

Parasite Epidemiology and Control

Empowering Rural Communities for Effective Larval Source Management: A Small-Scale Field Evaluation of a Community-Led Larviciding Approach to Control Malaria in South-Eastern Tanzania --Manuscript Draft--

Manuscript Number:	PAREPI-D-23-00078R3
Article Type:	Full length article
Section/Category:	Malaria, lymphatic filariasis, schistosomiasis.
Keywords:	Malaria control; larviciding; larval source management; biolarvicides; community engagement; Tanzania; Ifakara Health Institute
Corresponding Author:	Salum Abdallah Mapua Ifakara Health Institute Morogoro, TANZANIA, UNITED REPUBLIC OF
First Author:	Salum Abdallah Mapua
Order of Authors:	Salum Abdallah Mapua Alex J. Limwagu Dmitry Kishkinev Khamis Kifungo Ismail H. Nambunga Samuel Mziray Gwakisa John Wahida Mtiro Kusiryu Ukio Javier Lezaun Frederic Tripet Fredros O. Okumu
Abstract:	<p>Introduction</p> <p>Larval source management, particularly larviciding, is mainly implemented in urban settings to control malaria and other mosquito-borne diseases. In Tanzania, the government has recently expanded larviciding to rural settings across the country, but implementation faces multiple challenges, notably inadequate resources and limited know-how by technical staff. This study evaluated the potential of training community members to identify, characterize and target larval habitats of <i>Anopheles funestus</i> mosquitoes, the dominant vector of malaria transmission in south-eastern Tanzania.</p> <p>Methods</p> <p>A mixed-methods study was used. First, interviewer-administered questionnaires were employed to assess knowledge, awareness, and perceptions of community members towards larviciding (N = 300). Secondly community-based volunteers were trained to identify and characterize aquatic habitats of dominant malaria vector species, after which they treated the most productive habitats with a locally-manufactured formulation of the biolarvicide, <i>Bacillus thuringiensis</i> var <i>israelensis</i>. Longitudinal surveys of mosquito adults and larvae were used to assess impacts of the community-led larviciding programme in two villages in rural south-eastern Tanzania.</p> <p>Results</p> <p>At the beginning of the program, the majority of village residents were unaware of</p>

	<p>larviciding as a potential malaria prevention method, and about 20% thought that larvicides could be harmful to the environment and other insects. The trained community volunteers identified and characterized 360 aquatic habitats, of which 45.6% had <i>Anopheles funestus</i>, the dominant malaria vector in the area. The preferred larval habitats for <i>An. funestus</i> were deep and had either slow- or fast-moving waters. Application of biolarvicides reduced the abundance of adult <i>An. funestus</i> and <i>Culex</i> spp species inside human houses in the same villages, by 46.3% and 35.4% respectively. Abundance of late-stage instar larvae of the same taxa was also reduced by 74% and 42%, respectively.</p> <p>Conclusion</p> <p>This study demonstrates that training community members to identify, characterize, and target larval habitats of the dominant malaria vectors can be effective for larval source management in rural Tanzania. Community-led larviciding reduced the densities of adult and late-stage instar larvae of <i>An. funestus</i> and <i>Culex</i> spp. inside houses, suggesting that this approach may have potential for malaria control in rural settings. However, efforts are still needed to increase awareness of larviciding in the relevant communities.</p>
Response to Reviewers:	

Parasite Epidemiology and Control
Radarweg 29
1043 NX Amsterdam
The Netherlands

August 3rd, 2023

Dear Editor

I hope this finds you well.

It is with great enthusiasm that I submit my manuscript titled **“Empowering Rural Communities for Effective Larval Source Management: A Small-Scale Field Evaluation of a Community-Led Larviciding Approach to Control Malaria in South-Eastern Tanzania”**. evaluate methods to involve community members in identifying, characterizing, and targeting the main aquatic habitats of the dominant malaria vector species using biolarvicide, and the impact of such an intervention on the density of the most relevant vector species in rural areas of Tanzania. The study focused on villages in the Kilombero valley, southeastern Tanzania, a meso-endemic community where *An. funestus* mediates >90% of malaria transmission events.

This study was funded in parts by Wellcome trust International Masters Fellowship in Tropical Medicine and Hygiene, and Bill and Melinda Gates Foundation. These grants do not cover publication fees; I therefore request Parasite Epidemiology and Control to publish these outstanding findings, without charging fees.

I thank you in advance for your kind consideration and I look forward for your response.

Yours sincerely,



Salum A. Mapua
Corresponding author
+447827261098; smapua@ihi.or.tz



Dar es Salaam
PO Box 78373
Tel: 0 222 771 714
Fax: 0 222 771 714

Ifakara
PO Box 53
Tel: 0 232 625 164
Fax: 0 232 625 312

Bagamoyo
PO Box 74
Tel: 0 232 440 065
Fax: 0 232 440 064

Rufiji
PO Box 40 Ikwiriri
Tel: 0 232 010 007
Fax: 0 232 010 742

Mtwara
PO Box 1048
Tel: 0 232 333 487
Fax: 0 232 333 487

Kigoma
PO Box 1077
Tel: 0 282 803 655

September 17th, 2024

Dear Editor, *Parasite Epidemiology and Control*

Ref: Research paper re-submission (Manuscript number: PAREPI-D- 23-00078R2)

We thank the reviewer for reviewing this manuscript titled “**Empowering Rural Communities for Effective Larval Source Management: A Small-Scale Field Evaluation of a Community-Led Larviciding Approach to Control Malaria in South-Eastern Tanzania**”. We would like also to thank you for allowing us to resubmit a revised copy of the manuscript. In the revised manuscript, we have addressed all the comments and highlighted the revisions. We also provide below, point-by-point responses to the comments.

Yours Sincerely,



Salum A. Mapua
Corresponding author
Tel: (+447827261098)
Email: smapua@ihi.or.tz

Response to editor’s comments

Comment 1. I have completed my evaluation of your manuscript. The reviewers recommend reconsideration of your manuscript following minor revision and modification. I invite you to resubmit your manuscript after addressing the comments below.

Response: We thank the editor for the comment. In this revised manuscript, all comments from a reviewer have been addressed accordingly.

Response to reviewer's comments

Reviewer #3

Comment 1: Minor Improvements in Cohesion: Ensure that the transitions between sections (for example between the training description and the effectiveness assessment) are smooth and cohesive. This will help maintain a logical flow throughout the manuscript.

Response: We appreciate the suggestion and have revised the training sections to remove redundancy and improve the flow. Additionally, we have reviewed the entire manuscript to ensure consistency in flow.

Comment 2: Consistency in Writing Style: Review the manuscript for any remaining inconsistencies in writing style or formatting, particularly in sections where minor adjustments were previously noted.

For example in the revised response to comment 1 (Study aim) (129-134)

Rewording "involve" to "for involving" could improve grammatical flow (129).

"Primary" instead of "main" could add more specificity (130).

Combining the first two sentences into a smoother statement can improve readability.

Using "accounts for" instead of "mediates" looks clearer for a broader audience and aligns better with scientific writing (134).

Response: We are truly grateful for this comment. In addition to addressing the specific points highlighted by the reviewer, we have taken the opportunity to revise additional aspects for consistency throughout the manuscript. For instance, we have consistently formatted *An. funestus* s.s to "*An.funestus* s.s."

Comment 3: Clarify Impact of Findings: In the discussion-conclusion, clearly articulate the potential impact of the findings on malaria control programs, specifically addressing how these results can be applied or adapted to similar settings. The revised conclusion does touch on the impact of the findings but could be slightly enhanced to more explicitly articulate the potential implications for malaria control programs.

Response: We are grateful for the positive review. We have revised the conclusion section to ensure clarity on the potential impact of the findings and how they can be adapted in similar settings.

1 **Empowering Rural Communities for Effective Larval Source Management: A**
2 **Small-Scale Field Evaluation of a Community-Led Larviciding Approach to**
3 **Control Malaria in South-Eastern Tanzania**

4
5 Salum A. Mapua^{1,2*#}, Alex J. Limwagu^{1#}, Dmitry Kishkinev², Khamis Kifungo¹, Ismail H.
6 Nambunga¹, Samuel Mziray³, Gwakisa John⁴, Wahida Mtiro⁴, Kusirye Ukio⁴, Javier Lezaun⁵,
7 Frederic Tripet⁶, Fredros O. Okumu^{1,7,8,9}

8
9 1. Environmental Health and Ecological Sciences Department, Ifakara Health Institute,
10 P. O. Box 53, Morogoro, Tanzania

11 2. School of Life Sciences, Keele University, Huxley Building, Keele, Staffordshire ST5
12 5BG, UK

13 3. Tanzania Biotech Products Limited, P. O. Box 30119, Kibaha, Pwani, Tanzania

14 4. President's Office-Regional Administration and Local Government. Morogoro
15 Regional Secretariat, P.O. Box 650 Morogoro, Tanzania

16 5. Institute for Science, Innovation and Society; School of Anthropology and Museum
17 Ethnography, University of Oxford, 64 Banbury Road, Oxford OX2 6PN, UK

18 6. Swiss Tropical and Public Health Institute, Kreuzgasse 2, 4123 Allschwil, Switzerland

19 7. School of Public Health, Faculty of Health Sciences, University of the Witwatersrand,
20 Johannesburg, South Africa

21 8. School of life science and bioengineering, The Nelson Mandela African Institution of
22 Science and Technology, P. O. Box 447, Arusha, Tanzania

23 9. School of Biodiversity, One Health and Veterinary Medicine, University of Glasgow,
24 Glasgow G61 1QH, UK

25
26
27
28 * Correspondence: smapua@ihi.or.tz and fredros@ihi.or.tz

29 # These authors contributed equally to this work.
30
31
32
33

34 **Abstract**

35 **Introduction:** Larval source management, particularly larviciding, is mainly implemented in
36 urban settings to control malaria and other mosquito-borne diseases. In Tanzania, the
37 government has recently expanded larviciding to rural settings across the country, but
38 implementation faces multiple challenges, notably inadequate resources and limited know-
39 how by technical staff. This study evaluated the potential of training community members to
40 identify, characterize and target larval habitats of *Anopheles funestus* mosquitoes, the
41 dominant vector of malaria transmission in south-eastern Tanzania.

42

43 **Methods:** A mixed-methods study was used. First, interviewer-administered questionnaires
44 were employed to assess knowledge, awareness, and perceptions of community members
45 towards larviciding (N = 300). Secondly community-based volunteers were trained to identify
46 and characterize aquatic habitats of dominant malaria vector species, after which they
47 treated the most productive habitats with a locally-manufactured formulation of the
48 biolarvicide, *Bacillus thuringiensis* var *israelensis*. Longitudinal surveys of mosquito adults
49 and larvae were used to assess impacts of the community-led larviciding programme in two
50 villages in rural south-eastern Tanzania.

51

52 **Results:** At the beginning of the program, the majority of village residents were unaware of
53 larviciding as a potential malaria prevention method, and about 20% thought that larvicides
54 could be harmful to the environment and other insects. The trained community volunteers
55 identified and characterized 360 aquatic habitats, of which 45.6% had *Anopheles funestus*,
56 the dominant malaria vector in the area. The preferred larval habitats for *An. funestus* were
57 deep and had either slow- or fast-moving waters. Application of biolarvicides reduced the
58 abundance of adult *An. funestus* and *Culex* spp. species inside human houses in the same
59 villages, by 46.3% and 35.4% respectively. Abundance of late-stage instar larvae of the
60 same taxa was also reduced by 74% and 42%, respectively.

