5. Bioassay guided fractionation of *T. bentzoë* leaf extract

3.5.1. Effect of *T. bentzoë* fractions on HepG2 cell viability

HepG2 cells were used as a model to fractionate and characterise the bioactive components present in *T. bentzoë* leaf extract. The first round of fractionation was achieved using liquid-liquid portioning with organics solvents of increasing polarities. *T. bentzoë* butanol fraction, being most active, was further fractionated on a Sephadex LH-20 column. A total of 11 sub-fractions was derived with cytotoxic activity being limited between fractions F4 to F10 only (**Table 5**). Assessment of their chromatographic patterns revealed differences in the chemical compositions of the fractions, albeit multiple overlapping peaks highlighting the chemical complexity of the subfractions. Given the most potent cytotoxic activity of the subfraction F6 (IC_{50} = 15.2 ± 1.8 µg/mL), the further fractional separation of the latter was achieved using preparative HPLC to yield 11 HPLC subfractions (F6.1 – F6.11). Of these, HPLC subfraction F6.1 retained most of the cytotoxic activity, providing an IC_{50} value of 15.8 ± 0.3 µg/mL. By contrast, HPLC fractions F.2, F.7, F.9, F.10 and F.11 failed to effectively inhibit the growth of HepG2 cell cultures; hence no IC_{50} values were determined for these subfractions. A summary of the bioassay-guided fractionation employed is depicted in **figure 3**.

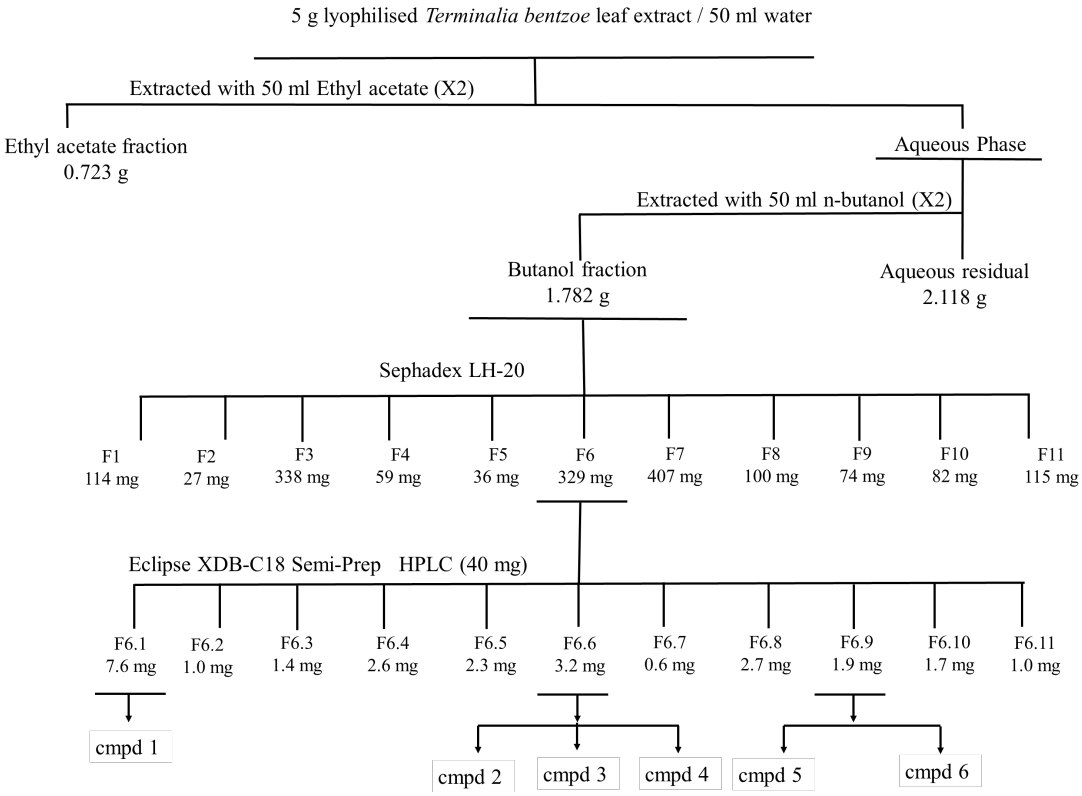


Figure 3. MTT- guided fractionation of *T. bentzoë* extract cytotoxicity against HepG2 cells. Cmpd: Compound. Cmpd 1= α and β- Punicalagin; compd 2= Isoterchebulin; compd 3 = Terflavin A; cmpd 4 = 3,4,6-trigalloyl-β-D-glucopyranose, cmpd 5 = 2''-O-galloyl-orientin; cmpd 6 = 2''-O-galloyl-isoorientin.