61

62 **Conclusion:** This study demonstrates that training community members to identify,
63 characterize, and target larval habitats of the dominant malaria vectors can be effective for
64 larval source management in rural Tanzania. Community-led larviciding reduced the
65 densities of adult and late-stage instar larvae of *An. funestus* and *Culex* spp. inside houses,
66 suggesting that this approach may have potential for malaria control in rural settings.
67 However, efforts are still needed to increase awareness of larviciding in the relevant
68 communities.

69

70 **Keywords:** Malaria control, larviciding, larval source management, biolarvicides,
71 community engagement, Tanzania, Ifakara Health Institute.

72

73 **Introduction**

74

75 Since 2000, malaria incidence and mortality have significantly decreased due primarily to
76 expanded vector control with insecticide-treated nets (ITNs) and indoor residual spraying
77 (IRS), along with improved diagnosis and treatment [1]. However, these gains began
78 plateauing around 2015, and many high-burden countries now report increases in cases [2].
79 The latest estimates from the World Health Organization (WHO) indicate that the disease
80 still causes around 249 million cases and 608,000 deaths, with a majority of this burden
81 being in Africa [2]

82

83 In addition to poor socio-economic circumstances and the generally weak health systems in
84 endemic communities [3,4], the continued burden of malaria is also associated with major
85 biological threats such as the widespread resistance of malaria vectors to insecticides [5,6]
86 and behavioral resistance to indoor interventions [7–9], as well as growing tolerance of the
87 malaria parasite (*Plasmodium* spp.) to anti-malarial drugs. Human behaviors, such as
88 staying outdoors unprotected during times of increased malaria transmission, also contribute
89 to ongoing malaria transmission [10–12]. Another important but often less discussed
90 challenge is that sustained malaria prevention is untenable without sufficient engagement of
91 local communities and other stakeholders [13,14]

92

93 Research has shown that involving rural communities in vector control programs is beneficial
94 for malaria prevention and control [14,15]. The success of such interventions is dependent
95 on the level of education and awareness of malaria prevention tools among community
96 members [16]. Studies conducted in Tanzania [17,18], Rwanda[19], Burkina Faso[20], Bioko
97 Island[21] and Malawi [22,23] have demonstrated the effectiveness of involving community
98 members in routine surveillance, larviciding, and house improvements for vector control.
99 Based on these studies, community engagement has the potential to play a significant role
100 in successful implementation of vector control programs in other African countries.

101

102 In Tanzania, the focus of malaria vector control has been on ITNs and IRS [24–27]. The
103 2014-2020 National Malaria Strategic Plan [28] also recommended deploying larviciding in
104 urban areas in line with the WHO guidelines [29,30], but this has recently been extended to
105 rural areas [31–33]. However, a recent study found limited community knowledge about
106 larviciding and other challenges, including insufficient funding and technical expertise [31],

107 which may hinder the sustainability of the program in rural settings where malaria burden is
108 higher. As in the urban settings, larviciding program in rural Tanzania involves the
109 application of biolarvicides to all water bodies, which is a massive undertaking that severely
110 constrains the program due to limited resources. The expansion of larviciding programs to
111 rural communities in particular requires a deeper understanding of the ecology of the
112 dominant malaria vector species, since the habitats may be more expansive and more
113 numerous than as envisioned in the current WHO guidelines [29,30].

114

115 Previous entomological studies in rural south-eastern Tanzania, where *An. arabiensis* and
116 *An. funestus* are the predominant malaria vectors, have shown that the latter species
117 **accounts for** over 90% of the ongoing malaria transmission [34,35]. *An. funestus* is also
118 highly resistant to common public health pesticides targeting adult mosquitoes [34,36] thus
119 requiring alternative control options (e.g. the use of the biocontrol agents such as, *Bacillus*
120 *thuringiensis israelensis* (*Bti*) or *Lysinibacillus sphaericus* (*Ls*)). While preliminary results
121 indicated that the aquatic habitats of immature *An. funestus* can be indeed few (i.e., only a
122 small fraction of habitats are occupied by *An. funestus*), fixed (i.e., they tend to be
123 permanent or semi-permanent) and findable (i.e., have unique characteristics allowing for
124 ease of identification) [37]. Expanded investigations (Msugupakulya *et al.*, Unpublished data)
125 suggest a more expansive range of habitats in different seasons, meaning that any effective
126 control would require extensive engagement with locals to effectively search, characterize
127 and target the habitats.

128

129 The aim of this study is to evaluate methods **for involving** community members in identifying,
130 characterizing, and targeting the **primary** aquatic habitats of the dominant malaria vector
131 species using biolarvicide, and to assess the impact of such an intervention on the density of
132 the most relevant vector species in rural areas of Tanzania. This study focused on villages in
133 the Kilombero valley, southeastern Tanzania, a meso-endemic community where *An.*
134 *funestus* **accounts for** over 90% of malaria transmission events.

135

136 **Methods**

137 **Study area**

138 The study was done in two villages, Igumbiro (8.35021°S, 36.67299°E) and Sululu
139 (8.00324°S, 36.83118°E), both located on the Kilombero valley flood plain, south-eastern
140 Tanzania. Igumbiro is at a slightly higher altitude (340 meters) above sea level than Sululu
141 (315 meters above sea level). Annual rainfall ranges from 1200 mm to 1400 mm, and the
142 temperature ranges from 20 °C to 32 °C in these villages [38,39]. The majority of residents

143 are subsistence rice farmers, but other crops such as sweet potatoes, beans, and maize are
144 also produced [38].

145
146 **Figure 1:** Map showing two villages in the Kilombero Valley, south-eastern Tanzania
147 where the study was conducted.

148 149 **Longitudinal entomological surveillance of adult mosquitoes**

150 Mosquitoes were routinely collected in the two villages during the same period from March
151 2020 to September 2021. Three different households were sampled nightly in each village
152 for four nights in a week, with one additional household sampled once per week as a
153 sentinel station. The collections were done using CDC light traps from 6pm to 6am for host-
154 seeking mosquitoes [40] and Prokopack aspirators from 6am to 7am for resting mosquitoes
155 [41]. Collected female mosquitoes were sorted by taxa and physiological state [42]. Recent
156 studies in these settings had determined that the *An. gambiae* s.l. and *An. funestus* group
157 consisted of *An. arabiensis* (100%) and *An. funestus* s.s. (>93%) respectively [34,43,44], but
158 also that majority of the transmission (>90%) is driven by *An. funestus* s.s. [34,45].

159 160 **Recruitment and training of community volunteers**

161 A total of 300 adult community members, 150 from each village, were randomly selected
162 using residents' lists provided by village authorities. An interviewer-administered
163 questionnaire, conducted in Swahili using KoboToolbox software [46], assessed the
164 community members' prior knowledge, awareness, and perception of disease-transmitting
165 mosquitoes and larviciding. The questionnaires were administered following written informed
166 consent. From these, ten community members from each village, meeting specific criteria,
167 were selected for entomological training with an emphasis on equal gender distribution. The
168 criteria were: i) ability to read and write properly, ii) involvement of the participant's
169 household in the entomological surveillance, iii) residency in the village for at least two
170 years, and iv) age between 18 and 50 years.

171
172 The training program for community volunteers was comprehensive, covering essential
173 topics related to mosquito larvae identification, GPS usage, and the application of
174 biolarvicides following WHO recommendations [29,30]. It consisted of both theoretical and
175 practical components. The theoretical aspect involved presentations and demonstrations
176 (Fig. 2) on mosquito life stages, the biology of malaria vectors, and malaria transmission.
177 The practical aspect included field visits, where volunteers engaged in hands-on training to
178 practice identifying and characterizing mosquito larvae habitats, using GPS for mapping, and

179 applying biolarvicides. Additionally, the practical sessions covered sampling techniques,
180 estimating larval density, processing and storing immature mosquitoes, and recording data.

181

182 The training spanned three full days, with each day divided into theoretical sessions in the
183 morning and practical sessions in the afternoon. Over the course of three days, three
184 simplified modules were covered: i) an introduction to the control and biology of malaria
185 vectors, ii) the identification and characterization of aquatic habitats of malaria vectors in
186 Ulanga and Kilombero districts, and iii) larval source management. Facilitators from the
187 Ifakara Health Institute and the government-owned biolarvicide plant (Tanzania Biotech
188 LTD) led the training, ensuring high-quality instruction and relevance to local contexts.
189 District malaria focal persons participated as liaisons, enhancing the connection between the
190 community, government, and the Ifakara Health Institute. Although a formal post-
191 assessment of the training program to evaluate the knowledge and skills acquired by
192 community members was not conducted due to budget and timeline constraints, participants'
193 skills were continuously assessed through hands-on activities during the practical sessions.

194

195 **Figure 2:** Theoretical and practical training of the community members for
196 identification and characterization of aquatic habitats and application of larviciding.

197

198 **Identification and characterization of the aquatic habitats of the dominant malaria** 199 **vectors, *Anopheles funestus***

200 The trained community members were deployed in their respective villages, with 2 km
201 transects allocated to each participation for the identification and characterization of water
202 bodies. Based on our previous survey [37], the focus was on types of habitats shown to be
203 favoured by *An. funestus* mosquitoes. For each aquatic habitat, a 350 ml larval dipper was
204 used to check for the absence or presence of *An. funestus* larvae based on WHO larval
205 survey guidelines [30]. Larvae counts and mosquito species identification were recorded per
206 dipper sample in a square meter habitat. Each dipper was treated as an individual
207 observation. Habitat types were defined, and environmental features were recorded.
208 Physicochemical parameters (temperature (°C), total dissolved solid (ppm), acidity (pH),
209 electrical conductivity (µS/cm) and dissolved oxygen (ppm)) were also measured, and GPS
210 coordinates were recorded for each habitat.

211

212 **Figure 3:** Example of the common *An. funestus* larvae habitats found in Sululu and
213 Igumbiro villages, south-eastern Tanzania.

214

215 **Application and monitoring of the efficacy of *Bacillus thuringiensis israelensis* (Bti)**

216 Following the habitat characterization and larval surveys, the trained community members
217 applied *Bti* (i.e., 10 ml in liquid formulation for every square meter) to all aquatic habitats
218 within the 2 km transect that has been identified as including *An. funestus* larvae. Our study
219 employed *Bti* serotype H-14, a product registered with both the Central Pesticide
220 Registration of Cuba and the Tropical Research Institute of Tanzania. This serotype is
221 manufactured under license by the LABIOFAM Enterprise Group and sourced from Tanzania
222 Biotech LTD, a local manufacturer. It is commercially available under the name BACTIVEC®
223 (ITU 1200/mg). Application of the larvicide in both villages was done using the IK Vector
224 Control Super Pressurized sprayers. This was followed by larval monitoring surveys after 24
225 hours, 7, 23 and 30 days. Data from the larval surveys conducted during aquatic habitat
226 characterization and after application were considered to enable pre and post intervention
227 assessment.

228

229 **Data analysis**

230 The open-source software, R programming language version 3.6.0 [47] was used for the
231 statistical analyses. Linear mixed effect models were used to assess the association
232 between number of female adult mosquitoes collected per night per house, and status of the
233 larviciding intervention (i.e., pre-larviciding and post-larviciding) in each village. In addition,
234 households were used as a random effect. The linear mixed effect model was fitted using
235 *lmer* function found within *lme4* package, whilst the variance estimator was set to Restricted
236 Maximum Likelihood (REML) [48]. Similar modelling was performed to determine
237 association between number of larvae per habitat per village and status of larviciding, with
238 the community member names as the random effect.

239

240 The questionnaire survey data was retrieved from KoboToolbox, cleaned, and coded for
241 easier statistical analyses in R programming language software. The data was used to
242 determine percentage of the community members that had knowledge and awareness of
243 larviciding intervention, but also their perceptions towards larviciding. The Generalized
244 Linear Model (GLM) was also used to investigate the association between community
245 members perception towards larviciding intervention and their socio-demographic
246 characteristics such as gender, age and literacy status.

247

248 GLMMs were used to determine the relationship between presence or absence of mosquito
249 larvae and environmental characteristics. Highly correlated independent variables were
250 removed, and the full model was fitted using a *glmer* function with binomial distribution and
251 *logit* link function. Model selection was performed by removing insignificant terms. Final

252 models were validated by assessing average residuals against expected values using
253 binned residual plots. The same approach was applied to reveal the association between
254 physicochemical attributes and occurrence of *An. funestus* larvae and other mosquitoes.

255

256 **Results**

257 **Knowledge, awareness and perception of community members towards larviciding**

258 A total of 300 community members (46% women and 54% men) participated in the
259 knowledge assessment on larviciding. Of these, only 39% were aware of larviciding as a
260 malaria control strategy, a majority of these being men (Table 3). While age of participants
261 did not influence their knowledge, those with secondary education were more than five times
262 more knowledgeable than those without formal education ($p < 0.001$). Most of the
263 participants had become aware of larviciding from family members or friends (Table 1). A
264 large proportion (86%) of the participants reported that they had never observed larviciding
265 being implemented in their respective villages. While 8-20% of the participants believed that
266 larvicides were potentially harmful to other organisms (insects, large animals, and fish),
267 more than 80% agreed that larviciding could be necessary against disease-transmitting
268 mosquitoes. Indeed, 71% were willing to participate in the implementation of larviciding
269 programs in their communities (Table 2).

Table 1: Knowledge and awareness of larviciding in the communities (N = 300)

Variable	Response	Percentage (n)
Awareness of larviciding (N = 300)	Yes	39 (117)
	No	61 (183)
Source of information (N = 117)	Family/Friends	65 (76)
	Radio/Television	14.5 (17)
	Village meeting	5.1 (6)
	Village health officer	6 (7)
	Village agricultural officer	0 (0)
	Malaria focal person	6 (7)
	Other sources	13.7 (16)
	Do not remember	1 (1)
Have you ever witnessed larviciding implemented in the community (N = 300)	Yes	8 (24)
	No	86 (258)
	Do not remember	6 (18)
Where are these larvicides manufactured (N = 300)	Domestic	6.7 (20)
	Abroad	6.7 (20)
	Both	1.3 (4)
	Do not know	85.3 (256)
Which is the first stage during larvicides application (N = 300)	Identification of aquatic habitats	29 (87)
	Community sensitization	28.3 (85)
	Estimation of larvicide quantity	1.7 (5)
	Spraying larvicides	3 (9)
	Other	3.3 (10)
	Do not know	34.7 (104)

272
273
274

Table 2: Perception of community members towards larviciding for the malaria prevention (N = 300)

Variable	Response	Percentage (n)
Have you ever participated in larviciding (N = 300)	Yes	1.3 (4)
	No	98.7 (296)
Do you think larvicides are harmful to insect (N = 300)	Yes	20 (60)
	No	31.3 (94)
	Do not know	48.7 (146)
Do you think larvicides are harmful to fish (N = 300)	Yes	8 (24)
	No	39.3 (118)
	Do not know	52.7 (158)
Do you think larvicides are harmful to animal (N = 300)	Yes	12.3 (37)
	No	40.7 (122)
	Do not know	47 (141)
Willingness to participate in larviciding (N = 300)	Yes	71 (213)
	No	29 (87)
Acceptance of larviciding (N = 300)	Agree	82.3 (247)
	Do not agree	4.7 (14)
	Neutral	13 (39)

275
276
277
278
279
280

281
282
283

Table 3: Association between socio-demographic characteristics and community perception towards larviciding

Category	Variable	Odds ratio (95% CI)	P-value
Sex	Female	1	-
	Male	1.84 (1.13-2.99)	0.01
Age category (in years)	18-25	1	-
	26-35	0.87 (0.46-1.68)	0.69
	36-45	0.66 (0.32-1.37)	0.26
	46-50	0.50 (0.15-1.59)	0.24
	Above 50	0.63 (0.32-1.21)	0.17
Education level	No formal education	1	-
	Primary	1.51 (0.85-2.68)	0.16
	Secondary	5.40 (2.20-13.24)	<0.001

284

285 **Habitat characteristics and mosquito species in different aquatic habitats**

286 A total of 360 aquatic habitats were surveyed, including 167 (46.4%) in Sululu and 193
287 (53.6%) in Igumbiro village (Table 4). Larvae of the dominant malaria vector, *An. funestus*,
288 exhibited a preference for naturally occurring or man-made wells that receive spring-fed
289 water, river streams, and puddles, rather than swamps (Table 5). *Culex* spp. and *An.*
290 *arabiensis* larvae displayed comparable preferences, except for the latter, which did not
291 favor man-made wells. Furthermore, *An. funestus* larvae were more commonly found in
292 water bodies with slow ($p < 0.001$) flow rate compared to other mosquito species (Table 5).
293 The presence of *An. funestus* larvae was found to be independent of vegetation type or
294 quantity. However, *An. arabiensis* showed a preference for habitats with floating vegetation
295 (Table 5). Furthermore, habitats with vegetation were less likely to host *Culex* spp. larvae.
296 Moreover, according to the modeling results, there was a positive correlation between the
297 depth of aquatic habitats and the likelihood of encountering *An. funestus* larvae. *Culex* spp.
298 larvae were negatively associated with the habitats having scarce or moderate algae
299 quantity, but the same was not observed for other mosquito species (Table 5). The
300 occurrence of mosquitoes was unaffected by the shading over the habitat, water clarity (i.e.,
301 clear, colored, or polluted), or habitat permanency (Table 5).

302

303 Acidity (pH), temperature (°C), electrical conductivity ($\mu\text{S}/\text{cm}$), total dissolved solids (ppm)
304 and dissolved oxygen (ppm) were measured in a total of 178 aquatic habitats in Igumbiro
305 village. Temperature, pH, total dissolved solids, and electrical conductivity were found to
306 have an impact on the occurrence of *Culex* spp. larvae. In contrast, only electrical
307 conductivity affected the presence of *An. arabiensis*, while pH specifically influenced the
308 presence of *An. funestus* (Table 6).

309

310

311
312Table 4: Characteristics of aquatic habitats of *Anopheles funestus* and other mosquito species

Parameter		No. habitats observed (%)	Habitats without larvae	Habitats with <i>An. funestus</i> larvae (%)	Habitats with other <i>Anopheles</i> larvae n(%)	Habitats with <i>Culicine</i> sp. larvae n(%)
Water movement	Stagnant	370 (49.7)	7 (1.9)	167 (45.1)	110 (29.7)	123 (33.2)
	Slow	354 (47.6)	11 (3.1)	164 (46.3)	90 (25.4)	147 (41.5)
	Fast	20 (2.7)	1 (5)	11 (55)	1 (5)	7 (35)
Water type	Permanent	525 (70)	10 (1.9)	279 (53.1)	150 (28.6)	230 (43.8)
	Temporary	219 (29.4)	9 (4.1)	63 (28.8)	51 (23.3)	47 (21.5)
Water colour	Clear	520 (69.9)	16 (3.1)	224 (43.1)	136 (26.2)	196 (37.7)
	Coloured	161 (21.6)	3 (1.8)	89 (55.3)	46 (28.6)	57 (35.4)
	Polluted	63 (8.5)	0	29 (46)	19 (30.2)	24 (38.1)
Habitat type	Swamp	168 (24.6)	2 (1.2)	97 (57.7)	41 (24.4)	76 (45.2)
	Stream	390 (56.2)	12 (3.1)	185 (47.4)	91 (23.3)	157 (40.3)
	Rice field	10 (1.6)	0	4 (40)	4 (40)	7 (70)
	Natural well	43 (3.7)	5 (11.6)	16 (37.2)	10 (23.3)	17 (39.5)
	Man-made well	32 (2.3)	0	7 (21.8)	7 (21.8)	3 (9.4)
	Puddle	101 (11.6)	0	33 (32.7)	48 (47.5)	17 (16.8)
Shade over habitat	None	425 (56.2)	10 (2.4)	196 (46.1)	127 (29.9)	145 (34.1)
	Partial	236 (31.9)	9 (3.8)	102 (43.2)	55 (23.3)	95 (40.3)
	Heavy	83 (11.8)	0	44 (53)	19 (22.9)	37 (44.6)
Vegetation quantity	None	257 (34.5)	12 (4.7)	114 (44.4)	57 (22.2)	96 (37.4)
	Scarce	68 (9.1)	0	34 (50)	15 (22)	27 (39.7)
	Moderate	217 (29.2)	6 (2.7)	103 (47.5)	57 (26.3)	78 (35.9)
	Abundant	202 (27.2)	1 (0.5)	91 (45.1)	72 (35.6)	76 (37.6)
Vegetation type	None	121 (16.3)	0	74 (61.2)	33 (27.3)	69 (57)
	Submerged	46 (6.2)	0	23 (50)	25 (54.3)	8 (17.4)
	Floating	99 (13.3)	0	70 (70.7)	34 (34.3)	53 (53.5)
	Emergent	478 (64.2)	19 (4)	175 (36.6)	109 (22.8)	147 (30.8)
Algae quantity	None	516 (69.4)	17 (3.3)	197 (26.5)	126 (16.9)	171 (23)
	Scarce	48 (6.5)	0	34 (70.8)	20 (41.7)	17 (35.4)
	Moderate	129 (17.3)	0	85 (65.9)	40 (31)	66 (51.2)
	Abundant	51 (6.9)	2 (3.9)	26 (51)	15 (29.4)	23 (45.1)
Habitat depth	Less than 10 cm	361 (48.5)	15 (4.2)	196 (54.3)	72 (19.9)	178 (49.3)
	Between 10-50cm	355 (47.7)	4 (1.1)	138 (38.9)	117 (33)	96 (27)
	Greater than 50cm	28 (3.8)	0	8 (28.6)	12 (42.9)	3 (10.7)

313
314

**No. habitats observed represents total number of observations per parameter.

**Rice field represents habitats specifically found within non-irrigational rice fields.

Table 5: Results of multivariate regression analysis of habitat characteristics and mosquito larvae