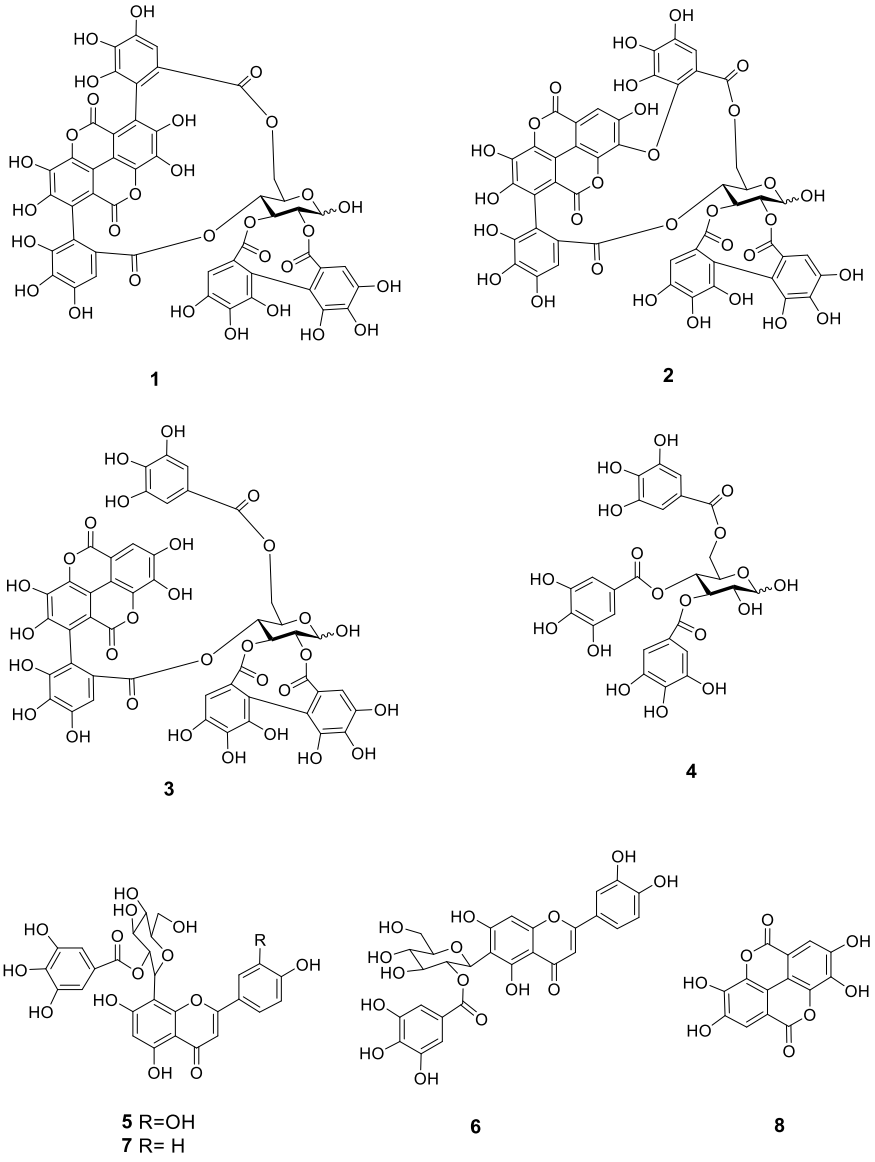
3.5.2. *Antioxidant potential of T. bentzoë fractions*

Both the ethyl acetate (22 ± 0.29 mmol Fe^{2+}) and butanol (21.01 ± 0.56 mmol Fe^{2+}) fractions were significantly ($p \leq 0.05$) higher compared to the aqueous residual fraction. Furthermore, the FRAP value of the organic fractions was greater as compared to the total extract. The butanol fraction of *T. bentzoë* was a more effective scavenger of DPPH• radical as opposed to ethyl acetate and aqueous

residual fractions (**Table 5**). As far as the butanol subfractions were concerned, notably F6, F7, and F8 had better antioxidant activity in FRAP and superoxide radical scavenging assays (**Table 5**).