Parameter		<i>Anopheles arabiensis</i>		<i>Anopheles funestus</i>		<i>Culex spp.</i>	
		OR (95% CIs)	p-value	OR (95% CIs)	p-value	OR (95% CIs)	p-value
Water movement	Stagnant	1	-	1	-	1	-
	Slow	2.21 (0.60-8.17)	0.23	37.83 (6.99- 204.81)	<0.001	0.59 (0.07-5.31)	0.64
	Fast	0.77 (0.12-4.99)	0.79	21.25 (2.82-160.02)	<0.05	0.61 (0.05-6.89)	0.69
Water type	Temporary	1	-	1	-	1	-
	Permanent	0.14 (0.01-1.87)	0.14	1.04 (0.14-7.91)	0.97	0.21 (0.01-3.73)	0.29
Water colour	Clear	1	-	1	-	1	-
	Coloured	1.44 (0.72-2.91)	0.30	1.97 (0.97-4.01)	0.06	0.88 (0.44-1.76)	0.72
	Polluted	0.71 (0.29-1.76)	0.46	1.78 (0.73-4.32)	0.20	1.20 (0.51-2.84)	0.68
Habitat type	Swamp	1	-	1	-	1	-
	Stream	0.35 (0.14-0.83)	0.02	6.89 (3.33-14.26)	<0.001	0.10 (0.04-0.27)	<0.001
	Rice field	1.17 (0.22-6.32)	0.85	9.49 (1.71-52.52)	<0.05	0.08 (0.01-0.76)	0.03
	Natural well	0.10 (0.03-0.36)	<0.001	4.26 (1.47-12.38)	<0.05	0.10 (0.03-0.38)	<0.001
	Man-made well	0.40 (0.11-1.44)	0.16	9.66 (3-31.04)	<0.001	0.09 (0.02-0.37)	<0.001
	Puddle	0.32 (0.11-0.91)	0.03	10.26 (4.02-26.16)	<0.001	0.07 (0.02-0.23)	<0.001
Shade over habitat	None	1	-	1	-	1	-
	Partial	0.62 (0.19-2.02)	0.43	1.72 (0.54-5.44)	0.36	1.01 (0.37-2.74)	0.98
	Heavy	0.48 (0.13-1.75)	0.27	1.78 (0.52-6.16)	0.36	0.57 (0.19-1.73)	0.32
Vegetation type	None	1	-	1	-	1	-
	Floating	2.65 (1.31-5.38)	<0.05	0.52 (0.24-1.11)	0.09	0.44 (0.21-0.89)	0.02
	Submerged	2.09 (0.95-4.61)	0.07	1.19 (0.53-2.69)	0.68	0.28 (0.11-0.75)	0.01
	Emergent	0.07 (0-1.09)	0.06	0.17 (0.02-1.50)	0.11	0.03 (0-0.68)	0.03
Vegetation quantity	None	1	-	1	-	1	-
	Scarce	0.27 (0.02-3.70)	0.33	0.35 (0.04-3)	0.34	0.08 (0-1.49)	0.09
	Moderate	0.17 (0.01-2.14)	0.17	0.49 (0.61-3.96)	0.50	0.11 (0-1.92)	0.13
	Abundant	0.14 (0.01-1.84)	0.14	0.31 (0.04-2.47)	0.27	0.07 (0-1.16)	0.06
Algae quantity	None	1	-	1	-	1	-
	Scarce	0.29 (0.06-1.32)	0.11	1.85 (0.56-6.10)	0.32	0.15 (0.04-0.60)	<0.05
	Moderate	0.76 (0.21-2.71)	0.67	2.32 (0.79-6.83)	0.13	0.18 (0.05-0.63)	<0.05
	Abundant	0.82 (0.21-3.23)	0.78	21.81 (0.56-5.89)	0.32	0.39 (0.10-1.497)	0.17
Habitat depth	Less than 10 cm	1	-	1	-	1	-
	Between 10-50 cm	2.87 (0.69-11.92)	0.15	116.34 (18.22-743.05)	<0.001	0.24 (0.03-2.24)	0.21
	Greater than 50cm	1.02 (0.16-6.57)	0.98	138.11 (18.48-1031.95)	<0.001	0.12 (0.01-1.45)	0.10

317 **Table 6:** Results of multivariate regression analysis of different physicochemical
 318 characteristics and their association with occurrence of mosquito larvae
 319

Physicochemical characteristic	<i>Anopheles arabiensis</i>		<i>Anopheles funestus</i>		<i>Culex</i> spp.	
	OR (95% CIs)	p-value	OR (95% CIs)	p-value	OR (95% CIs)	p-value
pH	0.89 (0.32-2.43)	0.25	10.01 (1.11-90.32)	0.04	0.22 (0.06-0.83)	<0.001
Temperature (°C)	0.80 (0.62-1.02)	0.08	0.74 (0.35-1.54)	0.42	3.08 (1.82-5.23)	0.03
Electrical conductivity (µS/cm)	1.02 (1-1.04)	0.02	0.98 (0.94-1.02)	0.24	1.13 (1.04-1.23)	<0.001
TDS (ppm)	1 (0.98-1.02)	0.94	1.01 (0.96-1.06)	0.67	0.80 (0.66-0.97)	<0.05
DO (ppm)	0.96 (0.86-1.08)	0.53	0.83 (0.41-1.68)	0.60	1 (0.75-1.34)	1

320
 321 **Effects of larviciding on the abundance of adult female mosquitoes and larvae**

322 In Igumbiro, there was a significant reduction of indoor densities for *An. funestus* ($p < 0.001$)
 323 and *Culex* spp. ($p < 0.001$) following larviciding. However, the densities of *An. arabiensis*
 324 significantly increased after larviciding ($p = 0.005$). In Sululu, no significant change was
 325 observed in the densities of *An. arabiensis* ($p = 0.268$) or *An. funestus* ($p = 0.119$), but there
 326 was a significant decline in densities of *Culex* spp. ($p = 0.035$) (Fig. 4).

327 For early instars larvae densities (Fig. 5), there was a significant reduction in the Igumbiro
 328 village for *An. arabiensis* ($p < 0.001$), *An. funestus* ($p < 0.001$) and *Culex* spp. ($p < 0.001$)
 329 but the densities remained statistically the same for *An. arabiensis* ($p = 0.404$) and *An.*
 330 *funestus* larvae ($p = 0.651$) in the Sululu village. The densities of *Culex* spp. at early instars
 331 were significantly increased after larviciding intervention ($p = 0.001$) in Sululu village (Fig. 5).
 332 At late instars, the densities of *An. arabiensis* ($p < 0.001$), *An. funestus* ($p < 0.001$) and
 333 *Culex* spp. ($p < 0.001$) were significantly reduced in Igumbiro village but not in Sululu, where
 334 only the *An. funestus* densities were marginally reduced ($p = 0.045$) after larviciding (Fig. 5).

335
 336 **Figure 4:** Effect of a single application of biolarvicides on abundance of adult *An.*
 337 *funestus*, *An. arabiensis* and *Culex* spp. inside houses in Sululu and Igumbiro
 338 villages, south-eastern Tanzania.

339
 340 **Figure 5:** Densities of early- and late-instar larvae of *An. funestus*, *An. arabiensis*
 341 and *Culex* spp. before and after a single application of biolarvicides in Sululu and
 342 Igumbiro villages, south-eastern Tanzania. A previous study [49] identified that *Culex*
 343 spp. mainly consisted of *Cx. pipiens pipiens* and *Cx. quinquefasciatus*.

344
 345 **Discussion**

346 Effective engagement of community members can ensure sustainability of public health
347 programmes [50,51], in part by creating a sense of ownership and responsibility [52,53]. A
348 recent study [31] emphasized the importance of such engagement to improve the larviciding
349 programs in rural Tanzania, following recent expansion of the practice beyond urban settings
350 [33]. This can be achieved through a combination of community sensitization meetings and
351 targeted training of community-based persons, as demonstrated in urban Dar es Salaam,
352 where community-owned resource persons previously enabled scale-up of larval source
353 management [17,54].

354

355 In rural Africa, where mosquito habitats can be more diverse and numerous, and where the
356 existing infrastructure may be inadequate for traditional LSM approaches, a detailed
357 understanding of the ecology of dominant malaria vectors is also necessary to optimize
358 resource use. For example, in the Kilombero valley, Tanzania, where just one of the many
359 vector species, *An. funestus*, now **accounts for** nine in every ten malaria infections [34,55], it
360 may be most appropriate to preferentially target habitats for this vector species. Previous
361 studies in the area have provided a baseline for such investigations by mapping the primary
362 characteristics of the vector species habitats [37] and also by highlighting the importance of
363 such knowledge for practical LSM practices [31]. This study tested the potential of training
364 community members to identify, characterize and target larval habitats of dominant malaria
365 vectors in rural south-eastern Tanzania. Biolarvicides were applied to aquatic habitats of *An.*
366 *funestus* mosquitoes in the two villages, and the immediate impact assessed by estimating
367 the larval densities and also adult densities of the mosquitoes in people's dwellings.

368

369 The current larviciding program in mainland Tanzania is implemented by community health
370 workers (i.e., secondary school graduates with a year of community health training) under
371 the supervision of ward health officers. Limited biolarvicide supply and inadequate funding
372 put severe constraints on implementation of the program, however. The present study has
373 demonstrated the effectiveness of involving lay members of the community in identifying and
374 characterizing aquatic habitats of the most competent malaria vector species and applying
375 the same biolarvicides as provided by the government. Involving community members in
376 larviciding is not uncommon in Tanzania [17,54], but this study demonstrates the role they
377 can play in a species-focused larviciding approach. The training of community members was
378 effective, and they identified habitats and environmental characteristics associated with the
379 occurrence of *An. funestus* larvae. The aquatic habitats identified by the trained community
380 members had similar characteristics as those previously identified by expert vector biologists
381 [37]. The number of *An. funestus* mosquitoes inside the houses in the villages was

382 significantly reduced after application of biolarvicides to the aquatic habitats identified by
383 community members. The study suggests that relying on the community to sustainably
384 implement the government-led larviciding program is possible, but recurring training would
385 be better to maximize the program's impact. Additionally, Tables 1 and 2 present the initial
386 awareness and perception of community members, which served as a basis for developing
387 the training modules. Although the subset of community members that underwent training
388 exhibited improved awareness and perception during and after the intervention, we did not
389 seek to quantify this improvement due to the project's tight timeline. The ability of trained
390 community members to carry out habitat identification and larviciding activities indicates that
391 capacity for larval control can be enhanced through a set of simplified yet engaging training
392 sessions.

393

394 However, it is important to note that the findings from this study may not be directly
395 generalizable to other rural settings or different ecological zones. The study was conducted
396 in two specific villages within the Kilombero Valley, which has unique environmental and
397 ecological characteristics. Factors such as mosquito species composition, habitat types, and
398 local community practices can vary significantly in different regions. Therefore, while the
399 training program and larviciding approach demonstrated effectiveness in this context, further
400 studies are necessary to evaluate the applicability and efficacy of similar interventions in
401 other rural areas with different ecological conditions.

402

403 In the two villages investigated, more than 300 aquatic habitats were surveyed, and it was
404 found that *An. funestus* larvae had a preference for habitats with slow or fast-moving waters,
405 such as streams. While this has been previously been linked to the higher levels of dissolved
406 oxygen and aeration in such waters [37,56], this current study did not find any significant
407 associations between dissolved oxygen levels and the presence of *An. funestus*. Similarly,
408 while previous studies have reported the preference of *An. funestus* for vegetated habitats
409 [42,57,58], no such association was observed here. Except for pH, the physicochemical
410 characteristics of aquatic habitats did not appear to have a significant association with the
411 occurrence of *An. funestus* larvae, which was also observed in other mosquito species,
412 particularly *Culex* spp. larvae. These differences may, in part, be due to the limited
413 geographical extent of the current study, which covered only two villages. It was also
414 observed that the main aquatic habitats of *An. funestus*, including streams and wells, were in
415 close proximity to agricultural activities, suggesting they may be constantly exposed to
416 agricultural pesticide wastes as previously observed in the area [59], potentially
417 exacerbating the challenge of insecticide resistance. Fortunately, the current larviciding

418 programs in Tanzania deploy biolarvicides (i.e., *Bacillus thuringiensis israelensis* and
419 *Lysinibacillus sphaericus*), which remain effective against pyrethroid-resistant malaria
420 vectors.