3.6. Characterisation of the cytotoxic components of T. bentzoë.

In an effort to identify the bioactive constituents in *T. bentzoë*, bioassay-guided fractionation was carried out. LC-MS analysis in conjunction with NMR spectroscopy of *T. bentzoë* butanol fraction 6 and its semi-prep HPLC sub-fractions allowed the identification of 8 phenolic compounds including α and β-punicalagin (**1**), isoterchebulin (**2**), terflavin A (**3**), 3,4,6-trigalloyl-β-D-glucopyranose (**4**), 2"-O-galloyl-orientin (**5**), 2"-O-galloyl-isoorientin (**6**), 2''-O-galloylvitexin (**7**), and ellagic acid (**8**) (**Figure 4**, **Table 6, Supplementary Figures S4)**. The bioactive HPLC subfractions F6.1 (IC_{50} value = 15.8 ± 0.3 µg/mL) revealed to be α and β- punicalagin (**1**) (**Supplementary Figures S5**), while a mixture of isoterchebulin (**2**), terflavin A (**3**), and 3,4,6-trigalloyl-β-D-glucopyranose (**4**) (**Supplementary Figures S6)** was identified from the active subfraction F6.6 (IC_{50} value = 19.6 ± 0.7 µg/mL). A mixture of 2"-O-galloyl-orientin (**5**) and 2"-O-galloyl-isoorientin (**6**) **(Supplementary Figures S7 and Table S1**), with a ratio of 5:2 based on the ^{1}H NMR integration of single hydrogen in each compound was identified from the non-active subfraction F6.9. 2''-O-galloylvitexin (**7**), and ellagic acid (**8**) were detected only by LC-MS from the butanol fraction 6 (**Supplementary Figures S4)**. Additionally, two simple phenolics, gallic acid and methyl gallate were detected in both the ethyl acetate and butanol fractions and identified by GC-MS analysis (**Supplementary figure S8)**. They were likely present in other less bioactive fractions. The levels of gallic and methyl gallate were quantified as 15.6 ± 1.1 and 26.2 ± 8.1 µg/mg total extract) (n=3) by HPLC, respectively.



368

369 **Figure 4.** Chemical structure of polyphenolic compounds (**1**-**8**) identified in *Terminalia bentzoë* L. leaf
370 extract.

371

Table 5. Cytotoxic and antioxidant potential of *T. bentzoë* leaf fractions and the isolated punicalagin (1).

T. bentzoë Fractions		IC_{50} (µg/mL) against HepG2 cells	FRAP[1]	DPPH[2]	Superoxide scavenging activity[2]
Ethyl acetate		20.8 ± 0.1	21.98 ± 0.29^{a,b}	1.17 ± 0.02^{d,e}	7.43 ± 0.14^{b}
Butanol		18.3 ± 3.4	21.01 ± 0.56^{b,c}	0.98 ± 0.06^{e}	7.65 ± 0.10^{b}
Aqueous residual		26.8 ± 5.5	15.29 ± 0.16^{e}	1.78 ± 0.07^{b,c}	10.70 ± 0.20^{a,b}
***T.bnetzoe butanol* subfractions**	**F1**	ND	3.04 ± 0.10^{g}	ND	ND
	F2	ND	13.62 ± 0.16^{f}	2.65 ± 0.04^{a}	16.90 ± 0.35^{a}
	F3	ND	13.80 ± 0.14^{e,f}	1.79 ± 0.12^{b,c}	10.80 ± 0.28^{a,b}
	F4	28.9 ± 1.5	14.42 ± 0.27^{e,f}	2.09 ± 0.08^{b}	8.96 ± 0.12^{b}
	F5	25.7 ± 2.9	18.46 ± 0.15^{d}	1.71 ± 0.07^{c}	8.71 ± 0.10^{b}
	F6	15.7 ± 1.8	23.01 ± 0.68^{a}	1.19 ± 0.09^{d,e}	7.03 ± 0.20^{b}
	F7	19.9 ± 4.7	20.94 ± 0.49^{b,c}	1.01 ± 0.07^{d,e}	7.15 ± 0.18^{b}
	F8	24.6 ± 5.9	19.86 ± 0.29^{c,d}	1.12 ± 0.06^{d,e}	8.76 ± 0.08^{b}
	F9	22.3 ± 3.1	18.82 ± 0.31^{d}	0.99 ± 0.06^{e}	9.33 ± 0.24^{b}
	F10	26.7 ± 6.2	18.47 ± 0.28^{d}	1.34 ± 0.04^{d}	9.05 ± 0.23^{b}
	F11	ND	0.09 ± 0.02^{h}	ND	ND
Active HPLC subfractions	F6.1 (Punicalagin **(1)**)	15.8 ± 0.3	-	-	-
	F6.3	27.5 ± 0.8	-	-	-
	F6.4	17.4 ± 0.4	-	-	-
	F6.5	22.1 ± 0.6	-	-	-
	F6.6	19.6 ± 0.7	-	-	-
	F6.8	18.6 ± 0.4	-	-	-

[1]Values are expressed in units of mmol Fe^{2+}/gFDW; [2]Values are expressed in units of µg/mL; Data represent mean with a standard error of the mean (n=3). Different letters between rows in each column represent significant differences between extracts (p <0.05). ND = IC_{50} value not reached at maximun test dose. "-" = Not tested.

Table 6. Identification of phenolic compound in *T. bentzoë* butanol fraction F6 using high-resolution mass spectrometry, NMR spectroscopy, and comparison with literature and available data.

Compound number	RT/min	Negative ESI-MS [M-H]$^-$	Molecular formula	Compound	References
1	0.76	1083.0555	$C_{48}H_{28}O_{30}$	Punicalagin	[33–36]
2	4.08	1083.0555	$C_{48}H_{28}O_{30}$	Isoterchebulin	[33]
3	4.58	1085.0710	$C_{48}H_{30}O_{30}$	Terflavin A	[37]
4	4.97	635.0867	$C_{27}H_{24}O_{18}$	3,4,6-trigalloyl-β-D-glucopyranose	[38]
5	6.25	599.1017	$C_{28}H_{24}O_{15}$	2"-O-galloyl-orientin	[34]
6	6.25	599.1017	$C_{28}H_{24}O_{15}$	2"-O-galloyl-isoorientin	[34]
7	6.53	583.1074	$C_{28}H_{24}O_{14}$	2″-O-Galloylvitexin	[34,35]
8	6.72	300.9987	$C_{14}H_6O_8$	Ellagic acid	[33]

4. Discussion

Medicinal plants are known to be the epicentre of numerous well established ethnomedicinal systems across the globe [4]. However, more than 84 % of these medicinal plants have been poorly studied in regards to their phytochemical compositions, clinical efficacy as well as their safety and toxicological profiles [4]. Moreover, forest cover is being uprooted across the world, at an unprecedented rate, threatening the survival of at least 15000 medicinal plant species [39]. Plant secondary metabolites have contributed enormously to the modern-day pharmaceutical industry by providing the chemical backbone for almost 25 % of the 1562 clinical agents as well as 60 % of the 246 oncologic drugs approved by the US Federal Drugs Administration between 1981 and 2014 [1,2,40]. As such it is of utmost importance to evaluate the unexplored terrestrial flora for their therapeutic potential, as the dwindling medicinal plant's species continues to stand as a rich repository to probe for novel chemotypes in the drug developmental process.

The current findings highlighted the richness of the different subclasses of polyphenolics, notably phenolics, flavonoids and proanthocyanidins, the distribution of which differed significantly ($p < 0.05$) among the investigated accessions (**Table 2**). The biosynthesis of flavonoids, in particular flavonols, are known to be upregulated in response to ultra-violet radiation [41]. The accumulation of flavonoids in the endemic plant leaves collected from Mauritius may be attributed to the high sunlight conditions and UV radiation which are characteristic to tropical islands like the Mascarene [42].