421

422 While the approach tested here was clearly effective, we did not investigate the optimal
423 timing of the larviciding. Instead, the single application was done in the dry season, when the
424 habitats were least numerous and least expansive. Also, a recent mathematical simulation
425 [60] suggested that the most effective timing for larviciding is during or at the beginning of
426 the rainy season but that was on assumption that the main vector species would be *An.*
427 *gambiae* s.s., which breed in temporary pools and whose populations peak during the wet
428 season. It however remains unclear what the optimal timing for larviciding would be in areas
429 dominated by *An. funestus*, which tends to occupy perennial habitats and therefore remains
430 important throughout the year. The results of this study may be useful for future modelling
431 exercises to assess such scenarios. Moreover, a distinct trend of aquatic habitat
432 recolonization was observed within one to two weeks after treatment, followed by a notable
433 reduction. This pattern suggests the residual effectiveness of *Bti*. Furthermore, it's important
434 to keep in mind that our results are based on a single larvicide application, which served
435 primarily to showcase the capability of trained community members in targeting disease-
436 transmitting mosquitoes. This also highlights the success of a simpler yet effective training
437 approach.

438

439 Tanzania has had great examples of cross-sectoral engagement for malaria prevention. In
440 addition to the supply of locally manufactured biolarvicides from the Tanzania biotechnology
441 industry, there have also been major investments in community sensitization and
442 engagement. For example, the Dar es Salaam Urban Malaria Control Programme (UMCP)
443 organized a highly effective community-based program of environmental management to
444 reduce densities of malaria vectors, which even included a degree of fundraising by the
445 communities [61]. More recently, the use of trained community owned resource persons to
446 deploy the larvicides and also monitor adult densities resulted in significant declines of
447 malaria in the city [17,18,54]. This current study demonstrate that such community-based
448 strategies can be expanded to rural settings such as the Kilombero Valley. This could
449 significantly reduce implementation costs, especially as community members are generally
450 willing to participate. We did not explicitly investigate the community's willingness to
451 participate without compensation in the current study. Nevertheless, previous studies have
452 indicated that such willingness is possible when there is a higher perceived level of safety
453 and acceptance of the product[19,62]. Already, the district-level malaria officials have been

454 conducting community sensitization programs to support larval source management [31].
455 However, it is evident that more efforts are needed given that significant proportions of the
456 rural community members (~60% in this study), remain unaware of the importance of
457 larviciding for malaria control.

458

459 Another important observation was that while *An. funestus* clearly prefer certain habitat
460 types, they do often cohabit with other mosquito species. In this study the application of the
461 biolarvicides in the habitats of *An. funestus* also reduced indoor densities of adult *Culex* spp.
462 but not *An. arabiensis*. Also, though *An. arabiensis* preferred irrigational rice fields, their
463 larva densities were also reduced in habitats that they shared with *An. funestus*. It can be
464 assumed therefore that the current strategy of applying larvicides to all aquatic habitats
465 could be effective as well, especially if there is adequate resources and manpower.
466 However, the findings also indicate that it might be more cost-effective to preferentially target
467 the most competent malaria vector species.

468

469 This study also had some limitations. Firstly, the use of a 2 km transects underestimated the
470 potential effectiveness of larviciding because mosquitoes can fly much longer, sometime up
471 to or beyond 4 km [63]. This means that mosquitoes could have emerged from habitats
472 beyond the transect, potentially reducing the larviciding efficacy. Additionally, the study did
473 not gather post-assessment feedback from community members who participated in the
474 program, primarily due to constraints related to the project timeline and available resources.
475 Their views and insights could have provided valuable information on the sustainability of the
476 approach and identified areas for improvement. Furthermore, the study focused solely on
477 assessing the impact of larviciding on the abundance of the major malaria vector, without
478 evaluating its effect on malaria prevalence in the villages. A recent simulation study [60]
479 suggests a reduction in malaria prevalence one month after larvicide application. However,
480 due to time limitations, the larvicides were only applied once in this study. While this may
481 have been sufficient for demonstrating the role of community members, it is itself inadequate
482 for assessing efficacy of the intervention on its own. Lastly, the application of biolarvicides
483 was limited to larval habitats that tested positive for *An. funestus* during the survey,
484 potentially reducing coverage by not treating habitats that could have harbored *An. funestus*
485 larvae but tested negative at the time of the survey.

486

487 In addition to these limitations, potential biases must be considered. Selection bias may
488 have been introduced during the recruitment of volunteers for the entomological training.
489 The criteria for selecting participants—such as the ability to read and write properly,
490 involvement in household entomological surveillance, a minimum residency of two years,

491 and an age range of 18 to 50 years may have excluded certain community members who
492 could have contributed valuable perspectives. This selection process might have favored
493 more educated or engaged individuals, potentially skewing the results. Additionally,
494 response bias in the questionnaires cannot be ruled out. Since the questionnaires were
495 administered by interviewers, there is a possibility that respondents provided socially
496 desirable answers rather than their true perceptions and knowledge levels about disease-
497 transmitting mosquitoes and larviciding. This could lead to an overestimation of the
498 community's baseline awareness and perception. To mitigate these biases in future studies,
499 a more inclusive selection process and the use of anonymous self-administered
500 questionnaires could be considered.

501 **Conclusion**

502
503
504 Previous studies have shown the effectiveness of larviciding in urban areas of Tanzania,
505 following WHO guidelines. However, the National Malaria Strategic Plan of Tanzania has
506 extended larviciding to rural areas, despite not strictly adhering to these guidelines. **This**
507 **study highlights the potential of species-focused community-led larviciding as a sustainable**
508 **intervention for malaria control in rural settings. The observed reduction in mosquito**
509 **densities demonstrates that, with proper training and community engagement, local**
510 **communities can successfully implement and maintain vector control strategies. These**
511 **findings suggest that this approach can be effectively adapted to other resource-limited rural**
512 **settings. Furthermore, by reducing reliance on centralized programs, this model promotes**
513 **self-sufficiency and community ownership, which are critical for the long-term sustainability**
514 **of malaria control efforts. The results have important implications for policy makers and**
515 **public health officials, as community-led interventions could complement existing vector**
516 **control strategies, thereby enhancing the overall impact of malaria control programs.**

517 518 **Abbreviations**

519 *Bti*: *Bacillus thuringiensis israelensis*; *Ls*: *Lysinibacillus sphaericus*; NMSP: National Malaria
520 Strategic Plan; ITN: Insecticide-Treated bed Net; IRS: Indoor Residual Spraying; TDS: Total
521 Dissolved Solids; DO: Dissolved Oxygen; pH: Potential of Hydrogen; GPS: Global
522 Positioning System; GLM: Generalized Linear Model; GLMM: Generalized Linear Mixed
523 Model; REML: Restricted Maximum Likelihood; UMCP: Urban Malaria Control Programme.

524 525 **Declarations**

526 **Ethics approval and consent to participate**

527 Ethical approvals for this project were obtained from Ifakara Health Institute's Institutional
528 Review Board (Protocol ID: IHI/IRB/No: 29 - 2019) and the Medical Research Coordinating

529 Committee (MRCC) at the National Institute for Medical Research, in Tanzania (Protocol ID:
530 NIMR/HQ/R.8a/Vol.IX/3517). Written consents were sought from all participants of this
531 study, after they had understood the purpose and procedure of the discussions.

532

533 **Consent for publication**

534 Permission to publish this study was obtained from National Institute for Medical Research,
535 in Tanzania (No: NIMR/HQ/P. 12 VOL.XXXVI 37).

536

537 **Availability of data and materials**

538 The data will be made available by the corresponding author upon reasonable request.

539

540 **Competing interests**

541 The authors declare no competing interests.

542

543 **Funding**

544 This study was supported by the Wellcome Trust International Masters Fellowship in
545 Tropical Medicine and Hygiene (Grant No. 212633/Z/18/Z) awarded to SAM, Bill and
546 Melinda Gates Foundation (Grant Number: OPP1177156) awarded to FOO. All grants were
547 held at Ifakara Health Institute.

548

549 **Author contributions**

550 SAM, JL, FT and FOO were involved in study design. SAM, AJL, IHN and KK were involved
551 in data collection. SAM conducted data analysis. DK contributed in designing and validating
552 the model selection part of the data analysis. SAM and AJL wrote the manuscript. KU, SM
553 and GJ facilitated training of the community members and vector surveillance officers, and
554 collaboration with district's malaria focal persons. FOO, JL, FT, DK, KU, GJ, WM, IHN and
555 SM provided thorough review of the manuscript. All authors read and approved the final
556 manuscript.

557

558 **Acknowledgments**

559 We would like to thank the vector surveillance officers and community members of Ulanga
560 and Kilombero districts for their participation in this study. We would also like to extend our
561 sincere gratitude to Ms. Anna Nyoni, Ms. Elihaika Minja, Dr. Marceline Finda, Ms. Noelia
562 Pama, Ms. Alice Ombella, Mr. Prosper Kobero, Mr. Gerald Tamayamali, Mr. Betwel
563 Msugupakulya and Mr. Japhet Kihonda for their assistance on the execution of the project.
564 We are also grateful to Mr. Augustino Mwambaluka and Ms. Rukia Njalambaha for their
565 transport and administrative supports respectively. Our sincere gratitude goes to Mr.

566 Nicholaus Banzi and Eng. Alejandro Gonzalez both from Tanzania Biotech Products Limited
567 for facilitating training of the community members and vector surveillance officers. Also, we
568 sincerely appreciate the initial inputs on the draft manuscript by Ms. Prisca Kweyamba, Drs.
569 Holly Matthews and Florian Noulin.

570

571 **Reference:**

572

573 [1] Bhatt S, Weiss DJ, Cameron E, Bisanzio D, Mappin B, Dalrymple U, et al. The effect of
574 malaria control on *Plasmodium falciparum* in Africa between 2000 and 2015. *Nature*. 526
575 (2015) 207–211. doi: 10.1038/nature15535.

576

577 [2] WHO. World malaria report 2023 [Internet]. 2023. Available

578 from: <https://www.who.int/publications/i/item/9789240086173>

579

580 [3] The malERA Consultative Group on Health Systems, A research agenda for malaria
581 eradication: Health systems and operational research, *PLoS Med*. 8 (2011) Preprint
582 at [doi:10.1371/journal.pmed.1000397](https://doi.org/10.1371/journal.pmed.1000397).

583

584 [4] Okumu F, Gyapong M, Casamitjana N, Castro MC, Itoe MA, Okonofua F, et al. What
585 Africa can do to accelerate and sustain progress against malaria. *PLOS Glob Public Health*.
586 2 (2022) e0000262. doi: 10.1371/journal.pgph.0000262.