Given the ubiquitous involvement of oxidative damage in carcinogenesis, antioxidant-rich secondary metabolites, notably, polyphenolics have attracted much interest in the search of novel and alternative treatment modalities for cancer [43,44]. In this vein, the *in vitro* antioxidant activities often correlated strongly with the growth inhibitory activity against cancer cell lines [45,46]. However, it is crucial to note that both ROS and antioxidants have a "Janus-faced" effect in cancer management, as their effect differs in the early events of cancer initiation, from those involved in the survival and propagation of an establish tumors [43,47]. On one hand, where moderate ROS level promotes the transformation of a normal cell to malignant cells, elevating the ROS level in cancer cells beyond the cell tolerance threshold may promote oxidative stress-induced cell cytotoxicity [47–49]. It should be noted that many standard chemotherapies cause cancer cell death via generation of ROS and excessive oxidation [50], therefore the effect of antioxidant polyphenols in combination with these chemotherapies on cancers must be fully evaluated *in vitro* and *in vivo* before use in patients.

The antioxidant mechanism of action of polyphenols are multifaceted, thus a panel of *in vitro* assay models allowed to gauge the antioxidant potential of the evaluated extracts [51]. *T. bentzoë* having the highest abundance of total phenolics and flavonoids also exhibited the most potent antioxidant activity in all six *in vitro* assays and was further evaluated for its cytotoxicity against cancer cell lines. However, the other six plant species might also show cytotoxicity through different mechanisms, which were not investigated in this study.

The cytotoxicity of different *Terminalia* species against multiple cancerous cell lines is documented. Investigation of the cytotoxic activity of methanolic leaf extracts of *T. arjuna* against human chronic myelogenous leukaemia cells led to the isolation of the bioactive ursolic acid (triterpenoid) [52]. Leaf extracts of *T. chebula* suppressed the growth of human breast and lung cancer cell lines [53]. Along a similar line, the leaf extract from *T. catappa* significantly ($p < 0.05$) suppressed the proliferation of human colorectal (SW480) cell line in a dose-dependent manner by downregulating the level of B-cell lymphoma 2 (BCL-2) gene expression while upregulating the level of Caspase 9 and Caspase 3, indicative of the mitochondrial pathway of apoptosis in SW480 cells [54].

The foremost aim of oncologic agents is to precisely target cancerous cells with minimal effect on non-malignant cells. *T. bentzoë* crude extract had a selective index value of approximately 2.5, indicating that the extract had some degree of selectivity towards HepG2 cells as opposed to non-malignant human ovarian epithelial cells. Nevertheless, it is crucial to also assess the cytotoxicity of the extract against a panel of normal cells of different tissue origins, to provide greater insight regarding its toxicity window and safety profile.

In line with the United States National Cancer Institute cytotoxicity guidelines [55,56], *T. bentzoë* crude extract with an IC_{50} value of 22.8 ± 1.3 µg/mL can be considered as a potent candidate for further investigation with regards to its anticancer potential. It is known that, following exposure to toxicants, a subpopulation of cells enters a dormant state. These dormant viable cells further retained their ability to replicate into stem-like progeny cells which might lead to the development of drug-resistance and cancer relapse [57,58]. Thus, while investigating the potential anticancer effect of extracts, it is warranted to also evaluate their long-term effect on the replicative ability of the cancer cell line. The colony formation assay is a simple and useful *in vitro* model, considered as the gold standard to predict the long-term sensitivity of cancer cell response to therapeutics treatment [57,59]. In this vein, the current study evidenced the ability of *T. bentzoë* to impede the replicative potential of HepG2 cells (**Figure 1**). Numerous anticancer agents, including polyphenols, are known to abrogate the limitless replicative ability of different cancer cell variants [60,61]. Catechin, catechin-3-O-gallate, 7-O-galloyl catechin, and methyl gallate purified from *Acasis hydaspica* is reported to suppress the long-term clonal proliferation of prostate cancer (PC-3) cells [62].

This study also attempted to delineate the mode of *T. bentzoë* induced HepG2 cell death. Flow cytometric analysis of annexin V-FITC and PI dual stained HepG2 cells treated with 40 µg/mL, revealed a significant ($p < 0.05$) increased in both Annexin V-FITC and PI fluorescence level, compared to the untreated control, thus indicating the putative activation of both apoptotic and necrotic pathways in HepG2 cells. Cells grown as monoculture are known to initiate apoptosis which is terminated by necrosis like events, also termed as secondary necrosis, due to the absence of phagocytic scavenger cells [63,64]. Numerous studies evidenced this type of cancer cell death following treatment with cytotoxic agents. Ellagic acid was reported to induce both apoptotic and necrotic cell death mechanism in human pancreatic cancer cell cultures [65]. A similar effect of *Lepidium sativum* and *Vitis vinifera* extract were highlighted in human breast and skin cancer cells, respectively [66,67].

Plant extracts and/or thereby derived phytochemicals are known to provoke cancer cell death by causing oxidative damage to genetic material mediated cell cycle arrest and subsequent cell death [68,69]. With this in mind, the DNA damage to HepG2 cellular DNA following *T. bentzoë* crude extracts treatment was investigated using the alkaline comet assay and scored in terms of tail length, tail intensity, and olive tail moment. The comet assay is a well-established and highly sensitive method for the detection of DNA damage and fragmentation pattern [70]. *T. bentzoë* induced 4 folds significantly ($p < 0.001$) higher DNA damaged to HepG2 cells compared to the untreated control, as reflected by the olive tail moments (**Table 4**). Consistent with the DNA damaging ability, *T. bentzoë* halted the cell cycle progression significantly ($p < 0.01$, *versus* control) in the G0/G1 phase (**Figure 2**). A similar observation was reported in lung cancer cells, where casuarinin, a tannin purified from *T. arjuna* L. bark, provoked apoptotic mechanism via DNA fragmentation and G0/G1 phase cell cycle arrest [71].

It is noteworthy that, the cytotoxicity of the extract may arise from the complex interplay of the cocktails of secondary metabolites present, which may act either synergistically or antagonistically to produce the overall results. Also, crude extracts often comprised a pool of inactive phytoconstituents that dilute the efficacy of the active components [72,73]. It is therefore of paramount importance to purify and identify the principal bioactive components to conduct further mechanistic studies to establish their molecular mode of action [72,74]. Moreover, isolating the lead compounds also allows for the potential structural modification in an attempt to enhance their selectivity and potency [75]. As such, the MTT-guided fractionation revealed that only six out of the eleven preparative HPLC subfractions retained the potent cytotoxicity of the crude extract (**Figure 3**). The HPLC subfraction F6.1 was 1.4 folds more potent as compared to the crude extract.

LC-MS analysis in conjunction with NMR spectroscopy allowed the identification of 8 phenolic compounds from the most bioactive subfraction F6. Punicalagin, isoterchebulin and ellagic acid have been previously reported from the bark extract of the same species collected in Réunion Island [33]. Although, the other identified compounds are being reported for the first time, to the best of our current knowledge, in *T. bentzoë* leaf, some were reported in the leaf extracts of other *Terminalia*

484 species. For instance, 2″-O-galloylvitexin and gallic acid were identified from *T. brachystemma* Welw.
485 ex Hiern and *T. mollis* M. Laws leaves, respectively [35]. Likewise, terflavin A was found in *T.*
486 *catappa* L. leaf [37].
487 Punicalagin and ellagic acid were reported to induce S Phase arrest and G0/G1 phase arrest in
488 HepG2 cells, respectively [76]. Moreover, punicalagin induced G0/G1 phase arrest in papillary
489 human thyroid carcinoma (BCPAP) cells via the NF-κB signaling pathway [77]. In human ovarian
490 cancer (A2780) cells, punicalagin treatment was associated with an increase in the number of cells
491 arrested at G1/S phase as well as the downregulation of the β-catenin signaling pathway [78]. Ellagic
492 acid administration in prostate cancer patients was associated with decreased prostate-specific
493 antigen as well as reduced chemotherapy-induced myelotoxicity [79]. It may be strongly proposed
494 that the overall cytotoxicity of *T. bentzoë* leaf extract may be a synergistic effect of the identified
495 compounds. Gallic acid was reported to induce S phase arrest in HepG2 cells [80] and be cytotoxic to
496 ovarian cancer cells [81]. Furthermore, gallic acid was also shown to impair centrosomal clustering
497 in human cervical cancer (Hela) cells, thus causing a mitotic catastrophe and ultimate cell G2/M phase
498 cell cycle arrest.