587

588 [5] Hancock PA, Hendriks CJM, Tangena JA, Gibson H, Hemingway J, Coleman M, et al.
589 Mapping trends in insecticide resistance phenotypes in African malaria vectors. *PLoS Biol*.
590 18 (2020) e3000633. doi: 10.1371/journal.pbio.3000633.

591

592 [6] Moyes CL, Athinya DK, Seethaler T, Battle KE, Sinka MS, Hadi MP, et al. Evaluating
593 insecticide resistance across African districts to aid malaria control decisions. *Proc Natl Acad
594 Sci USA*. 117 (2020) 22042–22050. doi: 10.1073/pnas.2002604117.

595

596 [7] Russell TL, Govella NJ, Azizi S, Drakeley CJ, Kachur SP, Killeen GF. Increased
597 proportions of outdoor feeding among residual malaria vector populations following
598 increased use of insecticide-treated nets in rural Tanzania. *Malar J.* 10 (2011) 1–10. doi:
599 10.1186/1475-2875-10-80.

600 [8] Moiroux N, Gomez MB, Pennetier C, Elanga E, Djènontin A, Chandre F, et al. Changes in
601 *Anopheles funestus* biting behavior following universal coverage of long-lasting insecticidal
602 nets in Benin. *J Infect Dis.* 206 (2012) 1622–1629. doi: 10.1093/infdis/jis565.

603

604 [9] Sougoufara S, Mocote Diédhiou S, Doucouré S, Diagne N, Sembène PM, Harry M, et al.
605 Biting by *Anopheles funestus* in broad daylight after use of long-lasting insecticidal nets: a
606 new challenge to malaria elimination. *Malar J.* 13 (2014) 125. doi: 10.1186/1475-2875-13-
607 125.

608

609 [10] Finda MF, Moshi IR, Monroe A, Limwagu AJ, Nyoni AP, Swai JK, et al. Linking human
610 behaviours and malaria vector biting risk in south-eastern Tanzania. *PLoS One.* 14 (6)
611 (2019) e0217414. doi: 10.1371/journal.pone.0217414.

612

613 [11] Matowo NS, Moore J, Mapua S, Madumla EP, Moshi IR, Kaindoa EW, et al. Using a new
614 odour-baited device to explore options for luring and killing outdoor-biting malaria vectors: a
615 report on design and field evaluation of the Mosquito Landing Box. *Parasit Vectors.* 6 (2013)
616 82. doi: 10.1186/1756-3305-6-82.

617

618 [12] Monroe A, Asamoah O, Lam Y, Koenker H, Psychas P, Lynch M, et al. Outdoor-sleeping
619 and other night-time activities in northern Ghana: implications for residual transmission and
620 malaria prevention. *Malar J.* 14 (2015) 343. doi: 10.1186/s12936-015-0543-4.

621

622 [13] Adhikari B, Pell C, Cheah PY. Community engagement and ethical global health
623 research. *Global Bioethics* 31(1) (2020) 1-12. doi: 10.1080/11287462.2020.1751880.

624

625 [14] Asale A, Kussa D, Girma M, Mbogo C, Mutero CM. Community based integrated vector
626 management for malaria control: lessons from three years' experience (2016–2018) in Botor-
627 Tolay district, southwestern Ethiopia. BMC Public Health 19 (2019) 1-14. doi:
628 10.1186/s12889-019-6674-0.

629

630 [15] Von Seidlein L, Peto TJ, Landier J, Nguyen T-N, Tripura R, Phommasone K,
631 Pongvongsa T, et al. The impact of targeted malaria elimination with mass drug
632 administrations on falciparum malaria in Southeast Asia: a cluster randomised trial. PLoS
633 Med 16 (2) (2019) e1002745. doi: 10.1371/journal.pmed.1002745.

634

635 [16] Mutero CM, Okoyo C, Girma M, Mwangangi J, Kibe L, Ng'ang'a P, Kussa D, Diro G,
636 Affognon H, Mbogo CM. Evaluating the impact of larviciding with *Bti* and community
637 education and mobilization as supplementary integrated vector management interventions
638 for malaria control in Kenya and Ethiopia. Malar J 19 (2020) 1-17. doi: 10.1186/s12936-020-
639 03429-9.

640

641 [17] Maheu-Giroux M, Castro MC. Impact of community-based larviciding on the prevalence
642 of malaria infection in Dar es Salaam, Tanzania. PLoS One 8(8) (2013) e71638. doi:
643 10.1371/journal.pone.0071638.

644

645 [18] Chaki PP, Mlacha Y, Msellemu D, Muhili A, Malishee AD, Mtema ZJ, Kiware SS, et al.
646 An affordable, quality-assured community-based system for high-resolution entomological
647 surveillance of vector mosquitoes that reflects human malaria infection risk patterns. Malar J
648 11 (2012) 1-18. doi: 10.1186/1475-2875-11-172.

649

650 [19] Hakizimana E, Ingabire CM, Rulisa A, Kateera F, van den Borne B, Muvunyi CM, van
651 Vugt M, et al. Community-based control of malaria vectors using *Bacillus thuringiensis* var.

652 *Israelensis* (*Bti*) in Rwanda. Int J Environ Res Public Health 19(11) (2022) 6699. doi:
653 10.3390/ijerph19116699.

654 [20] Dambach P, Baernighausen T, Traoré I, Ouedraogo S, Sié A, Sauerborn R, Becker N,
655 Louis VR. Reduction of malaria vector mosquitoes in a large-scale intervention trial in rural
656 Burkina Faso using *Bti* based larval source management. Malar J 18 (2019) 1-9. doi:
657 10.1186/s12936-019-2882-4.

658

659 [21] García GA, Fuseini G, Mba Nlang JA, Olo Nsue Maye V, Rivas Bela N, Wofford RN,
660 Weppelmann TA, et al. Evaluation of a multi-season, community-based larval source
661 management program on Bioko Island, Equatorial Guinea. Front Trop Dis 3 (2022) 846955.
662 doi: 10.3389/fitd.2022.846955.

663

664 [22] Van den Berg H, van Vugt M, Kabaghe AN, Nkalapa M, Kaotcha R, Truwah Z, Malenga
665 T, et al. Community-based malaria control in southern Malawi: a description of experimental
666 interventions of community workshops, house improvement and larval source management.
667 Malar J 17 (2018) 1-12. doi: 10.1186/s12936-018-2585-7.

668

669 [23] Phiri MD, McCann RS, Kabaghe AN, van den Berg H, Malenga T, Gowelo S, Tizifa T, et
670 al. Cost of community-led larval source management and house improvement for malaria
671 control: a cost analysis within a cluster-randomized trial in a rural district in Malawi. Malar J
672 20 (2021) 1-17. doi: 10.1186/s12936-021-04041-1.

673

674 [24] Kramer K, Mandike R, Nathan R, Mohamed A, Lynch M, Brown N, Mnzava A, Rimisho
675 W, Lengeler C. Effectiveness and equity of the Tanzania National Voucher Scheme for
676 mosquito nets over 10 years of implementation. Malar J 16 (2017) 1-13. doi:
677 10.1186/s12936-017-2046-y.

678

679 [25] Renggli S, Mandike R, Kramer K, Patrick F, Brown NJ, McElroy PD, Rimisho W, et al.
680 Design, implementation and evaluation of a national campaign to deliver 18 million free long-
681 lasting insecticidal nets to uncovered sleeping spaces in Tanzania. *Malar J* 12 (2013) 1-16.
682 doi: 10.1186/1475-2875-12-119.

683

684 [26] Mashauri FM, Kinung'hi SM, Kaatano GM, Magesa SM, Kishamawe C, Mwangi JR,
685 Nnko SE, Malima RC, Mero CN, Mboera LEG. Impact of indoor residual spraying of lambda-
686 cyhalothrin on malaria prevalence and anemia in an epidemic-prone district of Muleba,
687 north-western Tanzania. *Am J Trop Med Hyg* 88, 841 (2013). doi: 10.4269/ajtmh.12-0667.

688

689 [27] Kakilla C, Manjurano A, Nelwin K, Martin J, Mashauri F, Kinung'hi SM, Lyimo E, et al.
690 Malaria vector species composition and entomological indices following indoor residual
691 spraying in regions bordering Lake Victoria, Tanzania. *Malar J* 19, 1–14 (2020). doi:
692 10.1186/s12936-020-03547-4.

693

694 [28] Ministry of Health and Social Welfare. Tanzania National Malaria Strategic Plan 2014-
695 2020. Available from: [https://www.out.ac.tz/wp-content/uploads/2019/10/Malaria-Strategic-
696 Plan-2015-2020-1.pdf](https://www.out.ac.tz/wp-content/uploads/2019/10/Malaria-Strategic-Plan-2015-2020-1.pdf).

697

698 [29] World Health Organization. Guidelines for Malaria Vector Control. Guidelines for Malaria
699 Vector Control. (2019). Available
700 from: <https://www.who.int/publications/i/item/9789241550499>.

701

702 [30] World Health Organization. Larval Source Management: A Supplementary Malaria
703 Vector Control Measure. (2013). Available
704 from: <https://apps.who.int/iris/handle/10665/85379>.

705

706 [31] Mapua SA, Finda MF, Nambunga IH, Msugupakulya BJ, Ukio K, Chaki PP, et al.
707 Addressing key gaps in implementation of mosquito larviciding to accelerate malaria vector
708 control in southern Tanzania: results of a stakeholder engagement process in local district
709 councils. *Malar J.* 20 (2021) 1–14. doi: 10.1186/s12936-021-03769-z.
710
711 [32] MAELEZO TV. Tanzania President visit biolarvicide plant at Kibaha district. (2017).
712 Available from: <https://www.youtube.com/watch?v=4CzJcsxmptw>.
713
714 [33] The United Republic of Tanzania Ministry of Health, C. D. G. E. and C. National Malaria
715 Strategic Plan 2021-2025: Transitioning to Malaria Elimination in Phases. National Malaria
716 Control Program. November 2020. Available from: [http://api-hidl.afya.go.tz/uploads/library-](http://api-hidl.afya.go.tz/uploads/library-documents/1641210939-jH9mKCtz.pdf)
717 [documents/1641210939-jH9mKCtz.pdf](http://api-hidl.afya.go.tz/uploads/library-documents/1641210939-jH9mKCtz.pdf).
718
719 [34] Kaindoa, E. W., Matowo, N. S., Ngowo, H. S., Mkandawile, G., Mmbando, A., Finda, M.,
720 & Okumu, F. O. Interventions that effectively target *Anopheles funestus* mosquitoes could
721 significantly improve control of persistent malaria transmission in south–eastern Tanzania.
722 *PLoS One* 12, no. 5 (2017): e0177807. doi:10.1371/journal.pone.0177807.
723
724 [35] Lwetoijera, D. W., Harris, C., Kiware, S. S., Dongus, S., Devine, G. J., McCall, P. J., &
725 Majambere, S. Increasing role of *Anopheles funestus* and *Anopheles arabiensis* in malaria
726 transmission in the Kilombero Valley, Tanzania. *Malaria Journal* 13 (2014): 1-10.
727 doi:10.1186/1475-2875-13-331.
728
729 [36] Pinda, P. G., Eichenberger, C., Ngowo, H. S., Msaky, D. S., Abbasi, S., Kihonda, J.,
730 Bwanaly, H., & Okumu, F. O. Comparative assessment of insecticide resistance phenotypes
731 in two major malaria vectors, *Anopheles funestus* and *Anopheles arabiensis* in south-
732 eastern Tanzania. *Malaria Journal* 19 (2020): 1-11. doi:10.1186/s12936-020-03705-1.
733

734 [37] Nambunga, I. H., Ngowo, H. S., Mapua, S. A., Hape, E. E., Msugupakulya, B. J., Msaky,
735 D. S., Mhumbira, N. T., et al. Aquatic habitats of the malaria vector *Anopheles funestus* in
736 rural south-eastern Tanzania. *Malaria Journal* 19 (2020): 1-11. doi:10.1186/s12936-020-
737 03704-2.

738

739 [38] Gebrekidan, B. H., Heckelei, T., & Rasch, S. Characterizing farmers and farming system
740 in Kilombero Valley Floodplain, Tanzania. *Sustainability* 12 (17) (2020): 7114.
741 doi:10.3390/su12177114.

742

743 [39] Msofe, N. K., Sheng, L., & Lyimo, J. Land use change trends and their driving forces in
744 the Kilombero Valley Floodplain, Southeastern Tanzania. *Sustainability* 11 (2) (2019): 505.
745 doi:10.3390/su11020505.

746

747 [40] Mboera, L. E., Kihonda, J. A. P. H. E. T., Braks, M. A., & Knols, B. G. Influence of
748 centers for disease control light trap position, relative to a human-baited bed net, on catches
749 of *Anopheles gambiae* and *Culex quinquefasciatus* in Tanzania. *Am J Trop Med Hyg* 59 (4)
750 (1998): 595–596. doi:10.4269/ajtmh.1998.59.595.

751

752 [41] Maia, M. F., Robinson, A., John, A., Mgando, J., Simfukwe, E., & Moore, S. J.
753 Comparison of the CDC Backpack aspirator and the Prokopack aspirator for sampling
754 indoor-and outdoor-resting mosquitoes in southern Tanzania. *Parasites Vectors* 4 (2011): 1–
755 10. doi:10.1186/1756-3305-4-1.

756

757 [42] Gillies, M. T. & De Meillon, B. The Anophelinae of Africa south of the Sahara (Ethiopian
758 zoogeographical region). (1968): 343 pp.

759

760 [43] Ngowo, H. S., Kaindoa, E. W., Matthiopoulos, J., Ferguson, H. M. & Okumu, F. O.
761 Variations in household microclimate affect outdoor-biting behaviour of malaria vectors.
762 Wellcome Open Res 2, (2017).

763 [44] Ngowo, H. S., E. E. Hape, Matthiopoulos, J., Ferguson, H. M. & Okumu, F. O. Fitness
764 characteristics of the malaria vector *Anopheles funestus* during an attempted laboratory
765 colonization. Malaria J 20, (2021) 1-13.

766

767 [45] Mapua, S. A., E. E. Hape, J. Kihonda, H. Bwanary, K. Kifungo, M. Kilalangongono, E. W.
768 Kaindoa, H. S. Ngowo & F. O. Okumu. Persistently high proportions of *Plasmodium*-
769 infected *Anopheles funestus* mosquitoes in two villages in the Kilombero valley, South-
770 Eastern Tanzania. Parasite Epidemiol Control 18 (2022) e00264.

771

772 [46] Harvard Humanitarian Initiative. KoBoToolbox. Available
773 from: <https://www.kobotoolbox.org>.

774

775 [47] R Development Core Team. R: A Language and Environment for Statistical Computing.
776 R Foundation for Statistical Computing. 1 (2019) 409. Preprint
777 at: <https://doi.org/10.1007/978-3-540-74686-7>.

778

779 [48] Bates, D., M. Mächler, B. Bolker & S. Walker. Fitting linear mixed-effects models using
780 lme4. arXiv preprint arXiv:1406.5823 (2014).

781

782 [49] Matowo, N. S., Abbasi, S., Munhenga, G., Tanner, M., Mapua, S. A., Oullo, D.,
783 Koekemoer, L. L. et al. Fine-scale spatial and temporal variations in insecticide resistance in
784 *Culex pipiens* complex mosquitoes in rural south-eastern Tanzania. Parasites & Vectors 12
785 (2019) 1-13.

786

787 [50] Holder, H. D., Gruenewald, P. J., Ponicki, W. R., Treno, A. J., Grube, J. W., Saltz, R. F.,
788 Voas, R. B. et al. Effect of community-based interventions on high-risk drinking and alcohol-
789 related injuries. *JAMA* 284, no. 18 (2000) 2341-2347.

790 [51] Das, J. K., Salam, R. A., Arshad, A., Maredia, H., & Bhutta, Z. A. Community based
791 interventions for the prevention and control of Non-Helminthic NTD. *Infect Dis Poverty* 3
792 (2014) 1-12.

793

794 [52] Ingabire CM, Hakizimana E, Rulisa A, Kateera F, Van Den Borne B, Muvunyi CM, et al.
795 Community-based biological control of malaria mosquitoes using *Bacillus thuringiensis* var.
796 *israelensis* (*Bti*) in Rwanda: community awareness, acceptance and participation. *Malar J.*
797 16 (2017) 1–13. doi: 10.1186/s12936-017-2046-y.

798

799 [53] Gubler DJ, Clark GG. Community involvement in the control of *Aedes aegypti*. *Acta*
800 *Trop.* 61 (1996) 169–179. doi: 10.1016/0001-706X(95)00124-F.

801

802 [54] Geissbühler Y, Kannady K, Chaki PP, Emidi B, Govella NJ, Mayagaya V, et al. Microbial
803 larvicide application by a large-scale, community-based program reduces malaria infection
804 prevalence in urban Dar es Salaam, Tanzania. *PLoS One.* 4 (2009) e5107. doi:
805 10.1371/journal.pone.0005107.

806 [45] Mapua SA, Hape EE, Kihonda J, Bwanary H, Kifungo K, Kilalangongono M, et al.
807 Persistently high proportions of *Plasmodium*-infected *Anopheles funestus* mosquitoes in two
808 villages in the Kilombero valley, South-Eastern Tanzania. *Parasite Epidemiol Control.* 18
809 (2022) e00264. doi: 10.1016/j.parepi.2022.e00264.

810

811 [56] Tchigossou G, Akoton R, Yessoufou A, Djegbe I, Zeukeng F, Atoyebi SM, et al. Water
812 source most suitable for rearing a sensitive malaria vector, *Anopheles funestus* in the
813 laboratory. *Wellcome Open Res.* 2 (2017). doi: 10.12688/wellcomeopenres.12942.2.

814

815 [57] Gimnig JE, Ombok M, Kamau L, Hawley WA. Characteristics of larval anopheline
816 (Diptera: Culicidae) habitats in Western Kenya. *J Med Entomol.* 38 (2001) 282–288. doi:
817 10.1603/0022-2585-38.2.282.

818

819 [58] Cohuet A, Simard F, Toto JC, Kengne P, Coetzee M, Fontenille D. Species identification
820 within the *Anopheles funestus* group of malaria vectors in Cameroon and evidence for a new
821 species. *Am J Trop Med Hyg.* 69 (2003) 200–205. doi: 10.4269/ajtmh.2003.69.200.

822

823 [59] Matowo NS, Tanner M, Munhenga G, Mapua SA, Finda M, Utzinger J, et al. Patterns of
824 pesticide usage in agriculture in rural Tanzania call for integrating agricultural and public
825 health practices in managing insecticide-resistance in malaria vectors. *Malar J.* 19 (2020) 1–
826 16. doi: 10.1186/s12936-020-03623-9.

827

828 [60] Runge M, Mapua SA, Nambunga I, Smith TA, Chitnis N, Okumu FO, et al. Evaluation of
829 different deployment strategies for larviciding to control malaria: a simulation study. *Malar J.*
830 20 (2021) 1–14. doi: 10.1186/s12936-021-03916-x.

831

832 [61] De Castro MC, Yamagata Y, Mtasiwa D, Tanner M, Utzinger J, Keiser J, et al. Integrated
833 urban malaria control: a case study in Dar es Salaam, Tanzania. In *The Intolerable Burden of*
834 *Malaria II: What's New, What's Needed: Supplement to Volume 71 (2) of the American*
835 *Journal of Tropical Medicine and Hygiene.* American Society of Tropical Medicine and
836 Hygiene, 2004. doi: 10.4269/ajtmh.2004.71.103.

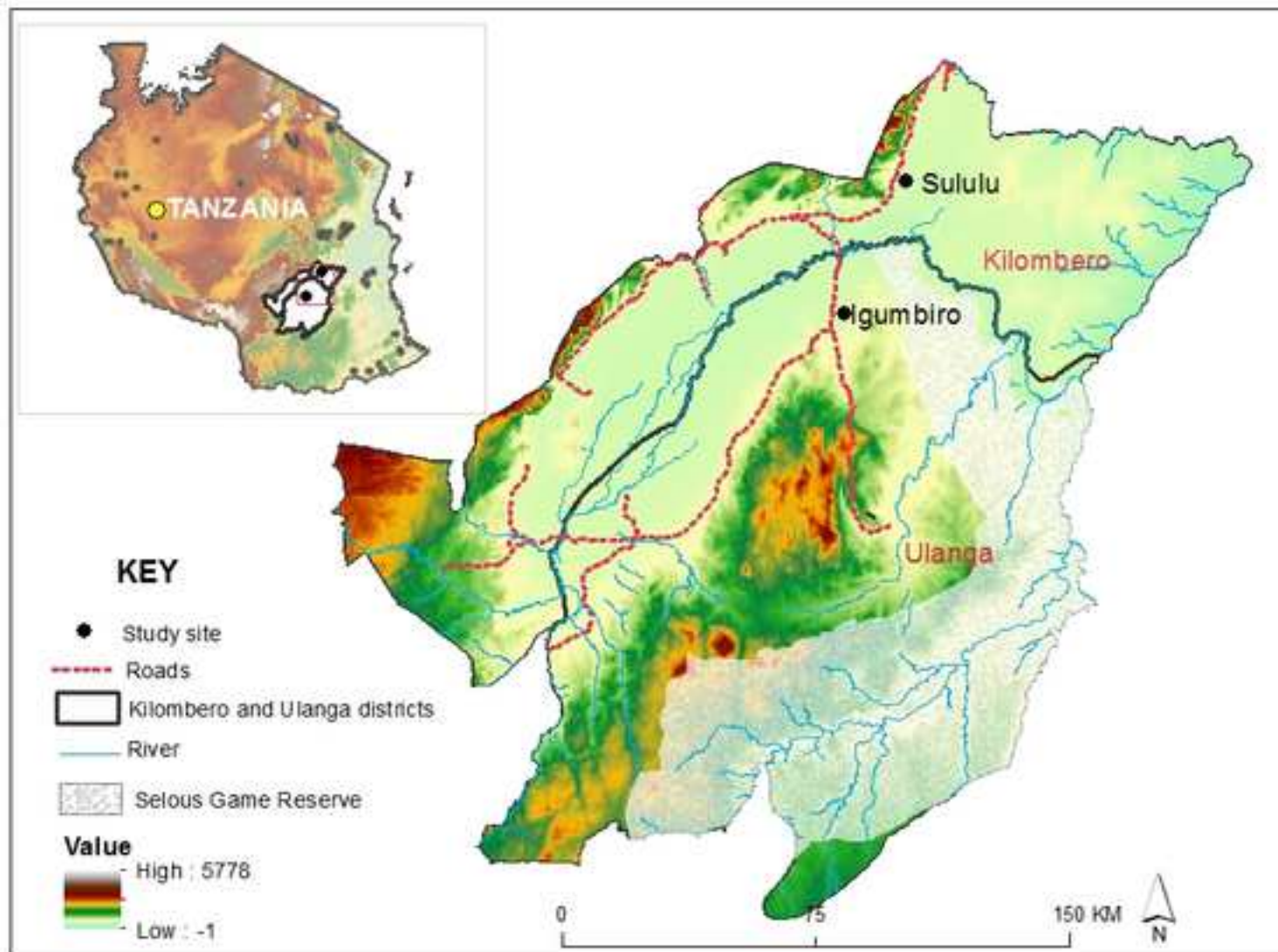
837

838 [62] Dambach P, Mendes Jorge M, Traoré I, Phalkey R, Sawadogo H, Zabré P, et al. A
839 qualitative study of community perception and acceptance of biological larviciding for malaria
840 mosquito control in rural Burkina Faso. *BMC Public Health.* 18 (2018) 1–11. doi:
841 10.1186/s12889-018-5841-8.