499 **5. Conclusion**

500 The findings evidenced the selective long-term cytotoxicity of the antioxidant-rich *T. bentzoë* leaf
501 extract against HepG2 cells. This plant which is also used traditionally in the mitigation of asthma,
502 haemorrhages, diarrhoea, and sexually transmissible diseases, amongst others, has also shown some
503 *in-vitro* anticancer activities against HepG2 cells. The cytotoxic nature of *T. bentzoë* leaf extract against
504 cancerous cells indicated that *T. bentzoë* leaf has the potential to be repurposed in the mitigation of
505 cancer as part of traditional medicine. The results so far generated, supports the hypothesis that *T.*
506 *bentzoë* extract induced apoptosis/necrosis cell death in HepG2 cells via the degradation of cellular
507 genetic material and subsequent arrest of the cell cycle progression in G0/G1 phase. Overall, the MTT-
508 guided fractionation of the crude extract allowed the characterisation of 10 phenolic compounds
509 including punicalagin, ellagic acid, gallic acid, and methyl gallate that are known to have established
510 *in vitro* and *in vivo* anticancer activities. However, the contribution of the other non-identified
511 phytoconstituents cannot be excluded and thus further evaluation of identified components is
512 warranted alongside the crude extract. The metabolite profiling of the leaf extract may be envisaged
513 in future studies. This will allow to precisely ascribe the bioactive molecules present in *T. bentzoë* leaf.
514 Taking into consideration the guidelines of the US national cancer institute as well as the selectivity
515 index observed in this study, *T. bentzoë* leaf extract revealed as a promising candidate that can be
516 exploited in the search for novel anticancer agents. The current study provided base line data for
517 more in-depth investigations of the chemotherapeutic potential of *T. bentzoë* plant extracts. Further
518 mechanistic investigation, directed towards delineating the molecular mechanisms via which the
519 purified bioactive entities target of the aberrant signaling pathways involved in carcinogenesis, is
520 needed to fuel *in-vivo* studies. Future animal studies and ideally human clinical trials are warranted
521 to establish the physiological doses for human administration.

522 **Supplementary Materials: F**igure S1: Cell viability profile of HepG2 cells treated with *T. bentzoë* extract and
523 fractions. Figure S2: Representatives annexin V-FITC/PI flow cytometric profile of HepG2 cells, 48 hours post
524 extract/control treatment. Figure S3: Representatives cell cycle histogram of HepG2 cells, 48 hours post
525 extract/control treatment. Figure S4: UPLC ESI MS of the butanol fraction 6 at the positive (A) and negative (B)
526 mode. Figure S5: ^{1}H NMR spectrum (A) and HSQC spectrum (B) for punicalagin (1) in an HPLC fraction at RT
527 2-3 min in methanol-d_4. Figure S6: ^{1}H NMR spectra for a mixture of isoterchebulin (2), terflavin A (3), and 3,4,6-
528 trigalloyl-beta-D-glucopyranose (4) in a HPLC fraction at RT18-20 min in acetone-d_6 (A) and methanol-d_4 (B).
529 Figure S7: ^{1}H NMR spectra for a mixture of 2"-O-galloyl-orientin (3) and 2"-O-galloyl-isoorientin (6) with a ratio
530 of 5:2 based on the integration of single hydrogen in each compound. Figure S8: GC –MS chromatogram of TMSi
531 derivatives of ethyl acetate (A) and butanol (B) fraction of *T. bentzoë*. Table S1 ^{1}H NMR spectral data (500 MHz,
532 CD_3OD) of the O-galloyl-C-glycosylflavones 7 and 8 (5:2) [" in ppm, multiplicities and J values (Hz) are given in
533 parentheses].

534 **Author Contributions:** Conceptualization, N.R and V.N; methodology, N.R., P.R, E.P., J.M., and T.D.; software,
535 N.R., P.R, E.P., J.M., and T.D.; validation, N.R., P.R, E.P., J.M., and T.D.; formal analysis, N.R., P.R, E.P., J.M., and

T.D.; investigation, N.R., P.R, E.P., J.M., and T.D.; resources, E.B., T.B., W.L., and V.N.; data curation, N.R., P.R, E.P., J.M., and T.D.; writing—original draft preparation, N.R.; writing—review and editing, P.R, T.D., T.B., W.L., E.B., and V.N.; visualization, N.R., E.P., J.M., and T.D ; supervision, T.B., W.L., and V.N.; project administration, T.B., V.N.; funding acquisition, T.B., W.L., and V.N. All authors have read and agreed to the published version of the manuscript.

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