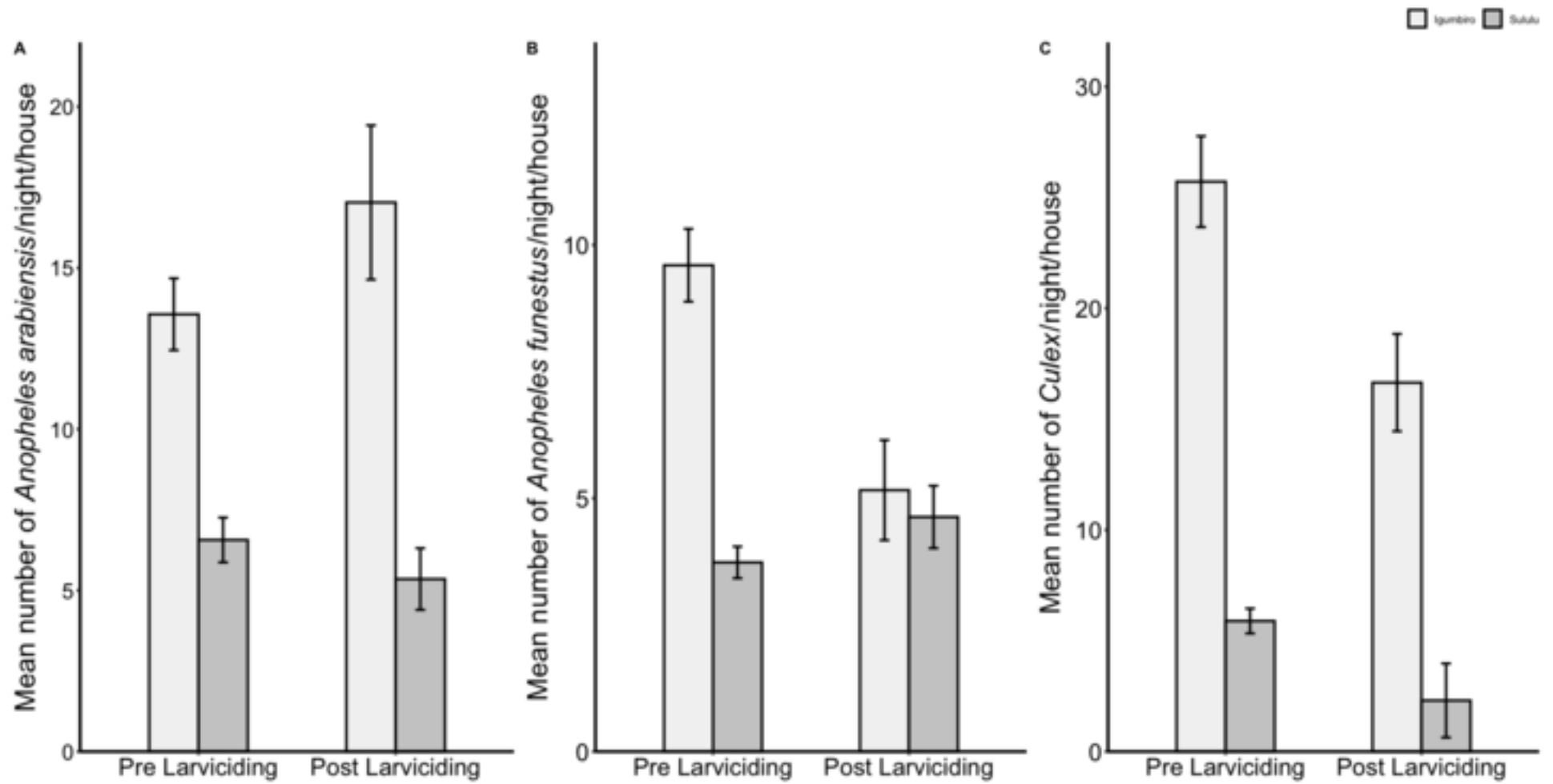
842

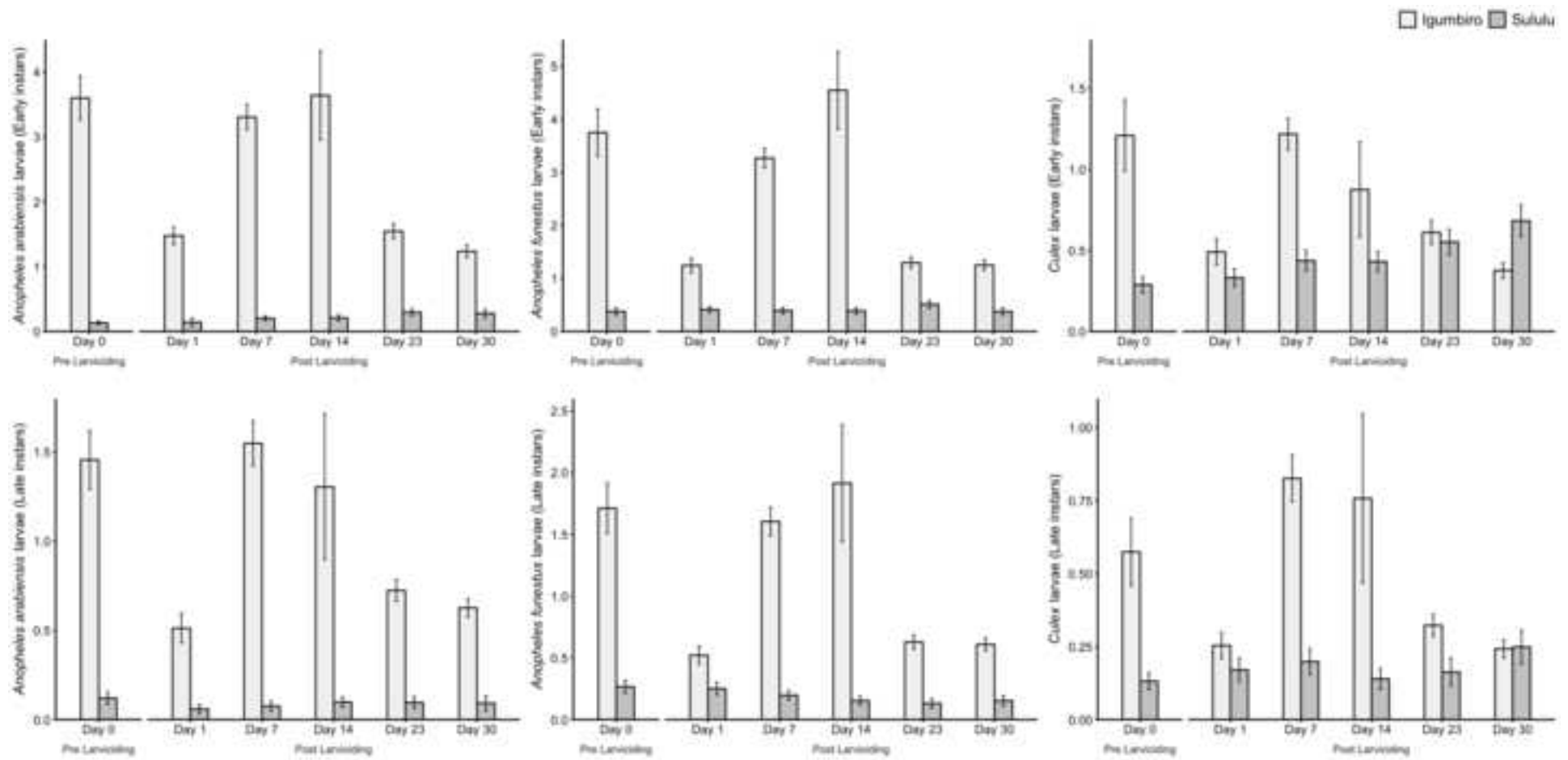
843 [63] Gillies MT. Studies on the dispersion and survival of *Anopheles gambiae* Giles in East
844 Africa, by means of marking and release experiments. Bull Entomol Res. 52 (1961) 99–127.
845 doi: 10.1017/S0007485300024797.











Parasite Epidemiology and Control
Radarweg 29
1043 NX Amsterdam
The Netherlands

August 3rd, 2023

Dear Editor

I hope this finds you well.

It is with great enthusiasm that I submit my manuscript titled **“Empowering Rural Communities for Effective Larval Source Management: A Small-Scale Field Evaluation of a Community-Led Larviciding Approach to Control Malaria in South-Eastern Tanzania”**. evaluate methods to involve community members in identifying, characterizing, and targeting the main aquatic habitats of the dominant malaria vector species using biolarvicide, and the impact of such an intervention on the density of the most relevant vector species in rural areas of Tanzania. The study focused on villages in the Kilombero valley, southeastern Tanzania, a meso-endemic community where *An. funestus* mediates >90% of malaria transmission events.

This study was funded in parts by Wellcome trust International Masters Fellowship in Tropical Medicine and Hygiene, and Bill and Melinda Gates Foundation. The aforementioned funders have no influence on the generated findings whatsoever. Data used to generate our findings can be accessed upon reasonable request to the corresponding author. In addition, Authors declare no conflict of interest.

I thank you in advance for your kind consideration and I look forward for your response.

Yours sincerely,



Salum A. Mapua

Corresponding author

+255 22 27261098; smapua@ihi.or.tz

PO Box 78373
Tel: 0 222 771 714
Fax: 0 222 771 714

PO Box 53
Tel: 0 232 625 164
Fax: 0 232 625 312

PO Box 74
Tel: 0 232 440 065
Fax: 0 232 440 064

Rufiji
PO Box 40 Ikwiriri
Tel: 0 232 010 007
Fax: 0 232 010 742

Mtwara
PO Box 1048
Tel: 0 232 333 487
Fax: 0 232 333 487

Kigoma
PO Box 1077
Tel: 0 282 803 655





Click here to access/download

e-Component

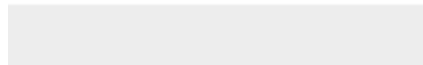
Binned Residual Plot_Anopheles funestus Model_Table
5.pdf



Click here to access/download

e-Component

Binned Residual Plot_Culex Model_Table 5.pdf





Click here to access/download

e-Component

Binned Residual Plot_Anopheles arabiensis
Model_Table 5.pdf

